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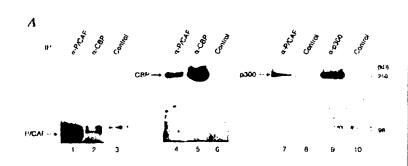
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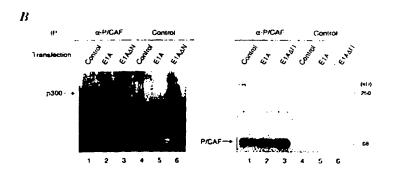
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(54) Title: P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

(57) Abstract

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein. The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided. Also provided are methods of screening for compounds that inhibit or stimulate the transcription modulating and histone acetyltransferase activity of P/CAF and p300/CBP.





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P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

BACKGROUND OF THE INVENTION

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Field of the Invention

The present invention provides a transcriptional co-factor, p300/CBP-associated factor (P/CAF), which modulates transcription through binding to the cellular transcription co-factors p300 and CBP and through acetylation of histones. Also provided are methods for screening for the presence of P/CAF and for substances which alter the transcription modulating effect and growth regulatory activity of P/CAF.

Background Art

Cellular proteins p300 and CBP are global transcriptional coactivators that are involved in the regulation of various DNA-binding transcriptional factors (Janknecht and Hunter, 1996). Recently, p300 was found to be very closely related to CBP, a factor that binds selectively to the protein kinase A-phosphorylated form of CREB (3-5). Cellular factors p300 and CBP exhibit strong amino acid sequence similarity and share the capacity to bind both CREB and E1A (6-8). Although neither p300 nor CBP by itself binds to DNA, each can be recruited to promoter elements via interaction with sequence-specific activators and functions to be a transcriptional adaptor. For simplicity, p300 and CBP will be termed p300/CBP in the context of discussing their shared functional properties.

p300/CBP is a large protein consisting of over 2,400 amino acids, known to interact with a variety of DNA-binding transcriptional factors including nuclear hormone receptors (13,57), CREB (3,4, 7), c-Jun/v-Jun (9,11), YY1 (10), c-Myb/v-Myb (12,58), Sap-1a (59), c-Fos (11) and MyoD (60). DNA-binding factors recruit p300/CBP not only by direct but also indirect interactions through cofactors, for example, nuclear 30 hormone receptors recruit p300/CBP directly as well as through indirect interactions, via SRC-1, which stimulates transcription by binding to various nuclear hormone receptors (13,61).



The transforming proteins encoded by adenovirus and several other small DNA tumor viruses disturb host cell growth control by interacting with cellular factors that normally function to repress cell proliferation. One of the most intensively studied of these viral proteins, the product of the adenovirus E1A gene, is itself sufficient for transformation (1). E1A transforming activity resides in two distinct domains, the targets of which include p300/CBP and products of the retinoblastoma (RB) susceptibility gene family (1,2). Interactions of E1A with p300/CBP and RB are thought to influence functionally distinct growth regulatory pathways, allowing the two domains to contribute additively to transformation (1).

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The paradigm for how E1A and functionally related viral proteins perturb cell growth regulation derives in large part from studies on their interactions with RB (1,2). The molecular function of E1A is based on its capacity to interfere with cellular protein-protein interactions. Since both E1A and various cellular targets bind to a site in RB termed the pocket domain (2), E1A can competitively disrupt the complex formation between RB and its cellular targets.

The second cellular factor implicated in E1A-dependent transformation, p300, is believed to inhibit G0/G1 exit, to activate certain enhancers, and to stimulate differentiation (1,2). E1A inhibits the p300/CBP-mediated transcriptional activation of many promoters (14). In one case that has been examined, the complex of p300 and YY1, E1A inhibits transcription without disrupting the complex (10).

The present invention provides a cellular protein designated P/CAF which binds to p300/CBP and plays an important role in both transcription and cell cycle regulation associated with a histone acetyltransferase activity. The present invention also provides a histone acetyltransferase activity in the p300/CBP cellular protein, thus providing targets for modulating transcription and cell cycle regulation in cells.

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SUMMARY OF THE INVENTION

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein.

The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided.

In addition, also provided is a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF and/or histone acetyltransferase activity, comprising contacting the substance with a system in which histone acetylation by P/CAF can be determined; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

Furthermore, the present invention provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF comprising contacting the substance with a system in which the p300 binding of P/CAF can be determined; determining the amount of p300 binding of P/CAF in the presence of the substance; and comparing the amount of p300 binding of P/CAF in the presence of the substance with the amount of p300 binding of P/CAF in the absence of the substance, a decreased amount of p300 binding of P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

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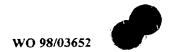


Also provided is a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:3 under conditions whereby a P/CAF/p300 complex can be formed, and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample.

The present invention additionally provides a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/antibody complex can be formed, and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample.

Also provided herein is an assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of P/CAF, comprising: contacting the substance with a system in which histone acetylation by P/CAF can be determined; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.

The present invention further provides an assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising: contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined; determining the amount of P/CAF binding of p300/CBP in the presence of the substance, and comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP



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In addition, an assay is provided for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP, comprising: contacting the substance with a system in which histone acetylation by p300/CBP can be determined, determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP.

Furthermore, the present invention provides an assay for screening substances for the ability to inhibit binding of a DNA-binding transcription factor to p300/CBP comprising: contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined; determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.

A method is also provided for inhibiting the transcription modulating activity of

P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier

Also provided in the present invention is a method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

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Furthermore, the present invention provides a method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

Finally, the present invention additionally provides a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

BRIEF DESCRIPTION OF THE FIGURES

Figs. 1A-B. Fig 1A: P/CAF-p300/CBP interaction *in vivo*. Cell extract was immunoprecipitated with rabbit anti-P/CAF (lanes 1, 4, and 7), rabbit anti-CBP (lanes 2 and 5), and mouse anti-p300 (lane 9) antibodies. For controls, cell extract was precipitated with rabbit control IgG (lanes 3, 6, and 8) or mouse anti-HA monoclonal antibody (lane 10). The precipitates were analyzed by immunoblotting with anti-P/CAF (lanes 1-3), anti-CBP (lanes 4-6), and anti-p300 (lanes 7-10) antibodies. The positions of non-specific bands are indicated by asterisks. Fig. 1B: E1A inhibits the P/CAF-p300 interaction *in vivo*. Osteosarcoma cells were transfected with either control vector (lanes 1 and 4) or E1A- (lanes 2 and 5) or E1AΔN- (lanes 3 and 6) expression vectors. Extract from the transfected subpopulation was immunoprecipitated with anti-P/CAF (lanes 1-3) or control (lanes 4-6) IgG. The precipitates were analyzed by immunoblotting with anti-p300 and anti-P/CAF.

Figs. 2A-F. P/CAF and E1A mediate antagonistic effects on cell cycle progression. HeLa cells (ATCC accession number CCL 2) were transfected by electroporation with 7 μ g of P/CAF-expression plasmid and/or 3 μ g of the full-length or the N-terminally deleted ($\Delta 2$ -36) E1A 12S-expression plasmid as indicated in the figure. These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into



pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1 μ g of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11 μ g. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 hours and subsequently labeled in medium containing 10 μ M bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32). Histograms show percentages of cells in G1 and S phases. Abscissa values represent fluorescence intensity of bound anti-BrdU antibodies in log scale.

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Fig. 3. Histone acetyltransferase activity of P/CAF. Activity of hGCN5 (lanes 1 and 4) and P/CAF (lanes 2 and 5) that acetylates free histones (lanes 1-3) or histones in the nucleosome core particle (35) (lanes 4-6) was measured as described (36). Each reaction contains 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. Note that the histone octamer dissociates into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE. The bands corresponding to acetylated histones H3 and H4 are indicated by arrows.

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DETAILED DESCRIPTION OF THE INVENTION

As used in the specification and in the claims, "a" can mean one or more, depending upon the context in which it is used.

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P/CAF protein and fragments

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones. The P/CAF protein can also bind to the amino acid region of SEQ ID NO 3

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(amino acid (aa) residues 1753 - 1966) of the cellular transcriptional factor, p300 (which has the complete amino acid sequence of SEQ ID NO:6 and the nucleotide sequence of SEQ ID NO:12), and the amino acid region of SEQ ID NO:6 (amino acid residues 1805 - 1854) of the cellular transcriptional factor, CBP (which has the complete amino acid sequence of SEQ ID NO:7 and the nucleotide sequence of SEQ ID NO:13). The P/CAF protein can be defined by any one or more of the typically used parameters. Examples of these parameters include, but are not limited to molecular weight (calculated or empirically determined), isoelectric focusing point, specific epitope(s), complete amino acid sequence, sequence of a specific region (e.g., N-terminus) of the amino acid sequence and the like.

For example, The P/CAF protein can consist of the amino acid sequence of SEQ ID NO:1 or the P/CAF protein can comprise the amino acid sequence of SEQ ID NO:2 which represents the carboxy terminal end of the P/CAF protein and contains the histone acetyltransferase activity, or the amino acid sequence of SEQ ID NO:4, which represents the amino terminal end of the P/CAF protein, containing the binding site for p300/CBP Because the amino-terminal region is specific for P/CAF it can be used to define and identify P/CAF.

As used herein, "purified" refers to a protein (polypeptide, peptide, etc.) that is sufficiently free of contaminants or cell components with which it normally occurs to distinguish it from the contaminants or other components of its natural environment. The purified protein need not be homogeneous, but must be sufficiently free of contaminants to be useful in a clinical or research setting, for example, in an assay for detecting antibodies to the protein. Greater levels of purity can be obtained using methods derived from well known protocols. Specific methods for purifying P/CAF proteins are known in the art.

As will be appreciated by those skilled in the art, the invention also includes those P/CAF polypeptides having slight variations in amino acid sequence which yield polypeptides equivalent to the P/CAF protein defined herein. Such variations may arise



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naturally as allelic variations (e.g., due to genetic polymorphism) or may be produced by human intervention (e.g., by mutagenesis of cloned DNA sequences), such as induced point, deletion, insertion and substitution mutants. Minor changes in amino acid sequence are generally preferred, such as conservative amino acid replacements, small internal deletions or insertions, and additions or deletions at the ends of the molecules. Substitutions may be designed based on, for example, the model of Dayhoff, et al. (37). These modifications can result in changes in the amino acid sequence, provide silent mutations, modify a restriction site, or provide other specific mutations.

Modifications to any of the P/CAF proteins or fragments can be made, while preserving the specificity and activity (function) of the native protein or fragment thereof. As used herein, "native" describes a protein that occurs in nature. The modifications contemplated herein can be conservative amino acid substitutions, for example, the substitution of a basic amino acid for a different basic amino acid. Modifications can also include creation of fusion proteins with epitope tags or known recombinant proteins or genes encoding them created by subcloning into commercial or non-commercial vectors (e.g., polyhistidine tags, flag tags, myc tag, glutathione-Stransferase [GST] fusion protein, xylE fusion reporter construct). Furthermore, the modifications can be such as do not affect the function of the protein or the way the protein accomplishes that function (e.g., its secondary structure or the ultimate result of the protein's activity). These products are equivalent to the P/CAF protein. The means for determining the function, way and result parameters are well known.

Having provided an example of a purified P/CAF protein, the invention also enables the purification of P/CAF homologs from other species and allelic variants from individuals within a species. For example, an antibody raised against the exemplary human P/CAF protein can be used routinely to screen preparations from different humans for allelic variants of the P/CAF protein that react with the P/CAF protein-specific antibody. Similarly, an antibody raised against an epitope, for example, from a conserved amino acid region of the human P/CAF protein can be used to routinely screen for homologs of the P/CAF protein in other species. A P/CAF protein can be

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routinely identified in and obtained from other species and from individuals within a species using the methods taught herein and others known in the art. For example, given the present sequence, the DNA encoding a conserved amino acid sequence can be used to probe genomic DNA or DNA libraries of an organism to predictably obtain the P/CAF gene for that organism. The gene can then be cloned and expressed as the P/CAF protein and purified according to any of a number of routine, predictable methods. An example of the routine protein purification methods available in the art can be found in Pei et al. (38).

A purified polypeptide fragment of the P/CAF protein is also provided. The term "fragment" as used herein regarding a P/CAF protein, means a molecule of at least five contiguous amino acids of P/CAF protein that has at least one function shared by P/CAF protein or a region thereof. These functions can include antigenicity, binding capacity, acetyltransferase activity and structural roles, among others. The P/CAF fragment can be specific for a recited source. As used herein to describe an amino acid sequence (protein, polypeptide, peptide, etc.), "specific" means that the amino acid sequence is not found identically in any other source. The determination of specificity is made routine by the availability of computerized amino acid sequence databases and sequence comparison programs, wherein an amino acid sequence of almost any length can be quickly and reliably checked for the existence of identical sequences. If an identical sequence is not found, the protein is "specific" for the recited source. For example, a P/CAF fragment can be species-specific (e.g., found in the P/CAF protein of humans, but not of other species).

A fragment of the P/CAF protein having histone acetyltransferase activity can consist of the amino acid sequence of SEQ ID NO.2. A fragment of the P/CAF protein which binds to the amino acid sequence of SEQ ID NO:3 on p300 and the amino acid sequence of SEQ ID NO:9 on CBP can consist of the amino acid sequence of SEQ ID NO 4. To the extent that these fragments are specific for P/CAF, they can be used to identify and define P/CAF.

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An antigenic fragment of P/CAF protein is provided. An antigenic fragment has an amino acid sequence of at least about five consecutive amino acids of a P/CAF protein amino acid sequence and binds an antibody or elicits an immune response in an animal. An antigenic fragment can be selected by applying the routine technique of epitope mapping to P/CAF protein to determine the regions of the proteins that contain epitopes reactive with antibodies or are capable of eliciting an immune response in an animal. Once the epitope is selected, an antigenic polypeptide containing the epitope can be synthesized directly, or produced recombinantly by cloning nucleic acids encoding the antigenic polypeptide in an expression system, according to standard methods.

Alternatively, an antigenic fragment of the antigen can be isolated from the whole P/CAF protein or a larger fragment of the P/CAF protein by chemical or mechanical disruption. Fragments can also be randomly chosen from a known P/CAF protein sequence and synthesized. The purified fragments thus obtained can be tested to determine their antigenicity and specificity by routine methods.

Nucleic Acids Encoding P/CAF Protein

An isolated nucleic acid that encodes a P/CAF protein is also provided. As used herein, the term "isolated" means a nucleic acid separated or substantially free from at least some of the other components of the naturally occurring organism, for example, the cell structural components commonly found associated with nucleic acids in a cellular environment and/or other nucleic acids. The isolation of nucleic acids can therefore be accomplished by techniques such as cell lysis followed by phenol plus chloroform extraction, followed by ethanol precipitation of the nucleic acids (39). It is not contemplated that the isolated nucleic acids are necessarily totally free of all non-nucleic acid components or all other nucleic acids, but that the isolated nucleic acids are isolated to a degree of purification to be useful in clinical, diagnostic, experimental, or other procedures such as, for example, gel electrophoresis, Southern, Northern or dot blot hybridization, or polymerase chain reaction (PCR).

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A skilled artisan in the field will readily appreciate that there are a multitude of procedures which may be used to isolate the nucleic acids prior to their use in other procedures. These include, but are not limited to, lysis of the cell followed by gel filtration or anion exchange chromatography, binding DNA to silica in the form of glass beads, filters or diatoms in the presence of high concentrations of chaotropic salts, or ethanol precipitation of the nucleic acids.

The nucleic acids of the present invention can include positive and negative strand RNA as well as DNA and can include genomic and subgenomic nucleic acids found in the naturally occurring organism. The nucleic acids contemplated by the present invention include double stranded and single stranded DNA of the genome, complementary positive stranded cRNA and mRNA, and complementary cDNA produced therefrom and any nucleic acid which can selectively or specifically hybridize to the isolated nucleic acids provided herein. Stringent conditions (further described below) are used to distinguish selectively or specifically hybridizing nucleic acids from non-selectively and non-specifically hybridizing nucleic acids.

An isolated nucleic acid that encodes a P/CAF protein can be species-specific (i.e., does not encode the P/CAF protein of other species and does not occur in other species). Examples of the nucleic acids contemplated herein include the nucleic acid of SEQ ID NO:10 as well as the nucleic acids that encode each of the P/CAF proteins or fragments thereof described herein P/CAF proteins and protein fragments can be routinely obtained as described herein and their structure (sequence) determined by routine means including the methods as used herein.

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P/CAF protein-encoding nucleic acids can be isolated from an organism in which they are normally found (e.g., humans), using any of the routine techniques. For example, a genomic DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest using one of the present P/CAF protein-encoding nucleic acids as a probe. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are

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commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers, which contain restriction sites, to the termini of the nucleic acid (See, for example, ref. 39).

P/CAF protein-encoding nucleic acids can also be synthesized. For example, a method of obtaining a DNA molecule encoding a specific P/CAF protein is to synthesize a recombinant DNA molecule which encodes the P/CAF protein. For example, nucleic acid synthesis procedures are routine in the art and oligonucleotides coding for a particular protein region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector.

Oligonucleotides complementary to or identical with the P/CAF proteinencoding nucleic acid sequence can be synthesized as primers for amplification reactions, such as PCR, or as probes to detect P/CAF protein encoding nucleic acids by various hybridization protocols (e.g., Northern blot; Southern blot; dot blot, colony screening, etc.).

Double-stranded molecules coding for relatively large proteins can readily be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein, followed by ligating these DNA molecules together. For example, Cunningham, et al. (40), have constructed a synthetic gene encoding the human growth hormone by first constructing overlapping and complementary synthetic oligonucleotides and ligating these fragments together. See also, Ferretti, et al. (41), wherein synthesis of a 1057 base pair synthetic bovine rhodopsin gene from synthetic oligonucleotides is disclosed.

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By constructing a P/CAF protein-encoding nucleic acid in this manner, one skilled in the art can readily obtain any particular P/CAF protein with modifications at any particular position or positions. See also, U.S. Patent No. 5,503,995 which describes an enzyme template reaction method of making synthetic genes. Techniques such as this are routine in the art and are well documented. DNA encoding the P/CAF protein or P/CAF protein fragments can then be expressed *in vivo* or *in vitro*.

The nucleic acid encoding the P/CAF protein can be any nucleic acid that functionally encodes the P/CAF protein. To functionally encode the protein (i.e., allow the nucleic acid to be expressed), the nucleic acid can include, but is not limited to, expression control sequences, such as an origin of replication, a promoter, regions upstream or downstream of the promoter, such as enhancers that may regulate the transcriptional activity of the promoter, appropriate restriction sites to facilitate cloning of inserts adjacent to the promoter, antibiotic resistance genes or other markers which can serve to select for cells containing the vector or the vector containing the insert, and necessary information processing sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites and transcription termination sequences as well as any other sequence which may facilitate the expression of the inserted nucleic acid.

Preferred expression control sequences are promoters derived from metallothionine genes, actin genes, immunoglobulin genes, CMV, SV40, adenovirus, bovine papilloma virus, etc. A nucleic acid encoding a P/CAF protein can readily be determined based upon the genetic code for the amino acid sequence of the P/CAF protein and many nucleic acid sequences will encode a P/CAF protein. Modifications in the nucleic acid sequence encoding the P/CAF protein are also contemplated. Modifications that can be useful are modifications to the sequences controlling expression of the P/CAF protein to make production of P/CAF protein inducible or repressible as controlled by the appropriate inducer or repressor. Such means are standard in the art (see, e.g., ref. 39). The nucleic acids can be generated by means standard in the art, such as by recombinant nucleic acid techniques, as exemplified in the examples herein, and by synthetic nucleic acid synthesis or in vitro enzymatic synthesis.



After a nucleic acid encoding a particular P/CAF protein of interest, or a region of that nucleic acid, is constructed, modified, or isolated, that nucleic acid can then be cloned into an appropriate vector, which can direct the *in vivo* or *in vitro* synthesis of that wild-type and/or modified P/CAF protein. The vector is contemplated to have the necessary functional elements that direct and regulate transcription of the inserted nucleic acid, as described above. The vector containing the P/CAF nucleic acid or nucleic acid fragment can be in a host (e.g., cell or transgenic animal) for expressing the nucleic acid. The P/CAF protein or fragment thereof can thus be produced in a host system containing the expression vector and its functional activity as described herein can be demonstrated according to methods well known in the art.

There are numerous *E. coli* (*Escherichia coli*) expression vectors known to one of ordinary skill in the art useful for the expression of proteins. Other microbial hosts suitable for use include bacilli, such as *Bacillus subtilis*, and other enterobacteria, such as *Salmonella*, *Serratia*, as well as various *Pseudomonas* species. These prokaryotic hosts can support expression vectors which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (Trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence and have ribosome binding site sequences, for example, for initiating and completing transcription and translation. If necessary, an amino terminal methionine can be provided by insertion of a Met codon 5' and in-frame with the gene sequence. Also, the carboxy-terminal extension of the protein can be removed using standard oligonucleotide mutagenesis procedures.

Additionally, yeast expression can be used. There are several advantages to yeast expression systems. First, evidence exists that proteins produced in yeast secretion systems exhibit correct disulfide pairing. Second, post-translational glycosylation is efficiently carried out by yeast secretory systems. The Saccharomyces cerevisiae prepro-alpha-factor leader region (encoded by the $MF\alpha-1$ gene) is routinely used to direct

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protein secretion from yeast (42). The leader region of pre-pro-alpha-factor contains a signal peptide and a pro-segment which includes a recognition sequence for a yeast protease encoded by the *KEX2* gene. This enzyme cleaves the precursor protein on the carboxyl side of a Lys-Arg dipeptide cleavage-signal sequence. The polypeptide coding sequence can be fused in-frame to the pre-pro-alpha-factor leader region. This construct is then put under the control of a strong transcription promoter, such as the alcohol dehydrogenase I promoter or a glycolytic promoter. The protein coding sequence is followed by a translation termination codon which is followed by transcription termination signals. Alternatively, the polypeptide encoding sequence of interest can be fused to a second protein coding sequence, such as Sj26 or β -galactosidase, used to facilitate purification of the resultant fusion protein by affinity chromatography. The insertion of protease cleavage sites to separate the components of the fusion protein is applicable to constructs used for expression in yeast.

Efficient post-translational glycosylation and expression of recombinant proteins can also be achieved in *Baculovirus* expression systems in insect cells.

Mammalian cells permit the expression of proteins in an environment that favors important post-translational modifications such as folding and cysteine pairing, addition of complex carbohydrate structures and secretion of active protein. Vectors useful for the expression of proteins in mammalian cells are characterized by insertion of the protein encoding sequence between a strong viral promoter and a polyadenylation signal. The vectors can contain genes conferring either gentamicin or methotrexate resistance for use as selectable markers. For example, the antigen and immunoreactive fragment coding sequence can be introduced into a Chinese hamster ovary (CHO) cell line using a methotrexate resistance-encoding vector. Presence of the vector RNA in transformed cells can be confirmed by Northern blot analysis and production of a cDNA or opposite strand RNA corresponding to the protein encoding sequence can be confirmed by Southern and Northern blot analysis, respectively. A number of other suitable host cell lines capable of secreting intact proteins have been developed in the art and include the CHO cell lines, HeLa cells, myeloma cell lines, Jurkat cells, and the like.



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Expression vectors for these cells can include expression control sequences, as described above. The vectors containing the nucleic acid sequences of interest can be transferred into the host cell by well-known methods, which vary depending on the type of cell host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cell hosts.

Alternative vectors for the expression of protein in mammalian cells, similar to those developed for the expression of human gamma-interferon, tissue plasminogen activator, clotting Factor VIII, hepatitis B virus surface antigen, protease Nexin I, and eosinophil major basic protein, can be employed. Further, the vector can include CMV promoter sequences and a polyadenylation signal available for expression of inserted nucleic acid in mammalian cells (such as COS7).

The nucleic acid sequences can be expressed in hosts after the sequences have been positioned to ensure the functioning of an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors can contain selection markers, e.g., tetracycline resistance or hygromycin resistance, to permit detection and/or selection of those cells transformed with the desired nucleic acid sequences (see, e.g., U.S. Patent 4,704,362).

The nucleic acids produced as described above can also be expressed in a host which is a non-human animal to create a transgenic animal, containing, in a germ or somatic cell, a nucleic acid comprising the coding sequence for all or a portion of the P/CAF protein, as well as all of the other regulatory elements required for expression of the P/CAF protein-encoding sequence. The animal will express the P/CAF gene or portion thereof to produce the P/CAF protein or protein fragment and such expression can be detected by determination of a particular phenotype unique to the transgenic animal expressing the transferred nucleic acid.

The nucleic acid can be the nucleic acid of SEQ ID NO:10, a nucleic acid having a nucleotide sequence which encodes the P/CAF protein, a nucleic acid having a nucleotide sequence which encodes the protein of SEQ ID NO:1, as well as the nucleic acids that encode the proteins comprising the fragments of SEQ ID NOS:2 and 4.

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The nucleic acids of the invention can contain substitutions or deletions which provide a particular phenotype of interest. For example, various deletions or base substitutions can be introduced into the nucleic acid encoding the P/CAF protein for the purpose of studying the effects of these particular deletions or substitutions on the transcription modulation activity of the P/CAF protein. These effects can be monitored by observation of such characteristics as growth and development of the animal, the ability to develop tumors, survival rates and the like. The gene construct introduced into the animal cells to produce the transgenic animal can contain any of the regulatory elements described above to modulate expression of the foreign genes. As used herein, the term "phenotype" includes morphology, biochemical profiles, changes in tumor formation and other parameters that are affected by the presence of the P/CAF protein.

The transgenic animals of the invention can also be used in a method for determining the effectiveness of administering a nucleic acid encoding a functional P/CAF protein to a subject in need of a functional P/CAF protein. First, a nucleic acid encoding a nonfunctional P/CAF protein can be introduced into the animal's cells and expressed to yield a characteristic phenotype. Then, using standard gene therapy techniques, a nucleic acid encoding a functional P/CAF protein can be introduced into the animal's cells and the effects on the animal's phenotypic characteristics can be determined.

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Having provided and taught how to obtain a nucleic acid that encodes a P/CAF protein, an isolated nucleic acid that encodes a fragment of P/CAF protein is also provided. The nucleic acid encoding the fragment can be obtained using any of the methods applicable to the nucleic acid encoding the entire P/CAF protein. The nucleic acid fragment can encode a species-specific P/CAF protein fragment (e.g., found in the



P/CAF protein of humans, but not in the P/CAF proteins of other species). Nucleic acids encoding species-specific fragments of P/CAF protein are themselves species-specific or allele-specific fragments of the P/CAF gene.

Examples of fragments of a nucleic acid encoding a fragment of the P/CAF protein can include the nucleic acid sequences which encode the amino acid sequences of the fragments of SEQ ID NOS:2 or 4. The same routine computer analyses used to select these examples of fragments can be routinely used to obtain others. Fragments of P/CAF-encoding nucleic acids can be primers for PCR or probes, which can be species-specific, gene-specific or allele-specific. P/CAF-encoding nucleic acid fragments can encode antigenic or immunogenic fragments of P/CAF protein that can be used in therapeutic assays or screening protocols. P/CAF gene fragments can encode fragments of P/CAF protein having histone acetylase activity and/or p300/CBP binding activity as described above, as well as other uses that may become apparent.

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An isolated nucleic acid of at least ten nucleotides that selectively hybridizes with the nucleic acid of SEQ ID NO:10 under selected conditions is provided. For example, the conditions can be PCR amplification conditions and the hybridizing nucleic acid can be a primer consisting of a specific fragment of the reference sequence or a nearly identical nucleic acid that hybridizes only to the exemplified P/CAF-encoding nucleic acid or allelic variants thereof.

The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF-encoding nucleic acid sequence of SEQ ID NO:10 under stringent conditions. The hybridizing nucleic acid can be a probe that hybridizes only to the exemplified P/CAF-encoding nucleic acid sequence. Thus, the hybridizing nucleic acid can be a naturally occurring species-specific allelic variant of the exemplified P/CAF gene. The hybridizing nucleic acid can also include insubstantial base substitutions that do not prevent hybridization under the stated stringent conditions or affect either the function of the encoded protein, the way the protein accomplishes that function (e.g., its

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secondary structure) or the ultimate result of the protein's activity. The means for determining these parameters are well known.

As used herein to describe nucleic acids, the term "selectively hybridizes" excludes the occasional randomly hybridizing nucleic acids as well as nucleic acids that encode other known homologs of the P/CAF protein. The selectively hybridizing nucleic acids of the invention can have at least 70%, 73%, 78%, 80%, 85%, 88%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% complementarity with the segment and strand of the sequence to which it hybridizes. This list is not intended to exclude percent complementarity values between these values. The nucleic acids can be at least 10, 15, 16, 17, 18, 20, 21, 23, 24, 25, 30, 35, 40, 50, 100, 150, 200, 300, 500, 550, 750, 900, 950, or 1000 nucleotides in length or any intervening length, depending on whether the nucleic acid is to be used as a primer, probe or for protein expression. The hybridizing nucleic acid can comprise a region of at least ten nucleotides (up to full length) that is completely complementary to a unique region of the nucleic acid to which it hybridizes.

The nucleic acid can be an alternative coding sequence for the P/CAF protein, or can be used as a probe or primer for detecting the presence of or obtaining the P/CAF protein. If used as primers, the invention provides compositions including at least two nucleic acids which selectively hybridize with different regions of the nucleic acid so as to amplify a desired region. Depending on the length of the probe or primer, it can range between 70% complementary bases and full complementarity and still hybridize under stringent conditions.

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For example, for the purpose of obtaining or determining the presence of a nucleic acid encoding the P/CAF protein, the degree of complementarity between the hybridizing nucleic acid (probe or primer) and the sequence to which it hybridizes (P/CAF DNA in a sample) should be at least enough to exclude hybridization with a nucleic acid from another species. The invention provides examples of these nucleic acids of P/CAF, so that the degree of complementarity required to distinguish selectively



hybridizing from nonselectively hybridizing nucleic acids under stringent conditions can be clearly determined for each nucleic acid. It should also be clear that the hybridizing nucleic acids of the invention will not hybridize with nucleic acids encoding unrelated proteins (hybridization is selective) under stringent conditions.

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"Stringent conditions" refers to the washing conditions used in a hybridization protocol. In general, the washing conditions should be a combination of temperature and salt concentration chosen so that the denaturation temperature is approximately 5-20°C below the calculated T_m of the nucleic acid hybrid under study. The temperature and salt conditions are readily determined empirically in preliminary experiments in which samples of reference DNA immobilized on filters are hybridized to the probe or protein encoding nucleic acid of interest and then washed under conditions of different stringencies. For example, the nucleic acid sequence of SEQ ID NO:10 was used as a specific radiolabeled probe for the detection of messenger RNA transcribed from the P/CAF gene by performing hybridizations under stringent conditions. The T_m of such an oligonucleotide can be estimated by allowing 2°C for each A or T nucleotide, and 4°C for each G or C. For example, an 18 nucleotide probe of 50% G+C would, therefore, have an approximate T_m of 54°C.

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The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF gene shown in the sequence set forth as SEQ ID NO:10 under stringent conditions. The invention further provides an isolated nucleic acid complementary to the nucleotide sequence set forth in SEQ ID NO:10.

25 Antibodies to the P/CAF protein

A purified antibody and an antiserum containing polyclonal antibodies that specifically bind the P/CAF protein or antigenic fragment are also provided. The term "bind" means the well understood antigen/antibody binding as well as other nonrandom association with an antigen. "Specifically bind" as used herein describes an antibody or other ligand that does not cross react substantially with any antigen other than the one specified, in this case, an antigen of the P/CAF protein. Antibodies can be made as

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described in Harlow and Lane (33). Briefly, purified P/CAF protein or an antigenic fragment thereof can be injected into an animal in an amount and in intervals sufficient to elicit a humoral immune response. Serum polyclonal antibodies can be purified directly, or spleen cells from the animal can be fused with an immortal cell line and screened for monoclonal antibody secretion, according to procedures well known in the art. Purified monospecific polyclonal antibodies that specifically bind the P/CAF antigen are also within the scope of the present invention. The antibodies of the present invention can bind the protein of claim 1, the protein of claim 2, the protein of claim 3 and/or the protein of claim 4, as well as any other proteins of the present invention.

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A ligand that specifically binds the antigen is also contemplated. The ligand can be a fragment of an antibody, such as, for example, an Fab fragment which retains P/CAF binding activity, or a smaller molecule designed to bind an epitope of the P/CAF antigen. The antibody or ligand can be bound to a substrate or labeled with a detectable moiety or both bound and labeled. The detectable moieties contemplated within the compositions of the present invention include those listed above in the description of the diagnostic methods, including fluorescent, enzymatic and radioactive markers.

The antibody can be bound to a solid support substrate or conjugated with a

detectable moiety or therapeutic compound or both bound and conjugated. Such
conjugation techniques are well known in the art. For example, conjugation of
fluorescent, radioactive or enzymatic moieties can be performed as described in the art
(33,43). The detectable moieties contemplated in the present invention can include
fluorescent, radioactive and enzymatic markers and the like. Therapeutic drugs
contemplated with the present invention can include cytotoxic moieties such as ricin A
chain, diphtheria toxin, pseudomonas exotoxin and other chemotherapeutic compounds

It is well understood by one of skill in the art that all of the above discussion regarding antibodies to P/CAF can also be applied with regard to production, characterization and use of antibodies which bind the p300/CBP protein or any of the DNA-binding transcription factors of this invention.



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Measuring the P/CAF protein in a sample

The present invention also provides a method for determining the presence and thus the amount of P/CAF protein in a biological sample. As used herein, a biological sample includes any tissue or cell which would contain the P/CAF protein. Examples of cells include tissues taken from surgical biopsies, isolated from a body fluid or prepared in an in vitro tissue culture environment.

One example of determining the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO.3 under conditions whereby a P/CAF/p300 complex can be formed, and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/p300 complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the p300 protein or by the detection of an antibody which binds p300 or the P/CAF protein, as taught in the Examples herein. Antibodies which bind p300 or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/p300 complexes by the detection of the binding of antibodies reactive with p300 or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as described below.

Alternatively, determination of the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:9 under conditions whereby a P/CAF/CBP complex can be formed, and determining the amount of the P/CAF/CBP complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/CBP complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the CBP protein or by the detection of an antibody which binds either CBP or the P/CAF protein, as taught in the Examples

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herein. Antibodies which bind CBP or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/CBP complexes by the detection of the binding of antibodies reactive with CBP or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as described below.

Another example of determining the amount of P/CAF in a biological sample comprises contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/ antibody complex can be formed and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample. Antibodies which bind P/CAF can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Determination of P/CAF/antibody complexes can be accomplished using various immunoassays as are available in the art, as described below.

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Immunoassays such as immunofluorescence assays, radioimmunoassays (RIA), immunoblotting and enzyme linked immunosorbent assays (ELISA) can be readily adapted for detection and measurement of P/CAF in a biological sample. Both polyclonal and monoclonal antibodies can be used in the assays. Available immunoassays are well known in the art and are extensively described in the patent scientific literature. See, for example, U.S. Patent Nos. 3,791,932, 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; and 4,098,876.

25 Screening assays for P/CAF

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur, determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the

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amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.

The present invention also provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF, comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and cell cycle progression suppressing activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the transcription modulating activity of P/CAF by interfering with the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.



Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 and P/CAF can occur; determining the amount of p300 binding to P/CAF in the presence of the substance; and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, a decreased amount of p300 binding to P/CAF in the presence of the substance, a decreased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO:3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

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Additionally provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur, determining the amount of CBP binding to P/CAF in the presence of the substance, and comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, a decreased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid sequence of SEQ ID NO.9 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance, determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase

activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further contemplates a bioassay for screening substances for the ability to stimulate the transcription modulating activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance; determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the transcription modulating activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples.

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Thus, the compound to be tested for the ability to stimulate the transcription modulating activity of P/CAF by increasing the histone acetyltransferase activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 to P/CAF can occur; determining the amount of p300 binding to P/CAF in the presence of the substance; and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, an increased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO:3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

Additionally provided in the present invention is a bioassay for screening substances for the ability to stimulate the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur; determining the amount of CBP binding to P/CAF in the presence of the substance, and comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, an increased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid sequence of SEQ ID NO.9 and P/CAF. Alternatively, the system can comprise a cell

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extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

Transcription modulating activity of P/CAF

The present invention contemplates a method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. For example, the substance can be identified according to the protocols provided herein as one that can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP or by inhibiting the histone acetyltransferase activity of P/CAF as well as by any other inhibitory mechanism as identified by the protocols provided herein. Inhibition of the transcription modulating activity of P/CAF in a subject is desirable, for example, to inhibit HIV TAT-mediated transcription and therefore, the method of the present invention can be used to treat HIV-infected subjects.

The substance can be in a pharmaceutically acceptable carrier. By "pharmaceutically acceptable" is meant a material that is not biologically or otherwise undesirable, i.e., the material may be administered to a subject, along with the substance, without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the pharmaceutical composition in which it is contained. The carrier would naturally be selected to minimize any degradation of the active ingredient and to minimize any adverse side effects in the subject.

The transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the P/CAF binding site or a substance which binds the P/CAF protein at the p300/CBP binding site, the ultimate result being that P/CAF and p300/CBP do not bind with one another and P/CAF cannot exert its transcription modulating and/or histone acetyltransferase effect. The substance can be a protein, such as an antibody which binds the P/CAF protein binding site at or near the p300/CBP

binding site, thereby preventing its binding or an antibody which binds the p300/CBP protein at or near the P/CAF binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on P/CAF or at the acetylation site on the histone, thereby preventing acetylation by P/CAF.

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The substance which binds p300/CBP, the P/CAF protein or the histone and has the net effect of inhibiting the transcription modulating effect and or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by mechanisms well known in the art.

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Alternatively, a nucleic acid encoding a protein which binds either to p300/CBP or the P/CAF protein and has the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors, as described below.

The substance which inhibits the transcription modulating effect and/or histone acetyltransferase activity of P/CAF can be an antisense RNA or an antisense DNA which binds the RNA or DNA of P/CAF, thereby preventing translation or transcription of the RNA or DNA encoding P/CAF and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF by inhibiting P/CAF production. The antisense RNA of the present invention can be generated from the nucleic acid of SEQ ID NO:14 (human) or SEQ ID NO:15 (mouse). Furthermore, the antisense DNA can be a phosphorothioate oligodeoxyribonucleotide having the nucleotide sequence of SEQ ID NO:16 (human) or of SEQ ID NO:17 (mouse). The mouse antisense RNA can be used to inhibit the activity of mouse P/CAF, having the nucleotide sequence of SEQ ID NO:18 and the amino acid sequence of SEQ ID NO:8 The present invention also contemplates an antisense nucleic acid sequence which can bind the DNA or RNA of any of the transcription factors or other proteins now known or later identified to bind P/CAF, thereby inhibiting expression of the gene products of

these proteins and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF.

The antisense nucleic acid can comprise a typical nucleic acid, but the antisense nucleic acid can also be a modified nucleic acid or a derivative of a nucleic acid such as a phosphorothioate analogue of a nucleic acid. The composition can comprise, for example, an antisense RNA that specifically binds an RNA encoded by the gene encoding the serum protein. Antisense RNAs can be synthesized and used by standard methods (62).

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Antisense RNA can inhibit gene expression by forming an RNA/RNA duplex between the antisense RNA and the RNA transcribed from the target gene. The precise mechanism by which this duplex formation decreases the production of the protein encoded by the endogenous gene probably involves binding of complementary regions of the normal sense mRNA and the antisense RNA strand with duplex formation in a manner that blocks RNA processing and translation. Alternative mechanisms include the formation of a triplex between the antisense RNA and duplex DNA or the formation of an DNA-RNA duplex with subsequent degradation of DNA-RNA hybrids by RNAse H. Furthermore, an antigene effect can result from certain DNA-based oligonucleotides via triple-helix formation between the oligomer and double-stranded DNA which results in the repression of gene transcription. Regardless of the specific molecular mechanism, the present invention results in inhibition of expression of the P/CAF gene by the introduced and replicated DNA resulting in inhibition of the transcription modulating and/or histone acetyltransferase activity of P/CAF, by a reduction in the expression of the nucleic acid to which the antisense nucleic acid is hybridized, and therefore a reduction of the gene product from the targeted gene.

The antisense nucleic acid may be obtained by any number of techniques known to one skilled in the art. One method of constructing an antisense nucleic acid is to synthesize a recombinant antisense DNA molecule. For example, oligonucleotide synthesis procedures are routine in the art and oligonucleotides coding for a particular

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protein or regulatory region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector. Double-stranded molecules coding for relatively large proteins or regulatory regions can be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein or regulatory region, followed by ligating these DNA molecules together. Once the appropriate DNA molecule is synthesized, this DNA can be cloned downstream of a promoter in an antisense orientation. Techniques such as this are routine in the art and are well documented.

An example of another method of obtaining an antisense nucleic acid is to isolate that nucleic acid from the organism in which it is found and clone it in an antisense orientation. For example, a DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector in an antisense orientation, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers which contain restriction sites to the termini of the nucleic acid. General methods are set forth in Sambrook et al. (39).

The DNA that is introduced into the cell is in an expression orientation that is antisense to a corresponding endogenous DNA or RNA of the cells. For example, where an endogenous DNA comprises a gene which encodes for a particular protein, the introduced DNA is in an expression orientation opposite the expression of the endogenous DNA, that is the DNA operatively linked to a promoter is in an antisense expression orientation relative to the corresponding endogenous gene. The introduced DNA may be homologous to the entire transcribed gene or homologous to only part of



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the transcribed gene. Alternatively, the sequence of the introduced DNA may be divergent to that of the endogenous DNA but only divergent to the extent that hybridization of the nucleic acids occurs, thereby preventing transcription. One skilled in the art can determine the maximum extent of this divergence by routine screening of antisense DNAs corresponding to an endogenous DNA of the cell. In this manner, one skilled in the art can readily determine which fragments, or alternatively the extent of homology of the fragments or the entire gene that is necessary to inhibit gene expression.

The antisense nucleic acids of the present invention can be made according to protocols standard in the art, as well as described in the Examples provided herein. The antisense nucleic acids can be administered to a subject according to the gene transduction protocols standard in the art, as described below.

The present invention also contemplates a method for stimulating the transcription modulating activity and/or histone acetyltransferase activity of P/CAF in a subject comprising administering to the subject a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a stimulatory affect on the transcription modulating and/or histone acetyltransferase activity of P/CAF. The substance can be one which has been identified, according to the protocols provided herein, to stimulate histone acetyltransferase activity in P/CAF or promote binding of P/CAF to p300/CBP. The stimulation of the transcription modulation activity and/or histone acetyltransferase activity of P/CAF in a subject is desirable, for example, to activate tumor suppressor p53 (which promotes apoptosis) or to activate the muscle differentiation factor, MyoD. Thus, the method of the present invention can be employed to treat cancer and to promote muscle differentiation in conditions where muscle differentiation is desired. The substance can be delivered to a cell in the subject by mechanisms well known in the art.

Further contemplated in the present invention is a method for promoting binding of P/CAF to p300/CBP in a subject, comprising administering to the subject a substance

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identified by the methods provided herein to promote binding of P/CAF to either p300 or CBP.

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Additionally, a nucleic acid encoding a protein which stimulates the transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

Also provided in the present invention is a method of inhibiting the cell cycle progression inducing effect of an oncoprotein which binds p300/CBP in a subject comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein, inducing expression of the nucleic acid in the cell to produce the P/CAF in an amount which will allow the P/CAF gene product to replace the oncoprotein bound to p300/CBP, whereby the replacement of the oncoprotein bound to p300/CBP by the P/CAF gene product inhibits the cell cycle progression inducing effect of the oncoprotein. The oncoprotein which binds p300/CBP in the cell can be the adenovirus E1A oncoprotein

A method for providing a functional P/CAF protein to a subject in need of the functional P/CAF protein is also provided, comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein and inducing expression of the nucleic acid to produce the functional P/CAF protein in the cell, thereby providing the functional P/CAF protein to the subject. The transduction of the vector nucleic acid into the subject's cells can be carried out according to standard gene therapy protocols well known in the art (see, for example, U.S. Patent No. 5,339,346).

Screening assays for p300/CBP

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance under conditions whereby histone acetylation by p300/CBP can occur,



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determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for acetyltransferase inhibiting ability.

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Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of a transcriptional factor to p300/CBP, comprising contacting a system in which the binding of a transcriptional factor to p300/CBP can be determined, with the substance under conditions whereby the binding of the transcriptional factor and p300/CBP can occur, determining the amount of transcriptional factor binding to p300/CBP in the presence of the substance; and comparing the amount of transcriptional factor binding to p300/CBP in the presence of the substance with the amount of transcriptional factor binding to p300/CBP in the absence of the substance, a decreased amount of transcriptional factor binding to p300/CBP in the presence of the substance indicating a substance that can inhibit the binding of a transcriptional factor to p300/CBP. The binding of a transcriptional factor to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising a transcriptional factor which binds p300/CBP and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both a transcriptional factor which binds p300/CBP and p300/CBP. The transcriptional factor which binds p300/CBP can be selected from, but is not limited to



the group consisting of nuclear hormone receptors, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YYI, Sap-1a, c-Fos, MyoD and SRC-1, as well as any other transcriptional factor now known or later identified to bind p300/CBP. The screening assay of the present invention can also be used to identify substances which inhibit the binding of p300/CBP to other components to which it is known to bind, for example, P/CAF, pp90_{RSK}, TFIIB, E1A, SV40 large T antigen, as well as any other substances now known or later identified to bind p300/CBP. Determination of the binding of a transcriptional factor or other substance to p300/CBP can be carried out as taught in the Examples herein as well as by protocols described in the literature.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance; determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, an increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the p300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for stimulating ability.

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of a component, which binds p300/CBP, to p300/CBP, comprising contacting a system in which the binding of the component to p300/CBP can



be determined, with the substance under conditions whereby the binding of the component to p300/CBP can occur; determining the amount of component binding to p300/CBP in the presence of the substance; and comparing the amount of component binding to p300/CBP in the presence of the substance with the amount of component binding to p300/CBP in the absence of the substance, an increased amount of component binding to p300/CBP in the presence of the substance indicating a substance that can stimulate the binding of the component to p300/CBP. The binding of the component to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising the component and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both the component and p300/CBP. The component which binds p300/CBP can be any of the transcriptional factors or other proteins which are known or are identified in the future to bind p300/CBP, as set forth above. Determination of the binding of the component to p300/CBP can be carried out as taught in the Examples provided herein and according 15 to protocols available in the literature.

Histone acetyltransferase activity of p300/CBP

A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject is provided in the present invention, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. The mechanism of the inhibitory action of the substance can be the inhibition of the binding of a DNA-binding transcription factor, such as, for example, a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD or SRC-1, to p300/CBP.

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The histone acetyltransferase activity of p300/CBP can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the transcription factor binding site or a substance which binds the transcription factor protein at the p300/CBP binding site, the ultimate result being that the transcription factor and p300/CBP do not bind with one another and p300/CBP cannot acetylate histones.

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The substance which binds either to the transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be identified according to the screening methods provided herein and delivered to a cell in the subject by mechanisms well known in the art. The substance can be a protein, such as an antibody which binds the p300/CBP protein binding site at or near the DNA-binding transcription factor binding site, thereby preventing its binding or an antibody which binds the DNA-binding transcription factor at or near the p300/CBP binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on p300/CBP (aa 1195-1673 on p300 or aa 1174-1850 on CBP) or at the acetylation site on the histone, thereby preventing acetylation by p300/CBP

Additionally, the substance can be a nucleic acid which can be expressed in the cell to produce a protein which inhibits the histone acetyltransferase activity of p300/CBP. For example, a nucleic acid encoding a protein which binds either to a transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors, as described below

The substance which inhibits the histone acetyltransferase activity of p300/CBP can be an antisense RNA or an antisense DNA which binds the RNA or DNA of p300/CBP thereby preventing translation or transcription of the RNA or DNA encoding p300/CBP and having the net effect of inhibiting the histone acetyltransferase activity of P/CAF by inhibiting p300/CBP production. The antisense RNA or DNA of the present invention can be produced and introduced into cells according to the same methods as set forth above for P/CAF antisense nucleic acids.

The present invention also contemplates a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject comprising administering to the



subject a histone acetyltransferase activity stimulating amount of a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a stimulatory affect on the histone acetyltransferase activity of p300/CBP. The substance can exert a stimulatory effect by promoting the binding of a DNA-binding transcription factor of the present invention to p300/CBP. The substance can be delivered to a cell in the subject by mechanisms well known in the art. A nucleic acid encoding a protein which stimulates the transcription modulating activity of p300/CBP can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

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Gene transduction

In the methods described above which include gene transduction into cells (i.e., addition of exogenous DNA into cells), the nucleic acids of the present invention can be in a vector for delivering the nucleic acids to the site for expression of the P/CAF protein. The vector can be one of the commercially available preparations, such as the pGM plasmid (Promega). Vector delivery can be by liposome, using commercially available liposome preparations or newly developed liposomes having the features of the present liposomes. Additionally, vector delivery can be via a viral system, including, but not limited to, retroviral, adenoviral and adeno-associated viral systems. Other delivery methods can be adopted and routinely tested according to the methods taught herein.

The modes of administration of the liposome will vary predictably according to the disease being treated and the tissue being targeted. For example, for treating cancer in either the lung or the liver, which are both sinks for liposomes, intravenous delivery is reasonable. For other localized cancers, as well as precancerous conditions, catheterization of an artery upstream from the target organ is a preferred mode of delivery, because it avoids significant clearance of the liposome by the lung and liver. For cancerous lesions at a number of other sites (e.g., skin cancer, localized dysplasias), topical delivery is expected to be effective and may be preferred, because of its convenience.

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Leukemias and other disorders involving dysregulated proliferation of certain isolatable cell populations may be more readily treated by ex vivo administration of the nucleic acid.

The liposomes may be administered topically, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally or the like, although intravenous or topical administration is typically preferred. The exact amount of the liposomes required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the severity of the disease being treated, the particular compound used, its mode of administration and the like. Thus, it is not possible to specify an exact amount. However, an appropriate amount may be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein.

Parenteral administration, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant level of dosage is maintained. See, e.g., U.S. Patent No. 3,610,795, which is incorporated by reference herein.

Topical administration can be by creams, gels, suppositories and the like Ex vivo (extracorporeal) delivery can be as typically used in other contexts.

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The present invention is more particularly described in the following examples which are intended as illustrative only since numerous modifications and variations therein will be apparent to those skilled in the art.

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EXAMPLES

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1. P/CAF studies.

5 Cloning and characterization of P/CAF protein.

In human cells, CBP binds to c-Jun in a phosphorylation-dependent manner in association with stimulation of transcription (9). In yeast, GCN4 is believed to be a c-Jun counterpart on the basis of similarities in DNA recognition (15) as well as the participation of both proteins in UV signaling pathways (16). Yeast genetic screening has led to the isolation of various cofactors for GCN4, including GCN5 (yGCN5), ADA2 (yADA2) and ADA3 (yADA3) (17-19). These factors are considered to function as a complex (or in a common pathway) based on genetic and protein-protein interaction studies (18-22). Finally, p300/CBP and yADA2 exhibit significant sequence similarity within a 50 amino acid region including a Zn²⁺ finger motif (3). Human counterparts to yGCN5, yADA2, or yADA3 that interact with p300/CBP to mediate transcriptional activation by c-Jun were searched for in various nucleotide sequence databases.

Comparison of the yGCN5 protein sequence with various databases (23) revealed significant similarities with the two randomly sequenced human cDNAs, ETS05039 (24) (P=4.0x10⁻¹⁵) and NIB2000-5R (P=6.5x10⁻⁹). Given that these cDNAs were truncated, human fetal liver and fetal brain cDNA libraries (Clontech) were screened with ETS05039 and NIB2000-5R, respectively and complete clones were isolated from the human fetal liver cDNA library. The complete sequences revealed that the ETS05039- and NIB2000-5R-derived clones are encoded by distinct genes but are highly related within the protein coding regions (68% identity at the DNA level; 75% identity and 86% similarity at the protein level). The former encodes an N-terminal region with no sequence similarity to any proteins in the databases besides the yGCN5-related C-terminal region, whereas the latter encodes only the yGCN5-related region. Given that p300/CBP-binding activity was observed in the former polypeptide as shown below, it was designated p300/CBP-associated factor (P/CAF), having the amino acid



sequence of SEQ ID NO:1 and the nucleotide sequence of SEQ ID NO:10 and the latter was named human GCN5 (hGCN5), having the amino acid sequence of SEQ ID NO:5 and the nucleotide sequence of SEQ ID NO:11.

Additionally, an RNA blot (Clontech) was hybridized with a random-primed probe made from the cDNA encoding P/CAF. RNA blotting indicated that transcripts detected by the P/CAF and hGCN5 cDNAs are ubiquitously expressed, but the former is most abundant in heart and skeletal muscle, whereas the latter is most abundant in pancreas and skeletal muscle.

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P/CAF-p300/CBP interaction in vitro

The P/CAF binding site was presumed to reside in the C terminal one third of CBP (residues 1,678-2,442) because it was observed that this region, when fused to a DNA binding domain, activates transcription (4) in a manner repressed by coexpression of 12S E1A. This region was divided into 6 overlapping fragments and each was expressed in *E. coli* as a glutathione-S-transferase (GST) fusion protein. GST-CBP fusions were incubated with recombinant P/CAF protein and, subsequently, purified using glutathione-Sepharose. Co-purified P/CAF was detected by immunoblotting analysis.

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To construct GST-fusions, various regions of CBP and p300 were amplified by PCR. A series of deletions of the CBP segment B was created by site-directed *in vitro* mutagenesis (30). These fragments were subcloned into pGEX-2T (Pharmacia). GST-fusions were expressed in *E. coli* and extracted with buffer B [20 mM Tris-HCl (pH 8.0), 5 mM MgCl₂, 10% glycerol, 1 mM AEBSF, 0.1% NP40, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 1 mM DTT] containing 0.1 M KCl for these experiments. GST-CBP-segment B was purified by glutathione-Sepharose and phenyl-Sepharose chromatographic steps, P/CAF, hGCN5, and E1A were expressed as FLAG-fusions in Sf9 cells via baculovirus vectors and affinity-purified with M2-agarose (ref. 30, Kodak-IBI). For interaction, a crude *E. coli* extract containing 20 pmol of GST-fusion was incubated with 40-60 pmol of P/CAF or E1A in a total volume of 50 μl of

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buffer B with 0.1 M KCl on ice for 10 min. Samples were further incubated with 10 µl (packed volume) of glutathione-Sepharose at 4°C for 30 min, washed four times with 200 µl of buffer B containing 0.1 M KCl, and eluted with 20 µl of buffer E [50 mM Tris-HCl (pH 8.0), 0.2 M KCl, 20 mM glutathione] for 60 min. Interacting proteins were detected by anti-FLAG immunoblotting or silver staining.

For p300 interactions, the segment spanning residues 1763-1966 (segment B') of p300, which is analogous to the CBP segment-B, was used. Twenty percent of the P/CAF and hGCN5 inputs and 100% of the E1A input were also analyzed. In the GST precipitation assays, almost identical amounts of the GST fusions were recovered in all samples. Interaction between P/CAF and CBP (segment B) was determined in the absence and in the presence of E1A. Control reactions with GST-CBP alone and without GST-CBP were also performed. Input proteins were analyzed.

Two CBP segments, A and B, interacted specifically with P/CAF. The stronger interaction was observed in the latter segment, which does not include the yADA2-like Zn²⁺ finger. Given that the CBP segment-B is well conserved in p300 (66% identity, 75% similarity), the binding of P/CAF to p300 *in vitro* was also analyzed. For this experiment, the p300 segment spanning residues 1763-1966, termed segment B', which is analogous to the CBP segment-B, was used. Like CBP, p300 interacted specifically with P/CAF. These studies demonstrated that P/CAF binds specifically to both p300 and CBP *in vitro*. In contrast to P/CAF, hGCN5 did not bind to CBP or p300.

These studies also demonstrated that the Zn²⁺ finger region of p300/CBP, which
shares sequence similarity with yADA2, is not essential for the interaction with P/CAF
Cloning of a human structural homolog of yADA2, termed hADA2 (25) has revealed
that, unlike the sequence similarity between p300/CBP and yADA2, which is restricted
to a 50 amino acid region, hADA2 shares extensive similarity (30% identity, 52%
similarity) to yADA2 over the entire protein sequence. Moreover, a computer search of
the complete genomic sequence of Saccharomyces cerevisiae revealed that yeast does

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not have counterparts of p300/CBP or P/CAF. Thus, the p300/CBP-P/CAF pathway may have been acquired during metazoan evolution.

Action of E1A in vitro 5

Previous reports indicated that E1A binds to both the p300 segment spanning residues 1767-1816 and the CBP segment spanning residues 1805-1854 (7). These interactions were reconfirmed in the present system; thus, both p300 and CBP segments covering the previously identified regions interacted with E1A.

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For further mapping, a series of deletions was introduced within the CBP segment-B and tested for interactions with P/CAF and E1A. Deletions of residues 1801-1825 or 1824-1851 markedly reduced interactions with both P/CAF and E1A, whereas deletion of residues 1850-1878 did not affect these interactions. Furthermore, deletion of residues 1801-1851 completely abolished interactions with both P/CAF and E1A. These data indicate that residues 1801-1851 of CBP are critical for interaction with both P/CAF and E1A. Taken together with the evidence that CBP segment A (aa residues 1,678-1,880) also binds to these factors, the above findings demonstrate that P7CAF and E1A bind to the same or very closely spaced sites on CBP.

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Evidence that both P/CAF and E1A recognize the same p300/CBP segments raises the possibility of direct competition between P/CAF and E1A for binding to p300/CBP. To test this possibility, a competition experiment was performed with the use of affinity purified recombinant proteins. The interaction of P/CAF with the CBPsegment B was progressively inhibited by the addition of increasing amounts of E1A. In contrast, no inhibition was caused by an E1A mutant which does not bind to p300/CBP (E1ADN). Similar results were obtained with the p300-segment B', leading to the conclusion that P/CAF and E1A compete for the same binding sites in p300/CBP.

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P/CAF-p300/CBP interaction in vivo

The *in vivo* interaction between P/CAF and p300/CBP was established by co-immunoprecipitation from a human osteosarcoma cell extract. Proteins in this extract were immunoprecipitated with rabbit anti-P/CAF, rabbit anti-CBP and anti-p300 antibodies. For controls, cell extract was precipitated with rabbit control IgG or mouse anti-HA monoclonal antibody. The precipitates were analyzed by immunoblotting with anti-P/CAF, anti-CBP and anti-p300 antibodies.

Osteosarcoma cells were transfected with either control vector or E1A- or

E1AAN-expression vectors. Extract from the transfected subpopulation was immunoprecipitated with anti-P/CAF or control IgG. The precipitates were analyzed by immunoblotting with anti-p300 and anti-P/CAF antibodies.

Rabbit anti-P/CAF antibody was raised to the P/CAF segment spanning residues 125-397 and purified by immunoaffinity chromatography (33). A mixture of 15 monoclonal antibodies raised to the human p300 segment spanning residues 1572-2371 (5) and rabbit polyclonal antibodies raised to the mouse CBP segment spanning residues 2-23 (for immunoprecipitation) and 1736-2179 (immunoblotting) were purchased from Upstate Biotechnology. Approximately 2 x 10⁷ human osteosarcoma U-2 OS cells (ATCC accession number HTB 96) were extracted with 10 ml of lysis buffer [25 mM 20 HEPES-KOH (pH 7.2), 150 mM potassium acetate, 2 mM EDTA, 1 mM DTT, 1 mM AEBSF, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 20 mM sodium fluoride, 0.1% NP40]. Two to 10 ml of extract were incubated with 2 µg of the respective antibody for four hours at 4°C. Fifty µl (packed volume) of protein-A Trisacryl (Pierce) were added and incubation was continued for two hours. The matrix 25 was washed four times with 1 ml of the lysis buffer, then boiled in 2x SDS sample buffer. Human osteosarcoma U-2 OS cells were transfected with 20 µg of the indicated plasmid and 1 µg of sorting plasmid (pCMV-IL2R) (31). The transfected subpopulation was purified by magnetic affinity cell sorting (32). Extract from approximately 2 x 10⁵ sorted cells was immunoprecipitated as described. 30

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Anti-P/CAF antibody specifically detected a 95 kDa protein, which is very close to the calculated value for the full-length P/CAF, in the immunoprecipitates. Anti-P/CAF antibody co-immunoprecipitated both CBP and p300. Similarly, anti-CBP antibody also co-immunoprecipitated P/CAF. However, anti-p300 antibody did not co-immunoprecipitate P/CAF. This is most likely due to steric interference since the anti-p300 antibody was raised to the p300 segment spanning residues 1572-2371 which includes the P/CAF binding region. These data demonstrate that P/CAF forms complexes with both p300 and CBP in vivo.

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10 Action of E1A in vivo

The *in vitro* experiments described herein indicate that P/CAF and E1A compete for the binding sites in p300/CBP. Thus, a study was conducted to determine whether E1A targets the endogenous interaction between P/CAF and p300. An E1A-expression vector was transiently transfected into human osteosarcoma cells and the transfected subpopulation was purified by cell sorting. Then, the interaction between P/CAF and p300 in transfected cells was examined by co-immunoprecipitation with anti-P/CAF antibody. The endogenous interaction of P/CAF with p300 was drastically inhibited by expression of E1A. On the other hand, no inhibition was observed by the E1A mutant lacking the p300 binding domain (E1AΔN), indicating that E1A disrupts the P/CAF-p300 complex *in vivo* through an interaction with p300.

Cell cycle regulation by P/CAF

Given that binding of P/CAF to p300/CBP is inhibited by E1A, experiments were performed to evaluate whether P/CAF, by binding to and forming a functional complex with p300, is involved in the regulation of entry into S phase. This possibility was addressed by examining whether transient expression of P/CAF would affect the rate of G1/S transit in HeLa cells. P/CAF negatively affected the distribution of cells between G1 and S phases in this assay.

HeLa cells were transfected by electroporation with 7 μ g of P/CAF-expression plasmid and/or 3 μ g of the full-length or the N-terminally deleted ($\Delta 2$ -36) E1A 12S-

expression plasmid as indicated. These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1 µg of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11 µg. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 h, and subsequently labeled in medium containing 10 µM bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32).

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The fraction of cells accumulating in S phase in control cultures was 23%, compared to 15% in P/CAF-transfected cells. This effect was reproducible in multiple independent experiments. In parallel experiments to verify the utility of this experimental protocol, plasmids encoding E2F-1, simian virus 40 small t, cyclin A or cyclin E increased the accumulation of cells in S phase, whereas plasmids encoding the cyclin-dependent kinase inhibitors p21 or p27 reduced the number of S phase cells.

On the basis of evidence that E1A and P/CAF compete for binding sites on p300, it seemed possible that cotransfection of P/CAF with E1A would oppose the mitogenic effect caused by E1A. As shown by the data herein, this is indeed the case. E1A alone has mitogenic activity in this experimental setting, while the E1A mutant lacking the p300 binding domain (E1ADN) has very weak activity. Comparable expression levels between wild type and mutant E1A in the transfected cells were revealed by immunoblotting analysis with anti-E1A. Intriguingly, when P/CAF was cotransfected with E1A, the mitogenic activity of E1A was significantly counteracted by P/CAF. These results show that P/CAF and E1A mediate antagonistic effects on cell cycle progression.

In the course of assessing P/CAF activity, it was also revealed that p300 is able to inhibit cell cycle progression under the same assay conditions. These findings suggest

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that P/CAF and p300, perhaps by forming a complex, act in concert to suppress cell cycle progression.

Histone acetyltransferase activity in P/CAF

Acetylation of the N-terminal histone tails has been considered to play a crucial role in accessibility of transcription factors to nucleosomal templates (26-27). Recently, yGCN5 has been identified as a histone acetyltransferase (28). On the basis of this information, intrinsic histone acetyltransferase activity in P/CAF and hGCN5 was examined. As substrates, the core histones (histones H2A, H2B, H3 and H4) and the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) were used.

Activity of hGCN5 and P/CAF that acetylates free histones or histones in the nucleosome core particle (35) was measured as described (36). Each reaction contained 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. The histone octamer dissociated into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE.

P/CAF and hGCN5 acetylated the core histones with almost the same efficiency. Both factors acetylated histones H3 and H4, but preferentially H3. In contrast, very weak or no acetylation by hGCN5 was detected in the nucleosome core particles. Remarkably, significant acetylation by P/CAF was observed in a nucleosomal context. Although all core histones are acetylated in the nucleus, P/CAF and hGCN5 did not acetylate histones H2A and H2B in vitro.

Direct function of P/CAF is likely to involve its intrinsic histone acetyltransferase activity. Although exact molecular mechanisms by which acetylation of core histones contribute to transcription remains undefined, acetylation of the histones is considered to play an important role in transcriptional regulation (26-27). The positively charged N-terminal tails of core histones are believed to affect nucleosome structure by interacting



with DNA at or near the nucleosome-spacer junction. Acetylation of the histone tails presumably destabilizes the nucleosome and facilitates access by regulatory factors. Likewise, there is a general correlation between the level of acetylation and transcriptional activity of nucleosomal domains. The findings of the present invention provide insights into the mechanisms of targeted histone acetylation.

Cellular factor p300/CBP binds to various sequence-specific factors that are involved in cell growth and/or differentiation, including CREB (3,4), c-Jun (9), Fos (11), c-Myb (12) and nuclear receptors (13). P/CAF could stimulate the activation function of these factors via promoter-specific histone acetylation. The present invention demonstrates that E1A appears to perturb normal cellular regulation by disrupting the connection between p300/CBP and its associated histone acetyltransferase.

II. p300/CBP studies.

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Purification of E1A associated histone acetyltransferase.

FLAG-epitope tagged E1A (or ΔE1A) was expressed in Sf9 cells (ATCC accession number CRL 1711) by infecting recombinant baculovirus (43). All purification steps were carried out at 4°C. Extract was prepared from infected cells by one cycle of freeze and thaw in buffer B (20 mM Tris-HCl, pH 8.0; 5 mM MgCl₂, 10% glycerol; 1 mM PMSF; 10 mMβ-mercaptoethanol, 0.1% Tween 20) containing 0.1 M KCl and the complete protease inhibitor cocktail (Boehringer Mannheim). To prepare E1A-immobilized beads, the extract was incubated with M2 anti-FLAG antibody agarose (Kodak-IBI) for four hours with rotating and subsequently washed with the same buffer three times. The resulting beads were incubated with HeLa (ATCC accession number CCL 2) nuclear extract for four to eight hours and thereafter washed with the same buffer six times. Finally, FLAG-E1A was eluted from the beads along with associated polypeptides by incubating with the same buffer containing 0.1 mg/ml FLAG peptide.

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For further purification, eluted polypeptides were dialyzed in 0.05 M KCl-buffer B and subsequently loaded onto a SMART Mono Q column (Pharmacia) equilibrated with the same 0.05 M KCl-buffer B. After washing, the column was developed with a linear gradient of 0.05-1.0 M KCl in buffer B. Mono Q fractions were concentrated with a MICROCON spin-filter (Amicon) and consequently loaded onto a SMART Superdex 200 column (Pharmacia) equilibrated with 0.1 M KCl-buffer B.

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Histone acetyltransferase assays

Filter binding assays were performed as described (80) with minor modifications. Samples were incubated at 30°C for 10-60 minutes in 30 ml of assay buffer containing 50 mM Tris-HCl, pH 8.0; 10% glycerol; 1 mM DTT; 1 mM PMSF; 10 mM sodium butyrate, 6 pmol of [3H]acetyl CoA (4.3 mCi/mmole, Amersham Life Science Inc.), and 33 mg/ml of calf thymus histones (Sigma Chemical Co.). In experiments where synthetic peptides were substituted for core histones, 50 pmol of each peptide were used. After incubation, the reaction mixture was spotted onto Whatman P-81 phosphocellulose filter paper and washed for 30 minutes with 0.2 M sodium carbonate buffer pH 9.2 at room temperature with 2-3 changes of the buffer. The dried filters were counted in a liquid scintillation counter.

PAGE analysis was done as above except that 90 pmol of [14C]acetyl CoA (55 mCi/mmole, Amersham Life Science Inc.) and 9 pmol of core histones or mononucleosomes were used. Core histones and mononucleosomes were prepared as described (35). For trypsin digestion, reaction mixtures were further incubated with various amounts of trypsin on ice for 30 minutes. The samples were analyzed on one dimensional SDS-PAGE gels or two dimensional gels, where the first dimension was an acid-urea-PAGE gel (44) and the second dimension was an SDS-PAGE gel.

Protein expression

For baculovirus expression, cDNAs corresponding to p300 portions of aa 1-670, aa 671-1194 and aa 1135-2414 were amplified by PCR (EXPAND High Fidelity PCR System, Boehringer Mannheim) as KpnI-NotI fragments. The resulting fragments were

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subcloned into a baculovirus transfer vector having the FLAG-tag sequence (43). The recombinant viruses were isolated using the BACULOGOLD system (Pharmingen), according to the manufacturer's protocol and were infected into Sf9 cells (ATCC accession number CRL 1711) to express FLAG-p300. Recombinant proteins were affinity purified with M2 anti-FLAG antibody-immobilized agarose (Kodak-IBI) according to the manufacturer's protocol.

For bacterial expression, cDNAs encoding the p300 portions and the CBP portion (aa 1174-1850) were first subcloned into the baculovirus transfer vector having the FLAG-tag as described above. Thereafter, the XhoI and NotI fragments encoding FLAG-p300 or FLAG-CBP fusions were resubcloned into the *E. coli* expression vector pET-28c (Novagene) digested with SalI and NotI. Recombinant proteins were expressed in *E. coli* BL21(DE3) and affinity purified with M2-antibody agarose.

15 Histone acetyltransferases that associate with E1A

Although the adenovirus E1A 12S protein (E1A) inhibits transcription in a variety of genes via direct binding to p300/CBP (45), E1A also stimulates transcription in some contexts (46). Thus, p300/CBP-bound E1A was tested to determine whether it might recruit histone acetyltransferases or deacetylases to regulate transcription. In addition, experiments were conducted as described below to determine if p300/CBP per se is a histone acetyltransferase.

Initially, recombinant FLAG-epitope tagged E1A was immobilized on anti-FLAG antibody beads. Immobilized E1A was incubated with a HeLa nuclear extract for affinity purification of E1A-associated polypeptides. FLAG-E1A was then eluted from the beads, along with E1A-associated polypeptides, by incubating with FLAG-peptide. Although E1A per se has no histone acetyltransferase activity, E1A recruited significant amounts of histone acetyltransferase activity from the nuclear extract. It is very unlikely that this activity is derived from P/CAF given that E1A and P/CAF cannot bind to p300/CBP simultaneously (43). Consistent with this, no P/CAF was detected in these fractions by immunoblotting.



The E1A N-terminus, a region that is not highly conserved among the various adenovirus serotypes, is involved in p300/CBP binding *in vivo*. Mutations in the N-terminal region lead to loss of the ability for p300/CBP binding without affecting RB binding (1,47). Thus, the requirement of the E1A N-terminal region for the recruitment of histone acetyltransferase activity was tested. In contrast to the wild type, the N-terminal deleted form of E1A (Δ N-E1A) recruited only a background level of acetyltransferase activity. In agreement with previous reports (47), the Δ N-E1A showed no ability to interact with p300/CBP, although it still retained the ability to interact with a variety of other polypeptides, including RB

To define the relationship between p300/CBP and histone acetyltransferase activity, affinity purified E1A-binding polypeptides were separated by Mono Q ion-exchange column. Both p300/CBP and the acetyltransferase activity were coeluted at 140 mM KCl, while most of polypeptides were eluted at 260 mM KCl. The active fraction of Mono Q column (~140 mM KCl) was further separated by Superdex-200 gel filtration column. Both p300/CBP and the acetyltransferase activity coeluted after the void volume, indicating that p300/CBP is involved in the histone acetyltransferase activity.

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p300 is a histone acetyltransferase

The data provided herein indicate that p300 per se, or a polypeptide(s) associated with p300, possesses histone acetyltransferase activity. To test the former possibility, the acetyltransferase activity of recombinant p300 was measured. p300 was divided into three fragments, each of which was expressed in and purified from Sf9 cells via a baculovirus expression vector. Histone acetyltransferase activity was readily detected in the C-terminal fragment containing amino acids 1135-2414, whereas no activity was found in the other fragments, demonstrating conclusively that p300 per se is a histone acetyltransferase.

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p300/CBP-histone acetyltransferase domain

To map the histone acetyltransferase domain of p300, a series of deletions was prepared. Given the poor conservation of the glutamine-rich region (aa 1815-2414) in the C. elegans p300/CBP homolog (6), the p300 fragment encoding aa 1135-1810 was expressed in and purified from E. coli. Importantly, this candidate region of p300 (aa 1135-1810) showed significant histone acetyltransferase activity. For further mapping within this region, a series of N-terminal deletions was constructed. Deletion of 60 residues, resulting in a fragment containing aa 1195-1810, had no effect on the acetyltransferase activity, whereas the deletion of 185 residues, yielding a fragment comprising aa residues 1320-1810, completely eliminated the acetyltransferase activity.

Next, a series of C-terminal deletions was analyzed to determine the requirement of the P/CAF (or E1A) -binding domain. The p300 fragments lacking the E1A binding domain (aa 1195-1760, 1195-1706 and 1195-1673) still retained the acetyltransferase activity, whereas the further truncated mutant (aa 1195-1652) completely lost the acetyltransferase activity. Consistent with these results, the internal deletion of residues 1418-1720 showed no acetyltransferase activity. These data demonstrate that the histone acetyltransferase domain is located between the bromodomain and the -E1A-binding domain. Given that the histone acetyltransferase domain is highly conserved between p300 and CBP (91% similarity), the corresponding region of CBP. aa residues 1174-1850, was expressed to confirm the acetyltransferase activity. As expected, comparable activity was detected, indicating that both p300 and CBP are histone acetyltransferases.

Among various acetyltransferases including histone acetyltransferases GCN5 and P/CAF, putative acetyl-CoA binding sites are conserved (48). However, multiple alignment analysis (49) showed that the p300/CBP histone acetyltransferase domain does not belong to this group. Moreover, comparison of the p300/CBP histone acetyltransferase domain with peptide sequence databases (23) showed no sequence similarity to any other proteins. Accordingly, this invention shows that p300/CBP 30 represents a novel class of acetyltransferases in that it does not have the conserved motif found among previously described acetyltransferases (48).



p300 acetylates all core histones in mononucleosomes

Substrate specificity for acetylation by p300 was also examined. As substrates, histone octamers and mononucleosomes (146 base pairs of DNA wrapped around the octamer of core histones) were used. Given that the histone octamer dissociates into dimers or tetramers under physiological conditions, the histone octamer is referred to here as core histones. When core histones were used, p300 acetylated all four proteins, but preferentially H3 and H4. More importantly, in a nucleosomal context, p300 acetylated all four core histones nearly stoichiometrically. In contrast, p300 acetylated neither BSA nor lysozyme.

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Hyperacetylated histones are believed to be linked with transcriptionally active chromatin (26,27,50,51). Hyperacetylated forms are found in histones H4, H3 and H2B, which have multiple acetylation sites *in vivo*. Thus, the level of acetylation by p300 was also tested.

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Mononucleosomes treated with p300 were analyzed by two-dimensional gel electrophoresis. A Coomassie blue-stained gel and the corresponding autoradiogram showed that a significant amount of histones, especially H4, were hyperacetylated. Importantly, acetylation levels by p300 were very close to those of hyperacetylated histones prepared from HeLa nuclei treated with sodium butyrate, a histone deacetylase inhibitor. In contrast, no acetylated forms were detected in the reaction without p300. These results indicate that p300 acetylates histones in mononucleosomes to the hyperacetylated state by targeting multiple lysine residues.

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p300 acetylates the four lysines in the histone H4 N-terminal tail in vitro which are acetylated in vivo

Lysines at positions 5, 8, 12 and 16 of histone H4 are acetylated *in vivo* (51). Recent studies with yeast histone acetyltransferases demonstrate the position-specific acetylation by distinct acetyltransferases, i.e., while cytoplasmic acetyltransferases for histone deposition and chromatin assembly modify



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positions 5 and 12, GCN5 modifies positions 8 and 16 (52). Accordingly, the positions of acetylation by p300 were also determined. A series of synthetic peptides containing acetylated lysines at various positions was used to determine the acetylation site-specificity of p300. Consistent with the two-dimensional gel electrophoresis analysis, the experiments with peptide substrates showed that p300 acetylates all four lysines in the histone H4 that are acetylated *in vivo*. These results are consistent with the view that deposition-related diacetylated histones are deacetylated during maturation of chromatin (53).

10 p300 preferentially acetylates the N-terminal histone tail

Histone acetyltransferases modify specific lysine residues in the N-terminal tail of core histones but not the C-terminal globular domain *in vivo* (26,27,50,51). Structural models of nucleosomes (54,55,56) suggest that most of the lysine residues in the C-terminal globular domain are buried. Therefore, experiments were conducted to examine whether restricted acetylation of the N-terminal tail resulted from the substrate specificity of the enzyme or inaccessibility of the enzyme to the core domain in nucleosomes. The globular domains of all core histones contain a long helix flanked on either side by a loop segment and short helix, termed the "histone fold" (54,55,56). The histone fold is involved in formation of the stable H2A-H2B and H3-H4 hetero-dimers, consisting of extensive hydrophobic contacts between the paired molecules. Therefore, it is likely that a histone monomer cannot fold properly, thereby increasing access of the histone acetyltransferase to the core domain. Based on these considerations, experiments were conducted to determine whether p300 acetylates free histone H4 in a N-terminal-specific manner.

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Histone H4 was acetylated with p300 and subsequently the histone tail was removed by partial digestion with trypsin. The distributions of radioactivity between intact and core histones were compared. While the globular core histone domain was predominant at the higher trypsin concentrations, radioactivity was detected mostly in the intact histone. These data demonstrate that p300 preferentially acetylates the N-terminal tail of histone H4.

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III. P/CAF interaction with MyoD

Tissue culture and transfection experiments

 C_2C_{12} mouse cells (ATCC accession number CRL 1772) were grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 20% fetal bovine serum (FBS) until they reached confluence. Differentiation was induced by switching medium to differentiation medium (DM), consisting of DMEM containing 2% horse serum. $C_3H/10T1/2$ fibroblasts (ATCC accession number CCL 226) were grown in DMEM supplemented with 10% FBS. Cells were transfected by the calcium phosphate precipitation method. Total amounts of transfected DNA were equalized by empty vector DNA. After 12 h incubation in medium containing the precipitated DNA, the cells were washed and incubated in fresh DMEM containing 10% FBS for an additional 24 h. Afterwards, differentiation was induced by incubating in DM for 36 to 72 h. Chloramphenicol acetyltransferase (CAT) assays were performed as previously described (64,69). The quantities of cell extracts used for CAT assays were normalized to β -galactosidase activity by cotransfection of 1 mg of the β -galactosidase expression vector, pON260.

Expression vectors used for transfection experiments are as follows: pCX-P/CAF for P/CAF (43), pCMV-bp300 for p300 (65), pCMV-p300 (1869-2414) (64) and pCMV-p300 (1514-1922) (60) for p300 wild type and mutants, pE1A12S, pE1A12S R2G, pE1A12S D2-36 and pE1A12S D121-130 for E1A wild type and mutants (66,67,68), and pEMSV-MyoD for MyoD (64)

The antisense P/CAF RNA expression vector, pcDNA3 P/CAF-AS, was created as follows. The 2.5 Kb EcoRI-KpnI fragment containing the entire P/CAF open reading frame was isolated from pCX-P/CAF (43). This fragment was subcloned into the

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EcoRI-KpnI sites of plasmid pcDNA3 (Invitrogen) so that the antisense P/CAF RNA is driven under the CMV promoter. Reporter genes employed were 4RE-CAT and MCK-CAT (69). 4RE-CAT is driven by a synthetic promoter containing 4 copies of the E-box, whereas MCK-CAT is driven by the native MCK promoter (nucleotides -1256 to +7).

Microinjection and immunofluorescence

Cells were grown on small glass slides, subdivided into numbered squares of 2 mm x 2 mm and microinjected with purified and concentrated antibodies, as previously described (70). For immunofluorescence, cells were fixed in either 2% paraformaldehyde or 1:2 methanol/acetone solution, preincubated with 5% BSA/PBS and incubated with the primary antibodies for 30 min at 37° C. Subsequently, antibody was visualized by incubating with either rhodamine- or fluorescein-conjugated secondary antibody for 30 min at 37° C. Injected antibodies were stained with a rhodamine-conjugated secondary antibody and nuclei were counter-stained by DAPI as previously described (69).

Antibodies employed are as follows; rabbit polyclonal affinity purified anti-P/CAF antibody (43), rabbit polyclonal anti-p300/CBP antiserum (71), mouse monoclonal anti-MyoD antibody (clone 5.8A, kindly provided by P. Houghton), goat polyclonal anti-c-Jun affinity purified antibody (Santa Cruz) and rabbit pre-immune serum.

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Immunoprecipitation and DNA affinity purification

Cells were resuspended in lysis buffer (20 mM NaPO₄, 150 mM NaCl, 5mM MgCl₂, 0.1% NP40, 1 mM DTT, 10 mM sodium fluoride, 0.1 mM sodium vanadate, 1 mM phenylmethylsulfonyl-fluoride and 10 mg/ml each of leupeptin, aprotinin and pepstatin). After 30 min incubation on ice, samples were centrifuged at 12,000 x g for 30 min and supernatants were used as cell extracts. Extracts were pre-cleared by



incubating with rabbit pre-immune serum and protein A/G Plus-Agarose (Santa Cruz) for 2 h at 4 C. For immunoprecipitation, the supernatants were incubated with the respective antibodies for 3 h at 4 C. Protein A/G Plus-Agarose was added, and incubation continued for 3 h. The matrix was washed with lysis buffer, then boiled in 2 X SDS sample buffer. Immunoblotting was performed by using the ECL chemiluminescent detection kit (Amersham) according to the manufacturer's protocol

Affinity purification of E-box-bound complexes was done as previously described (69). Briefly, 100 ng of the biotinylated double stranded DNA containing the E-box were immobilized on streptavidin-conjugated magnetic beads and incubated with 500 mg of cell extracts in the presence of poly dI-dC. After extensive washing, bound proteins were eluted with SDS sample buffer and analyzed by immunoblotting

In vitro protein-protein interaction assays

The CBP-B fragment and its deletion derivatives were expressed as 15 GST-fusions described previously (43). MyoD and E1A (43) were expressed as FLAG-fusion proteins in Sf9 cells via a baculovirus expression system and affinity-purified on M2 anti-FLAG antibody-agarose (Kodak-IBI). Crude E. coli extracts containing GST-fusions were incubated with various amounts of MyoD and/or E1A in 50 ml of buffer B (20 mM Tris-HCl, pH 8.0, 0.1 M KCl, 5 mM MgCl₂, 10% 20 glycerol, and 0.1% Nonidet P-40) on ice for 10 min. GST-precipitation was performed as described (43). MyoD and E1A were detected by immunoblotting with anti-FLAG M2 antibody. For the interaction between P/CAF and MyoD, 1.5 pmol of FLAG-P/CAF and 15 pmol of FLAG-MyoD were incubated in 50 ml of buffer B on ice for 10 min. The mixture was further incubated with 2 mg of anti-P/CAF (43) or anti-hADA2 antibody for 60 min. The immunocomplexes were precipitated by incubation with 10 ml of protein A-Trisacryl (Pierce) and rotated for 1-4 hr at 4oC. The matrix was washed 4 times with 200 ml of buffer B and boiled in 10 ml of 2 X SDS sample buffer. The proteins were resolved on a 4%-20% gradient SDS-PAGE and subjected to immunoblotting with the anti-FLAG M2 antibody. The blot was developed 30 with the SUPERSIGNAL chemiluminescent substrates (Pierce).



P/CAF coactivates muscle-specific transcription

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P/CAF and MyoD were co-transfected into mouse C3H10T1/2 fibroblasts, and MvoD-mediated transcription was determined from reporter activity driven by the artificial (4RE) and the naturally-occurring muscle creatine kinase (MCK) promoters. Overexpression of P/CAF stimulated MyoD-dependent transcription several folds in both promoters. Similar results were obtained for the myoD activated myogenin promoter Transcriptional activation was further stimulated by co-transfecting with MyoD, P/CAF and p300 expression vectors, suggesting that P/CAF may function by forming a complex with p300/CBP. Consistent with the lack of DNA binding capacity in P/CAF, overexpression of P/CAF alone did not increase the basal transcriptional activity of either enhancer. To test whether P/CAF and p300/CBP function in the same pathway, two dominant negative forms of p300 were employed which specifically inhibit p300/CBP-mediated transcription (60,64). The p300 segment spanning residues 1514-1922 inhibits the MyoD-dependent activation via direct interaction with MyoD (60), whereas the p300 segment spanning residues 1869-2414 inhibit it without direct interaction (64). Both dominant negative mutants inhibited MyoD-coactivation by P/CAF), suggesting that P/CAF and p300/CBP function in the same pathway.

For further elucidation of the activation mechanism by P/CAF, the effect of E1A, which inhibits MyoD-dependent transcription and differentiation (66,72,73) via direct interaction with p300/CBP (65,78), was tested. Expression of E1A in C3H10T1/2 fibroblasts inhibited stimulation of MyoD-directed transcription by P/CAF overexpression. E1A mutants lacking p300/CBP-binding activity, E1A D2-36 and E1A R2G (67,79), had almost no effect. On the other hand, an E1A mutant retaining p300/CBP-binding activity, E1A D121-130, behaved like the wild type. Since E1A associates with p300/CBP, but not with P/CAF, these results suggest that P/CAF functions in MyoD-directed transcription via interaction with p300/CBP

To address the role of P/CAF as a myogenic coactivator in a more relevant environment, P/CAF was overexpressed in proliferating C2C12 myoblasts which express

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endogenous myogenic bHLH factors. As observed in fibroblasts, overexpression of P/CAF stimulated muscle specific transcription. Concomitant expression of exogenous p300 increased P/CAF-mediated coactivation. The repression exerted by wild type E1A, but not mutant E1A D2-36, on P/CAF coactivation of MyoD was also observed in muscle cells.

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Similar experiments were performed with myogenic cell lines that were stably transformed with wild type or mutant E1A-expressing vectors (66). Coactivation by P/CAF was inhibited by wild type E1A or the E1A mutant that retains p300/CBP-binding activity (E1A\Delta121-130). In contrast, E1A mutants that lack p300/CBP-binding (E1A \Delta2-36 and E1A R2G) allowed transcriptional coactivation by P/CAF. Taken together, these experiments show that P/CAF coactivates MyoD-directed transcription via interaction with p300/CBP.

15 P/CAF stimulates myogenic differentiation

Given that P/CAF potentiates MyoD-directed transcription, the ability of P/CAF to assist MyoD in promoting myogenic differentiation was investigated. To this aim, C3H10T1/2 fibroblasts were transiently transfected with P/CAF and MyoD expression vectors. An expression vector for the green fluorescent protein (GFP) was co-transfected to identify transfected cells. After incubation in differentiation medium, the myogenic conversion of transfected cells was determined by simultaneous expression of the GFP and the differentiation-specific marker myosin heavy chain (MHC). Forced expression of MyoD in fibroblasts caused muscle differentiation in 12% of the transfected fibroblasts. This myogenic conversion was 20% by co-expressing MyoD and P/CAF. As observed in transcription experiments, stimulation of differentiation by P/CAF was counteracted by co-transfection with the p300 dominant negative mutant, p300 (1869-2414). Consistent with a general role for coactivators, overexpression of P/CAF alone was unable to differentiate fibroblasts.

Similar experiments were done using proliferating C2C12 myoblasts in which the differentiation program is already committed. Most of the myoblasts differentiated into

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myotubes by overexpressing P/CAF, whereas only a modest effect was observed by overexpressing p300. In contrast, differentiation was inhibited slightly by overexpressing c-Jun. This inhibitory effect presumably was caused by titration of p300/CBP, which associates directly with c-Jun (74). A similar inhibition was observed in the p300 dominant negative mutant. Consistent with the transcriptional effect, E1A almost completely inhibited differentiation. The E1A mutant RG2, lacking p300/CBP-binding capability but retaining the retinoblastoma protein (Rb)-binding capability, only partially inhibited differentiation, although this same mutant inhibited transcription as severely as the wild type. Taken together, these data show that P/CAF stimulates muscle differentiation by coactivating MyoD function via p300/CBP association.

P/CAF is essential for myogenic transcription and differentiation

To test the necessity of P/CAF for myogenic transcription, experiments were conducted whereby P/CAF synthesis was inhibited by expressing antisense P/CAF RNA A vector from which the P/CAF mRNA is transcribed in the antisense orientation (P/CAF-AS) was transfected with P/CAF and MyoD expression vectors into fibroblasts and MyoD-dependent transcription was examined. Cotransfection of the antisense expression vector strongly inhibited MyoD-dependent transcription below the level of induction elucidated by MyoD alone, demonstrating that expression of P/CAF antisense RNA inhibits not only the coactivation exerted by exogenous P/CAF but also that of endogenous P/CAF. These results indicate that P/CAF is essential for MyoD-dependent transcription.

Studies were also carried out to determine whether expression of P/CAF antisense RNA inhibits myogenic differentiation. C3H10T1/2 fibroblasts were transiently transfected with various expression vectors with or without the P/CAF antisense RNA expression vector. Expression of P/CAF antisense RNA reduced MyoD-mediated myogenic conversion of fibroblasts. Expression of P/CAF antisense RNA also counteracted the stimulatory effect of both P/CAF and p300 on myogenic differentiation. These data support the view that P/CAF and p300/CBP coactivate

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MyoD-dependent transcription in the same pathway. More drastic inhibition was observed in C2C12 myoblasts in similar experiments. Therefore, it can be concluded that P/CAF is essential for transcription of muscle specific genes and hence differentiation into myotubes.

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To further confirm the essential role of P/CAF for myogenic differentiation, we blockage experiments by antibody microinjection were performed. Antibodies were injected into the cytoplasm of proliferating C2C12 myoblasts to prevent the nuclear transport of newly synthesized target proteins. After incubating in the differentiation medium, the degree of differentiation was determined. Microinjection of an anti-P/CAF antibody almost completely inhibited differentiation. Similar results were obtained by microinjecting anti-p300/CBP antibodies. Although microinjection of either anti-p300/CBP or P/CAF antibody was sufficient to inhibit differentiation, an even greater inhibition was observed by coinjecting both of them. Microinjection of anti-P/CAF or anti-p300/CBP antibody did not interfere with induction of p53 by DNA damaging agents, showing specificity of the inhibition by the antibodies. In contrast to anti-P/CAF or anti-p300/CBP antibodies, the injection of anti-MyoD antibody only partially inhibited differentiation, supporting the view of functional redundancy between MyoD and Myf-5 (75,76). Injection of anti-c-Jun antibody or control antibody did not interfere with muscle differentiation.

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Similar experiments were performed with C3H10T1/2 fibroblasts stably expressing MyoD. In these cells, either anti-p300/CBP or anti-P/CAF antibody completely inhibited muscle differentiation. In contrast to myoblasts, anti-MyoD antibody completely blocked differentiation in the fibroblasts expressing MyoD. Anti-c-Jun and control antibodies did not interfere with differentiation. Taken together, these results demonstrate that P/CAF and p300/CBP are indispensable for activation of the myogenic program.

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The data described above indicate that P/CAF stimulates MyoD-directed transcription via association with p300/CBP. Thus, experiments were conducted to

p300/CBP, P/CAF and MyoD form a multimeric complex in vivo

investigate whether P/CAF, p300/CBP and MyoD could associate in a complex.

First, cellular extracts derived from C2C12 myotubes were subjected to immunoprecipitation. Both anti-MyoD and anti-p300/CBP antibodies co-precipitated P/CAF. In a complementary experiment, both anti-p300/CBP and anti-P/CAF antibodies also co-precipitated MyoD, suggesting that these factors form a multimeric protein complex in myotubes.

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Next, attempts were made to detect this complex on the E-box, the DNA binding site for MyoD. Immobilized DNA containing an E-box sequence was incubated with myotube extracts. After extensive washing, P/CAF, p300/CBP and MyoD were analyzed by immunoblotting. P/CAF, p300/CBP and MyoD were all affinity purified on the immobilized DNA, whereas they were not purified on the control DNA lacking the E-box. Given that P/CAF and p300/CBP per se cannot bind to DNA, these observations indicate that P/CAF and p300/CBP are recruited through MyoD at the E-box sites to form a multi-protein complex.

20 Complex formation is inhibited by viral transforming factors

Since the oncoviral proteins E1A and large T antigen inhibit myogenic transcription and differentiation, the effect of these factors on the formation of complexes on the E-box was tested. Importantly, very small amouts of P/CAF and p300/CBP were co-purified on the E-box from myocyte extracts which stably express E1A or large T antigen, although MyoD was detected under these conditions. The lower recovery of MyoD from E1A-expressing muscle cells could reflect the low level of MyoD in the extracts (66). These results indicate that E1A and large T antigen dissociate P/CAF and p300/CBP from MyoD without altering MyoD binding to DNA.

Consistent with the previous observations that transiently expressed E1A prevents interaction between P/CAF and p300/CBP in vivo (43), the association

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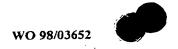
between p300/CBP and P/CAF was abolished in myoblasts stably transformed by wild type E1A but not in those clones transformed with the E1A mutant R2G unable to bind p300/CBP. Similarly, the interaction between p300/CBP and P/CAF was abolished by large T antigen but not by the mutant protein that localizes into the cytoplasm (77).

Interaction between MyoD, P/CAF and CBP in vitro

Previous interaction experiments *in vitro* indicate that the CBP region spanning residues 1801 to 1850 is crucial for interaction with both P/CAF and E1A (43). While most sequence-specific factors bind to CBP sites distinct from the P/CAF/E1A binding sites, MyoD interacts with an overlapping CBP fragment called the CH3 region (60,64,65). To understand how P/CAF, p300/CBP and MyoD associate, the CBP sites important for MyoD binding were mapped more precisely. Consistent with previous reports (60,64,65), the CBP fragment spanning residues 1801-2000 (fragment B) bound MyoD. Moreover, deletion of residues 1801 to 1850 within fragment B completely abolished interaction with MyoD, which is similar to the results obtained with P/CAF and E1A. Importantly, an internal deletion of residues 1850-1878 abolished the MyoD interaction with CBP, while it did not affect binding of E1A or P/CAF (43). These results suggest that MyoD and P/CAF bind to distinct sites of p300/CBP, albeit the binding sites may overlap. Moreover, a direct interaction was observed between MyoD and P/CAF, which may contribute to stabilization of the multimeric complex.

These data show that E1A prevents not only p300/CBP-interaction with P/CAF but also that with MyoD in vivo. To obtain evidence that this inhibition is due to the direct action by E1A, competition experiments were performed in vitro. Importantly, the interaction between CBP and MyoD was strongly inhibited by addition of E1A, implicating that E1A inhibits myogenic transcription by disrupting multiple interactions.

Although the present process has been described with reference to specific details of certain embodiments thereof, it is not intended that such details should be





regarded as limitations upon the scope of the invention except as and to the extent that they are included in the accompanying claims.

Throughout this application various publications are referenced by numbers

within parentheses. Full citations for these publications are as follows. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

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SEQUENCE LISTING

- (1) GENERAL INFORMATION.
- (i) APPLICANT: The United States of America, as repesented by the Secretary, Department of Health and Human Services, c/o National Institutes of Health, Office of Technology Transfer, 6011 Executive Boulevard, Suite 325, Rockville, Maryland 20842
- (ii) TITLE OF THE INVENTION: METHODS AND COMPOSITIONS FOR p300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF

- (iii) NUMBER OF SEQUENCES: 18
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 - (E) COUNTRY: USA
 - (F) ZIP: 30303
- (V) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: DOS
 - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE: 23-JUL-1997
- (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: Corresponding U.S. Serial No. 60/022,273
 - (B) FILING DATE: 23-July-1996
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Miller, Mary L
 - (B) REGISTRATION NUMBER: 39,303
 - (C) REFERENCE/DOCKET NUMBER: 14014.0238/P
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 404/688-0770
 - (B) TELEFAX: 404/688-9880
 - (C) TELEX:
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 832 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
- . (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: None





(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1: Met Ser Glu Ala Gly Gly Ala Gly Pro Gly Gly Cys Gly Ala Gly Ala Gly Ala Gly Ala Gly Pro Gly Ala Leu Pro Pro Gln Pro Ala Ala Leu Pro Pro Ala Pro Pro Gln Gly Ser Pro Cys Ala Ala Ala Ala Gly Gly Ser Gly Ala Cys Gly Pro Ala Thr Ala Val Ala Ala Ala Gly Thr Ala Glu Gly Pro Gly Gly Gly Ser Ala Arg Ile Ala Vai Lys Lys Ala Gln Leu Arg Ser Ala Pro Arg Ala Lys Lys Leu Glu Lys Leu Gly Val Tyr Ser Ala Cys Lys Ala Glu Glu Ser Cys Lys Cys Asn Gly Trp Lys Asn Pro Asn Pro Ser Pro Thr Pro Pro Arg Ala Asp Leu Gln Gln Ile Ile Val Ser Leu Thr Glu Ser Cys Arg Ser Cys Ser His Ala Leu Ala Ala His Val Ser His Leu Glu Asn Val Ser Glu Glu Met Asn Arg Leu Leu Gly Ile Val Leu Asp Val Glu Tyr Leu Phe Thr Cys Val His Lys Glu Glu Asp Ala Asp Thr Lys Gln Val Tyr Phe Tyr Leu Phe Lys Leu Leu Arg Lys Ser Ile Leu Gln Arg Gly Lys Pro Val Val Glu Gly Ser Leu Glu Lys Lys Pro Pro Phe Glu Lys Pro Ser Ile Glu Gln Gly Val Asn Asn Phe Val Gln Tyr Lys Phe Ser His Leu Pro Ala Lys Glu 235 . . Arg Gin Thr Ile Val Glu Leu Ala Lys Met Phe Leu Asn Arg Ile Asn Tyr Trp His Leu Glu Ala Pro Ser Gln Arg Arg Leu Arg Ser Pro Asn Asp Asp Ile Ser Gly Tyr Lys Glu Asn Tyr Thr Arg Trp Leu Cys Tyr Cys Asn Val Pro Gln Phe Cys Asp Ser Leu Pro Arg Tyr Glu Thr Thr . 300 Gln Val Phe Gly Arg Thr Leu Leu Arg Ser Val Phe Thr Val Met Arg Arg Gln Leu Leu Glu Gln Ala Arg Gln Glu Lys Asp Lys Leu Pro Leu Glu Lys Arg Thr Leu Ile Leu Thr His Phe Pro Lys Phe Leu Ser Met Leu Glu Glu Glu Val Tyr Ser Gln Asn Ser Pro Ile Trp Asp Gln Asp Phe Leu Ser Ala Ser Ser Arg Thr Ser Gln Leu Gly Ile Gln Thr Val Ile Asn Pro Pro Pro Val Ala Gly Thr Ile Ser Tyr Asn Ser Thr Ser Ser Ser Leu Glu Gln Pro Asn Ala Gly Ser Ser Pro Ala Cys Lys Ala Ser Ser Gly Leu Glu Ala Asn Pro Gly Glu Lys Arg Lys Met Thr Asp Ser His Val Leu Glu Glu Ala Lys Lys Pro Arg Val Met Gly Asp

Ile Pro Met Glu Leu Ile Asn Glu Val Met Ser Thr Ile Thr Asp Pro

Ala Ala Met Leu Gly Pro Glu Thr Asn Phe Leu Ser Ala His Ser Ala



Arg Asp Glu Ala Ala Arg Leu Glu Glu Arg Arg Gly Val Ile Glu Phe 490 485 His Val Val Gly Asn Ser Leu Asn Gln Lys Pro Asn Lys Lys Ile Leu 510 505 500 Met Trp Leu Val Gly Leu Gln Asn Val Phe Ser His Gln Leu Pro Arg 520 Met Pro Lys Glu Tyr Ile Thr Arg Leu Val Phe Asp Pro Lys His Lys 535 Thr Leu Ala Leu Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe 550 555 Arg Met Phe Pro Ser Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val 570 565 Thr Ser Asn Glu Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His 590 585 580 Leu Lys Glu Tyr His Ile Lys His Asp Ile Leu Asn Phe Leu Thr Tyr 605 600 595 Ala Asp Glu Tyr Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys 620 615 Glu Ile Lys Ile Pro Lys Thr Lys Tyr Val Gly Tyr Ile Lys Asp Tyr 635 630 Glu Gly Ala Thr Leu Met Gly Cys Glu Leu Asn Pro Arg Ile Pro Tyr 650 645 Thr Glu Phe Ser Val Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys 670 665 660 . Leu Ile Glu Arg Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu 685 680 675 Ser Cys Phe Lys Asp Gly Val Arg Gln Ile Pro Ile Glu Ser Ile Pro 700 695 Gly Ile Arg Glu Thr Gly Trp Lys Pro Ser Gly Lys Glu Lys Ser Lys 715 710 Glu Pro Arg Asp Pro Asp Gln Leu Tyr Ser Thr Leu Lys Ser Ile Leu 735 730 725 Gln Gln Val Lys Ser His Gln Ser Ala Trp Pro Phe Met Glu Pro Val 750 740 745 Lys Arg Thr Glu Ala Pro Gly Tyr Tyr Glu Val Ile Arg Ser Pro Met 760 765 Asp Leu Lys Thr Met Ser Glu Arg Leu Lys Asn Arg Tyr Tyr Val Ser 780 775 Lys Lys Leu Phe Met Ala Asp Leu Gln Arg Val Phe Thr Asn Cys Lys 790 795 Glu Tyr Asn Ala Pro Glu Ser Glu Tyr Tyr Lys Cys Ala Asn Ile Leu 805 -810 815 Glu Lys Phe Phe Ser Lys Ile Lys Glu Ala Gly Leu Ile Asp Lys 820 . 825

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 481 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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 Trp
 Asp
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 Arg
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 Gln
 Leu
 Gly
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 Gln
 Thr

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 Asn
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Ser Ser Ser Leu Glu Gln Pro Asn Ala Gly Ser Ser Pro Ala Cys Lys Ala Ser Ser Gly Leu Glu Ala Asn Pro Gly Glu Lys Arg Lys Met Thr Asp Ser His Val Leu Glu Glu Ala Lys Lys Pro Arg Val Met Gly Asp Ile Pro Met Glu Leu Ile Asn Glu Val Met Ser Thr Ile Thr Asp Pro Ala Ala Met Leu Gly Pro Glu Thr Asn Phe Leu Ser Ala His Ser Ala Arg Asp Glu Ala Ala Arg Leu Glu Glu Arg Arg Gly Val Ile Glu · 135 Phe His Val Val Gly Asn Ser Leu Asn Gln Lys Pro Asn Lys Lys Ile Leu Met Trp Leu Val Gly Leu Gln Asn Val Phe Ser His Gln Leu Pro Arg Met Pro Lys Glu Tyr Ile Thr Arg Leu Val Phe Asp Pro Lys His Lys Thr Leu Ala Leu Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe Pro Ser Gln Gly Phe Thr Glu Ile Val Phe Cys Ala 210 215 Val Thr Ser Asn Glu Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn - 230 His Leu Lys Glu Tyr His Ile Lys His Asp Ile Leu Asn Phe Leu Thr Tyr Ala Asp Glu Tyr Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Glu Ile Lys Ile Pro Lys Thr Lys Tyr Val Gly Tyr Ile Lys Asp Tyr Glu Gly Ala Thr Leu Met Gly Cys Glu Leu Asn Pro Arg Ile Pro Tyr Thr Glu Phe Ser Val Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu Arg Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe Lys Asp Gly Val Arg Gln Ile Pro Ile Glu Ser Ile Pro Gly Ile Arg Glu Thr Gly Trp Lys Pro Ser Gly Lys Glu Lys Ser Lys Glu Pro Arg Asp Pro Asp Gln Leu Tyr Ser Thr Leu Lys Ser Ile Leu Gln Gln Val Lys Ser His Gln Ser Ala Trp Pro Phe Met Glu Pro Val Lys Arg Thr Glu Ala Pro Gly Tyr Tyr Glu Val Ile Arg Ser Pro Met Asp Leu Lys Thr Met Ser Glu Arg Leu Lys Asn Arg Tyr Tyr Val Ser Lys Lys Leu Phe Met Ala Asp Leu Gln Arg Val Phe Thr Asn Cys Lys Glu Tyr Asn Ala Pro Glu Ser Glu Tyr Tyr Lys Cys Ala Asn Ile Leu Glu Lys Phe Phe Phe Ser Lys Ile Lys Glu Ala Gly Leu Ile Asp Lys

(2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 203 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single



(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Arg Val Val Gln His Thr Lys Gly Cys Lys Arg Lys Thr Asn Gly Gly Cys Pro Ile Cys Lys Gln Leu Ile Ala Leu Cys Cys Tyr His Ala Lys 25 20 His Cys Gln Glu Asn Lys Cys Pro Val Pro Phe Cys Leù Asn Ile Lys 45 40 . Gln Lys Leu Arg Gln Gln Gln Leu Gln His Arg Leu Gln Gln Ala Gln 60 - 55 Met Leu Arg Arg Arg Met Ala Ser Met Arg Thr Gly Val Val Gly Gln 75 70 Gln Gln Gly Leu Pro Ser Pro Thr Pro Ala Thr Pro Thr Thr Pro Thr 85 . 90 . Gly Gln Gln Pro Thr Thr Pro Gln Thr Pro Gln Pro Thr Ser Gln Pro 105 100 Gln Pro Thr Pro Pro Asn Ser Met Pro Pro Tyr Leu Pro Arg Thr Gln 125 120 115 Ala Ala Gly Pro Val Ser Gln Gly Lys Ala Ala Gly Gln Val Thr Pro 140 135 130 Pro Thr Pro Pro Gln Thr Ala Gln Pro Pro Leu Pro Gly Pro Pro Pro 155 150 Thr Ala Val Glu Met Ala Met Gln Ile Gln Arg Ala Ala Glu Thr Gln 170 165 Arg Gln Met Ala His Val Gln Ile Phe Gln Arg Pro Ile Gln His Gln 185 180 Met Pro Pro Met Thr Pro Met Ala Pro Met Gly 200 195

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 351 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ser Glu Ala Gly Gly Ala Gly Pro Gly Gly Cys Gly Ala Gly Ala 10 Gly Ala Gly Ala Gly Pro Gly Ala Leu Pro Pro Gln Pro Ala Ala Leu 25 Pro Pro Ala Pro Pro Gln Gly Ser Pro Cys Ala Ala Ala Ala Gly Gly 40 Ser Gly Ala Cys Gly Pro Ala Thr Ala Val Ala Ala Ala Gly Thr Ala 55 Glu Gly Pro Gly Gly Gly Ser Ala Arg Ile Ala Val Lys Lys Ala 75 70 Gln Leu Arg Ser Ala Pro Arg Ala Lys Lys Leu Glu Lys Leu Gly Val 95 85 Tyr Ser Ala Cys Lys Ala Glu Glu Ser Cys Lys Cys Asn Gly Trp Lys 110 105 100 Asn Pro Asn Pro Ser Pro Thr Pro Pro Arg Ala Asp Leu Gln Gln Ile 125 120 Ile Val Ser Leu Thr Glu Ser Cys Arg Ser Cys Ser His Ala Leu Ala



Ala His Val Ser His Leu Glu Asn Val Ser Glu Glu Glu Met Asn Arg 150 155 Leu Leu Gly Ile Val Leu Asp Val Glu Tyr Leu Phe Thr Cys Val His 165 170· Lys Glu Glu Asp Ala Asp Thr Lys Gln Val Tyr Phe Tyr Leu Phe Lys 180 185 Leu Leu Arg Lys Ser Ile Leu Gln Arg Gly Lys Pro Val Val Glu Gly 205 195 200 Ser Leu Glu Lys Lys Pro Pro Phe Glu Lys Pro Ser Ile Glu Gln Gly 215 220 Val Asn Asn Phe Val Gln Tyr Lys Phe Ser His Leu Pro Ala Lys Glu 230 235 Arg Gln Thr Ile Val Glu Leu Ala Lys Met Phe Leu Asn Arg Ile Asn 250 255 245 Tyr Trp His Leu Glu Ala Pro Ser Gln Arg Arg Leu Arg Ser Pro Asn 260 265 Asp Asp Ile Ser Gly Tyr Lys Glu Asn Tyr Thr Arg Trp Leu Cys Tyr 285 280 275 Cys Asn Val Pro Gln Phe Cys Asp Ser Leu Pro Arg Tyr Glu Thr Thr 295 Gln Val Phe Gly Arg Thr Leu Leu Arg Ser Val Phe Thr Val Met Arg 310 315 320 Arg Gln Leu Leu Glu Gln Ala Arg Gln Glu Lys Asp Lys Leu Pro Leu 335 325 Glu Lys Arg Thr Leu Ile Leu Thr His Phe Pro Lys Phe Leu Ser 345 350 340

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 476 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Leu Glu Glu Glu Ile Tyr Gly Ala Asn Ser Pro Ile Trp Glu Ser 10 Gly Phe Thr Met Pro Pro Ser Glu Gly Thr Gln Leu Val Pro Arg Pro 25 Ala Ser Val Ser Ala Ala Val Val Pro Ser Thr Pro Ile Phe Ser Pro 40 45 Ser Met Gly Gly Gly Ser Asn Ser Ser Leu Ser Leu Asp Ser Ala Gly 55 Ala Glu Pro Met Pro Gly Glu Lys Arg Thr Leu Pro Glu Asn Leu Thr 7.5 70 Leu Glu Asp Ala Lys Arg Leu Arg Val Met Gly Asp Ile Pro Met Glu 95 Leu Val Asn Glu Val Met Leu Thr Ile Thr Asp Pro Ala Ala Met Leu 110 105 100 Gly Pro Glu Thr Ser Leu Leu Ser Ala Asn Ala Ala Arg Asp Glu Thr 120 125 Ala Arg Leu Glu Glu Arg Arg Gly Ile Ile Glu Phe His Val Ile Gly 135 140 Asn Ser Leu Thr Pro Lys Ala Asn Arg Arg Val Leu Leu Trp Leu Val 150 155 Gly Leu Gln Asn Val Phe Ser His Gln Leu Pro Arg Met Pro Lys Glu 165 170 175 Tyr Ile Ala Arg Leu Val Phe Asp Pro Lys His Lys Thr Leu Ala Leu 185



Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe Pro 200 195 Thr Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val Thr Ser Asn Glu 220 215 Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His Leu Lys Glu Tyr 235 -230 His Ile Lys His Asn Ile Leu Tyr Phe Leu Thr Tyr Ala Asp Glu Tyr 250 245 Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Asp Ile Lys Val 260 265 Pro Lys Ser Arg Tyr Leu Gly Tyr Ile Lys Asp Tyr Glu Gly Ala Thr 280 275 Leu Met Glu Cys Glu Leu Asn Pro Arg Ile Pro Tyr Thr Glu Leu Ser 300 ,295 His Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu Arg . 315 310 Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe Lys 330 335 325 Glu Gly Val Arg Gln Ile Pro Val Glu Ser Val Pro Gly Ile Arg Glu 345 Thr Gly Trp Lys Pro Leu Gly Lys Glu Lys Gly Lys Glu Leu Lys Asp 355 360 Pro Asp Gln Leu Tyr Thr Thr Leu Lys Asn Leu Leu Ala Gln Ile Lys 380 375 370 Ser His Pro Ser Ala Trp Pro Phe Met Glu Pro Val Lys Lys Ser Glu 395 390 Ala Pro Asp Tyr Tyr Glu Val Ile Arg Phe Pro Ile Asp Leu Lys Thr 410 405 Met Thr Glu Arg Leu Arg Ser Arg Tyr Tyr Val Thr Arg Lys Leu Phe 425 420 Val Ala Asp Leu Gln Arg Val Ile Ala Asn Cys Arg Glu Tyr Asn Pro 440 435 Pro Asp Ser Glu Tyr Cys Arg Cys Ala Ser Ala Leu Glu Lys Phe Phe 455 450 Tyr Phe Lys Leu Lys Glu Gly Gly Leu Ile Asp Lys 470

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2414 amino acids
- . (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

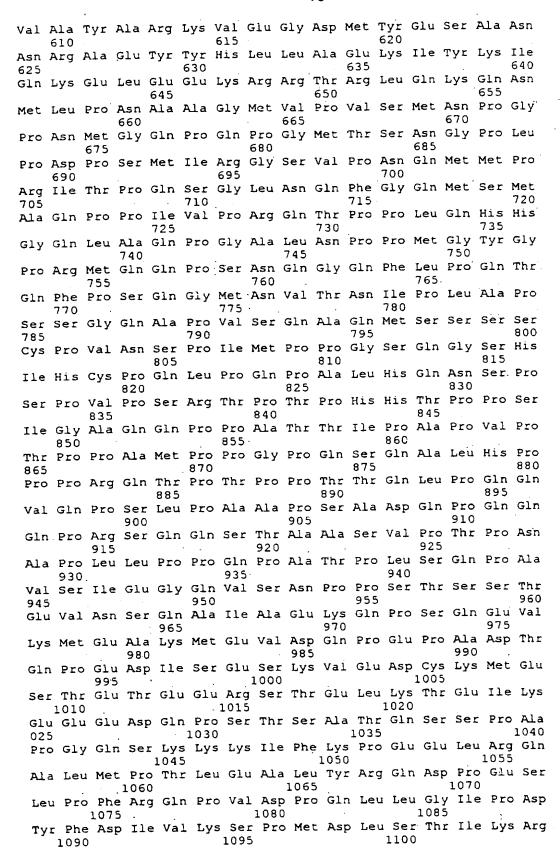
 Met
 Ala
 Glu
 Asn
 Val
 Val
 Glu
 Pro
 Gly
 Pro
 Pro
 Ser
 Ala
 Lys
 Arg
 Pro
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 Arg
 Pro
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 Inches





Pro Gly Leu Gly Leu Ile Asn Ser Met Val Lys Ser Pro Met Thr Gln Ala Gly Leu Thr Ser Pro Asn Met Gly Met Gly Thr Ser Gly Pro Asn Gln Gly Pro Thr Gln Ser Thr Gly Met Met Asn Ser Pro Val Asn Gln Pro Ala Met Gly Met Asn Thr Gly Thr Asn Ala Gly Met Asn Pro Gly Met Leu Ala Ala Gly Asn Gly Gln Gly Ile Met Pro Asn Gln Val Met Asn Gly Ser Ile Gly Ala Gly Arg Gly Arg Gln Asp Met Gln Tyr Pro Asn Pro Gly Met Gly Ser Ala Gly Asn Leu Leu Thr Glu Pro Leu Gln Gln Gly Ser Pro Gln Met Gly Gly Gln Thr Gly Leu Arg Gly Pro Gln Pro Leu Lys Met Gly Met Met Asn Asn Pro Asn Pro Tyr Gly Ser Pro Tyr Thr Gln Asn Pro Gly Gln Gln Ile Gly Ala Ser Gly Leu Gly Leu Gln Ile Gln Thr Lys Thr Val Leu Ser Asn Asn Leu Ser Pro Phe Ala 280 285 Met Asp Lys Lys Ala Val Pro Gly Gly Gly Met Pro Asn Met Gly Gln Gln Pro Ala Pro Gln Val Gln Gln Pro Gly Leu Val Thr Pro Val Ala Gln Gly Met Gly Ser Gly Ala His Thr Ala Asp Pro Glu Lys Arg Lys Leu Ile Gln Gln Gln Leu Val Leu Leu His Ala His Lys Cys Gln Arg Arg Glu Gln Ala Asn Gly Glu Val Arg Gln Cys Asn Leu Pro His Cys Arg Thr Met Lys Asn Val Leu Asn His Met Thr His Cys Gln Ser Gly Lys Ser Cys Gln Val Ala His Cys Ala Ser Ser Arg Gln Ile Ile Ser His Trp Lys Asn Cys Thr Arg His Asp Cys Pro Val Cys Leu Pro Leu Lys Asn Ala Gly Asp Lys Arg Asn Gln Gln Pro Ile Leu Thr Gly Ala Pro Val Gly Leu Gly Asn Pro Ser Ser Leu Gly Val Giy Gln Gln Ser Ala Pro Asn Leu Ser Thr Val Ser Gln Ile Asp Pro Ser Ser Ile 4.60 Glu Arg Ala Tyr Ala Ala Leu Gly Leu Pro Tyr Gln Val Asn Gln Met Pro Thr Gln Pro Gln Val Gln Ala Lys Asn Gln Gln Asn Gln Gln Pro Gly Gln Ser Pro Gln Gly Met Arg Pro Met Ser Asn Met Ser Ala Ser Pro Met Gly Val Asn Gly Gly Val Gly Val Gln Thr Pro Ser Leu Leu Ser Asp Ser Met Leu His Ser Ala Ile Asn Ser Gln Asn Pro Met Met Ser Glu Asn Ala Ser Val Pro Ser Leu Gly Pro Met Pro Thr Ala Ala . 550 Gln Pro Ser Thr Thr Gly Ile Arg Lys Gln Trp His Glu Asp Ile Thr Gln Asp Leu Arg Asn His Leu Val His Lys Leu Val Gln Ala Ile Phe Pro Thr Pro Asp Pro Ala Ala Leu Lys Asp Arg Arg Met Glu Asn Leu



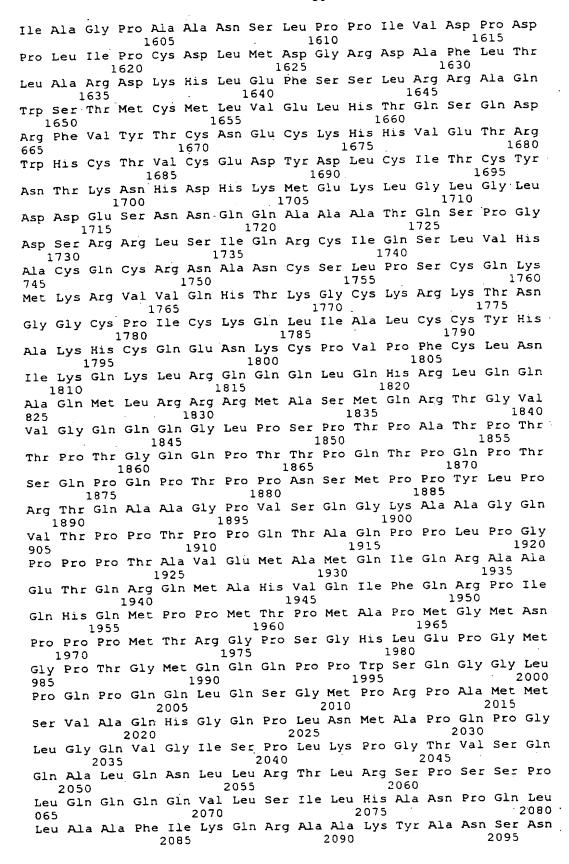






Lys Leu Asp Thr Gly Gln Tyr Gln Glu Pro Trp Gln Tyr Val Asp Asp 1110 1115 105 Ile Trp Leu Met Phe Asn Asn Ala Trp Leu Tyr Asn Arg Lys Thr Ser 1125 1130 - 1135 Arg Val Tyr Lys Tyr Cys Ser Lys Leu Ser Glu Val Phe Glu Glu Glu 1140 1145 1150 Ile Asp Pro Val Met Gln Ser Leu Gly Tyr Cys Cys Gly Arg Lys Leu 1155 1160 1165 Glu Phe Ser Pro Gln Thr Leu Cys Cys Tyr Gly Lys Gln Leu Cys Thr 1175 1180 Ile Pro Arg Asp Ala Thr Tyr Tyr Ser Tyr Gln Asn Arg Tyr His Phe 1190 1195 Cys Glu Lys Cys Phe Asn Glu Ile Gln Gly Glu Ser Val Ser Leu Gly 1210 1215 1205 Asp Asp Pro Ser Gln Pro Gln Thr Thr Ile Asn Lys Glu Gln Phe Ser 1220 1225 1230 Lys Arg Lys Asn Asp Thr Leu Asp Pro Glu Leu Phe Val Glu Cys Thr 1235 1240 1245 Glu Cys Gly Arg Lys Met His Gln Ile Cys Val Leu His His Glu Ile 1250 1255 1260 Ile Trp Pro Ala Gly Phe Val Cys Asp Gly Cys Leu Lys Lys Ser Ala 265 1270 1275 1280 Arg Thr Arg Lys Glu Asn Lys Phe Ser Ala Lys Arg Leu Pro Ser Thr 1285 1290 1295 Arg Leu Gly Thr Phe Leu Glu Asn Arg Val Asn Asp Phe Leu Arg Arg 1300 1305 1310 Gln Asn His Pro Glu Ser Gly Glu Val Thr Val Arg Val Val His Ala 1320 1325 1315 Ser Asp Lys Thr Val Glu Val Lys Pro Gly Met Lys Ala Arg Phe Val 1335 1340 Asp Ser Gly Glu Met Ala Glu Ser Phe Pro Tyr Arg Thr Lys Ala Leu 1350 1355 Phe Ala Phe Glu Glu Ile Asp Gly Val Asp Leu Cys Phe Phe Gly Met 1370 1375 1365 His Val Gln Glu Tyr Gly Ser Asp Cys Pro Pro Pro Asn Gln Arg Arg 1380 1385 1390 Val Tyr Ile Ser Tyr Leu Asp Ser Val His Phe Phe Arg Pro Lys Cys 1395 1400 1405 Leu Arg Thr Ala Val Tyr His Glu Ile Leu Ile Gly Tyr Leu Glu Tyr 1410 1415 1420 Val Lys Lys Leu Gly Tyr Thr Thr Gly His Ile Trp Ala Cys Pro Pro 1435 1430 Ser Glu Gly Asp Asp Tyr Ile Phe His Cys His Pro Pro Asp Gln Lys 1450 1445 Ile Pro Lys Pro Lys Arg Leu Gln Glu Trp Tyr Lys Lys Met Leu Asp 1465 Lys Ala Val Ser Glu Arg Ile Val His Asp Tyr Lys Asp Ile Phe Lys 1480 1485 1475 Gln Ala Thr Glu Asp Arg Leu Thr Ser Ala Lys Glu Leu Pro Tyr Phe 1490 1495 1500 Glu Gly Asp Phe Trp Pro Asn Val Leu Glu Glu Ser Ile Lys Glu Leu 1510 1515 1520 505 Glu Gln Glu Glu Glu Arg Lys Arg Glu Glu Asn Thr Ser Asn Glu 1525 1530 1535 Ser Thr Asp Val Thr Lys Gly Asp Ser Lys Asn Ala Lys Lys Lys Asn 1540 1545 1550 Asn Lys Lys Thr Ser Lys Asn Lys Ser Ser Leu Ser Arg Gly Asn Lys 1555 1560 1565 Lys Lys Pro Gly Met Pro Asn Val Ser Asn Asp Leu Ser Gln Lys Leu 1570 1575 1580 Tyr Ala Thr Met Glu Lys His Lys Glu Val Phe Phe Val Ile Arg Leu 1590 1595 ...





Pro Gln Pro Ile Pro Gly Gln Pro Gly Met Pro Gln Gly Gln Pro Gly 2110 2105 Leu Gln Pro Pro Thr Met Pro Gly Gln Gln Gly Val His Ser Asn Pro 2120 2125 2115 Ala Met Gln Asn Met Asn Pro Met Gln Ala Gly Val Gln Arg Ala Gly 2140 2135 Leu Pro Gln Gln Gln Pro Gln Gln Gln Leu Gln Pro Pro Met Gly Gly 2155 2160 2150 Met Ser Pro Gln Ala Gln Gln Met Asn Met Asn His Asn Thr Met Pro 2165 2170 Ser Gln Phe Arg Asp Ile Leu Arg Arg Gln Gln Met Met Gln Gln Gln 2185 2190 Gln Gln Gly Ala Gly Pro Gly Ile Gly Pro Gly Met Ala Asn His 2205 2195 2200 Asn Gln Phe Gln Gln Pro Gln Gly Val Gly Tyr Pro Pro Gln Pro Gln 2210 2215 2220 Gln Arg Met Gln His His Met Gln Gln Met Gln Gly Asn Met Gly 2230 2235 Gln Ile Gly Gln Leu Pro Gln Ala Leu Gly Ala Glu Ala Gly Ala Ser 2245 2250 Leu Gln Ala Tyr Gln Gln Arg Leu Leu Gln Gln Met Gly Ser Pro 2260 2265 2270 Val Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu Pro Asn Gln 2275 2280 2285 Ala Gln Ser Pro His Leu Gln Gly Gln Gln Ile Pro Asn Ser Leu Ser 2290 2295 2300 Asn Gln Val Arg Ser Pro Gln Pro Val Pro Ser Pro Arg Pro Gln Ser 2310 2315 Gln Pro Pro His Ser Ser Pro Ser Pro Arg Met Gln Pro Gln Pro Ser 2325 2330 Pro His His Val Ser Pro Gln Thr Ser Ser Pro His Pro Gly Leu Val 2350 2345 Ala Ala Gln Ala Asn Pro Met Glu Gln Gly His Phe Ala Ser Pro Asp 2360 2365 2355 Gln Asn Ser Met Leu Ser Gln Leu Ala Ser Asn Pro Gly Met Ala Asn 2375 2380 Leu His Gly Ala Ser Ala Thr Asp Leu Gly Leu Ser Thr Asp Asn Ser 2390 2395 Asp Leu Asn Ser Asn Leu Ser Gln Ser Thr Leu Asp Ile His 2410

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2441 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (11) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

 Met
 Ala
 Glu
 Asn
 Leu
 Leu
 Asp
 Gly
 Pro
 Pro
 Asn
 Pro
 Lys
 Arg
 Ala
 Lys

 Leu
 Ser
 Ser
 Pro
 Gly
 Phe
 Ser
 Ala
 Asn
 Asp
 Asn
 Thr
 Asp
 Phe
 Gly
 Ser

 Leu
 Phe
 Asp
 Leu
 Pro
 Asp
 Glu
 Leu
 Ile
 Pro
 Asn
 Gly

 Glu
 Leu
 Ser
 Leu
 Asn
 Ser
 Gly
 Asn
 Leu
 Val
 Pro
 Asp
 Ala
 Ala
 Ser

 Glu
 Leu
 Ser
 Leu
 Leu
 Leu
 Leu
 Leu
 Pro
 Asp
 Ala
 Ala
 Ser

 Glu
 Leu
 Ser
 Gly
 Asn
 Leu
 Leu
 Val
 Pro
 Asp
 Ala
 Ala
 Ser

 Lys
 His
 Lys
 Gln
 Leu
 Ser
 Glu
 Leu
 Leu
 Leu
 Arg
 Gly
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Ile	Asn	Pro	Gly	Ile 85	Gly	Asn	Val	Ser	Ala 90	Ser	Ser	Pro	Val	Gln (95	Gln
_			Gly 100					105					110		
		115	Ala				120					125			
	130	Asn	Leu			135					140				
1/15	Ala		Gln		150					155					100
Val			Ser	165					170					1/3	
			Phe 180					185					190		
-		195	Leu				200					205			
	210		Leu			215					220				
225			Pro		230					235					240
			Gln	245					250					255	
			Ala 260					265					2/0		
		275	Gly				280					285			
	290	_	Val			295					300				
305			Ala Met		310					315					320
			Ala	325					330					335	
			340 1 Gln					345					350		
		355					360					365			
	370)	. Lys			375					380				
385			s Gln		390					395)				400
-			s Asr	405)				410)				415	
	_		420 a Sei)				425	•				430		
-		43					440)				445			
	45	o .	r Sei			455	5			o Ile	460 Asp	}			Met
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Th	r Gl	n Le	u Gli	489 n Pro	5 o Glr	n Val	l Pro				n Pro	Alā	Glr	495 Pro	
Al	a Hi	s Gl	50 n Gl:	0 n Me	t Ar	g Th.				a Le	u Gly	Asr	510 Asr		Met
Se	r Va	51 1 Pr	.5 :0 Al	a Gl	y Gl	y Il	520 e T h:) r Th	r As	p Gl	n Gl	525 n Pro		Asr	Leu
	e Se	0 . r Gl	u Se	r Al				r Se	r Le	u Gl	540 y Ala		r Ası	n Pro	Leu 560
5 4 Me	5 t As	n As	p Gl			n Se	r Gl	y As	n Il 57	55 e Gl 0		r Le	u Se	r Thi	: Ile
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Pro Thr Ala Ala Pro Pro Ser Ser Thr Gly Val Arg Lys Gly Trp His Glu His Val Thr Gln Asp Leu Arg Ser His Leu Val His Lys Leu Val Gln Ala Ile Phe Pro Thr Pro Asp Pro Ala Ala Leu Lys Asp Arg Arg Met Glu Asn Leu Val Ala Tyr Ala Lys Lys Val Glu Gly Asp Met Tyr . 640 Glu Ser Ala Asn Ser Arg Asp Glu Tyr Tyr His Leu Leu Ala Glu Lys Ile Tyr Lys Ile Gln Lys Glu Leu Glu Glu Lys Arg Arg Thr Arg Leu His Lys Gln Gly Ile Leu Gly Asn Gln Pro Ala Leu Pro Ala Ser Gly Ala Gln Pro Pro Val Ile Pro Pro Ala Gln Ser Val Arg Pro Pro Asn Gly Pro Leu Pro Leu Pro Val Asn Arg Met Gln Val Ser Gln Gly Met Asn Ser Phe Asn Pro Met Ser Leu Gly Asn Val Gln Leu Pro Gln Ala Pro Met Gly Pro Arg Ala Ala Ser Pro Met Asn His Ser Val Gln Met Asn Ser Met Ala Ser Val Pro Gly Met Ala Ile Ser Pro Ser Arg Met Pro Gln Pro Pro Asn Met Met Gly Thr His Ala Asn Asn Ile Met Ala Gln Ala Pro Thr Gln Asn Gln Phe Leu Pro Gln Asn Gln Phe Pro Ser Ser Ser Gly Ala Met Ser Val Asn Ser Val Gly Met Gly Gln Pro Ala Ala Gln Ala Gly Val Ser Gln Gly Gln Glu Pro Gly Ala Ala Leu Pro Asn Pro Leu Asn Met Leu Ala Pro Gln Ala Ser Gln Leu Pro Cys Pro Pro Val Thr Gln Ser Pro Leu His Pro Thr Pro Pro Pro Ala Ser Thr Ala Ala Gly Met Pro Ser Leu Gln His Pro Thr Ala Pro Gly Met Thr Pro Pro Gln Pro Ala Ala Pro Thr Gln Pro Ser Thr Pro Val Ser Ser Gly Gln Thr Pro Thr Pro Gly Ser Val Pro Ser Ala Ala Gln Thr Gln Ser Thr Pro Thr Val Gln Ala Ala Ala Gln Ala Gln Val Thr Pro Gln Pro Gln Thr Pro Val Gln Pro Pro Ser Val Ala Thr Pro Gln Ser Ser Gln Gln Pro Thr Pro Val His Thr Gln Pro Pro Gly Thr Pro Leu Ser Gln Ala Ala Ala Ser Ile Asp Asn Arg Val Pro Thr Pro Ser Thr Val Thr Ser Ala Glu Thr Ser Ser Gln Gln Pro Gly Pro Asp 98C Val Pro Met Leu Glu Met Lys Thr Glu Val Gln Thr Asp Asp Ala Glu 1000 1005 Pro Glu Pro Thr Glu Ser Lys Gly Glu Pro Arg Ser Glu Met Met Glu Glu Asp Leu Gln Gly Ser Ser Gln Val Lys Glu Glu Thr Asp Thr Thr 1030 1035 1040 Glu Gln Lys Ser Glu Pro Met Glu Val Glu Glu Lys Lys Pro Glu Val 1050 1055 Lys Val Glu Ala Lys Glu Glu Glu Asn Ser Ser Asn Asp Thr Ala





Ser Gln Ser Thr Ser Pro Ser Gln Pro Arg Lys Lys Ile Phe Lys Pro 1075 1080 1085 Glu Glu Leu Arg Gln Ala Leu Met Pro Thr Leu Glu Ala Leu Tyr Arg 1090 1095 1100 Gln Asp Pro Glu Ser Leu Pro Phe Arg Gln Pro Val Asp Pro Gln Leu 1110 1115 Leu Gly Ile Pro Asp Tyr Phe Asp Ile Val Lys Asn Pro Met Asp Leu 1135 1125 1130 Ser Thr Ile Lys Arg Lys Leu Asp Thr Gly Gln Tyr Gln Glu Pro Trp 1140 1145 1150 Gln Tyr Val Asp Asp Val Arg Leu Met Phe Asn Asn Ala Trp Leu Tyr 1155 1160 1165 Asn Arg Lys Thr Ser Arg Val Tyr Lys Phe Cys Ser Lys Leu Ala Glu 1170 1175 1180 Val Phe Glu Gln Glu Ile Asp Pro Val Met Gln Ser Leu Gly Tyr Cys 185 1190 1195 Cys Gly Arg Lys Tyr Glu Phe Ser Pro Gln Thr Leu Cys Cys Tyr Gly 1205 1210 1215 Lys Gln Leu Cys Thr Ile Pro Arg Asp Ala Ala Tyr Tyr Ser Tyr Gln 1220 1230 Asn Arg Tyr His Phe Cys Gly Lys Cys Phe Thr Glu Ile Gln Gly Glu 1235 1240 1245 Asn Val Thr Leu Gly Asp Asp Pro Ser Gln Pro Gln Thr Thr Ile Ser 1255 1260 Lys Asp Gln Phe Glu Lys Lys Lys Asn Asp Thr Leu Asp Pro Glu Pro 265 1270 1275 Phe Val Asp Cys Lys Glu Cys Gly Arg Lys Met His Gln Ile Cys Val 1285 1290 1295 Leu His Tyr Asp Ile Ile Trp Pro Ser Gly Phe Val Cys Asp Asn Cys 1300 1305 Leu Lys Lys Thr Gly Arg Pro Arg Lys Glu Asn Lys Phe Ser Ala Lys 1315 1320 1325 Arg Leu Gln Thr Thr Arg Leu Gly Asn His Leu Glu Asp Arg Val Asn 1330 1335 1340 Lys Phe Leu Arg Arg Gln Asn His Pro Glu Ala Gly Glu Val Phe Val 1350 1355 1360 Arg Val Val Ala Ser Ser Asp Lys Thr Val Glu Val Lys Pro Gly Met 1365 1370 1375 Lys Ser Arg Phe Val Asp Ser Gly Glu Met Ser Glu Ser Phe Pro Tyr 1380 1385 Arg Thr Lys Ala Leu Phe Ala Phe Glu Glu Ile Asp Gly Val Asp Val 1395 . 1400 1405 Cys Phe Phe Gly Met His Val Gln Asp Thr Ala Leu Ile Ala Pro His 1415 1420 Gln Ile Gln Gly Cys Val Tyr Ile Ser Tyr Leu Asp Ser Ile His Phe 1435 1440 1430 Phe Arg Pro Arg Cys Leu Arg Thr Ala Val Tyr His Glu Ile Leu Ile 1445 1450 1455 Gly Tyr Leu Glu Tyr Val Lys Lys Leu Val Tyr Val Thr Ala His Ile 1465 1470 1460 Trp Ala Cys Pro Pro Ser Glu Gly Asp Asp Tyr Ile Phe His Cys His 1475 1480 1485 Pro Pro Asp Gln Lys Ile Pro Lys Pro Lys Arg Leu Gln Glu Trp Tyr 1490 1495 1500 Lys Lys Met Leu Asp Lys Ala Phe Ala Glu Arg Ile Ile Asn Asp Tyr 1515 505 1510 Lys Asp Ile Phe Lys Gln Ala Asn Glu Asp Arg Leu Thr Ser Ala Lys 1530 1535 1525 Glu Leu Pro Tyr Phe Glu Gly Asp Phe Trp Pro Asn Val Leu Glu Glu 1540 1545 1550 Ser Ile Lys Glu Leu Glu Gln Glu Glu Glu Glu Arg Lys Lys Glu Glu 1560 1555





Ser Thr Ala Ala Ser Glu Thr Pro Glu Gly Ser Gln Gly Asp Ser Lys 1575 1580 Asn Ala Lys Lys Lys Asn Asn Lys Lys Thr Asn Lys Asn Lys Ser Ser 1590 1595 1600 Ile Ser Arg Ala Asn Lys Lys Lys Pro Ser Met Pro Asn Val Ser Asn 1610 1615 1605 Asp Leu Ser Gln Lys Leu Tyr Ala Thr Met Glu Lys His Lys Glu Val 1620 1625 1630 Phe Phe Val Ile His Leu His Ala Gly Pro Val Ile Ser Thr Gln Pro 1640 1645 Pro Ile Val Asp Pro Asp Pro Leu Leu Ser Cys Asp Leu Met Asp Gly 1655 1660 Arg Asp Ala Phe Leu Thr Leu Ala Arg Asp Lys His Trp Glu Phe Ser 1670 1675 1680 Ser Leu Arg Arg Ser Lys Trp Ser Thr Leu Cys Met Leu Val Glu Leu 1685 1690 1695 His Thr Gln Gly Gln Asp Arg Phe Val Tyr Thr Cys Asn Glu Cys Lys 1700 1705 1710 His His Val Glu Thr Arg Trp His Cys Thr Val Cys Glu Asp Tyr Asp 1715 1720 1725 Leu Cys Ile Asn Cys Tyr Asn Thr Lys Ser His Thr His Lys Met Val 1730 1735 1740 Lys Trp Gly Leu Gly Leu Asp Asp Glu Gly Ser Ser Gln Gly Glu Pro 1750 1755 1760 745 Gln Ser Lys Ser Pro Gln Glu Ser Arg Arg Leu Ser Ile Gln Arg Cys 1765 1770 1775 Ile Gln Ser Leu Val His Ala Cys Gln Cys Arg Asn Ala Asn Cys Ser 1780 1785 1790 Leu Pro Ser Cys Gln Lys Met Lys Arg Val Val Gln His Thr Lys Gly 1800 1805 Cys Lys Arg Lys Thr Asn Gly Gly Cys Pro Val Cys Lys Gln Leu Ile 1820 1815 Ala Leu Cys Cys Tyr His Ala Lys His Cys Gln Glu Asn Lys Cys Pro 1830 1835 Val Pro Phe Cys Leu Asn Ile Lys His Asn Val Arg Gln Gln Gln Ile 1850 • 1855 1845 Gln His Cys Leu Gln Gln Ala Gln Leu Met Arg Arg Arg Met Ala Thr 1865 1870 1860 Met Asn Thr Arg Asn Val Pro Gln Gln Ser Leu Pro Ser Pro Thr Ser 1880 1885 Ala Pro Pro Gly Thr Pro Thr Gln Gln Pro Ser Thr Pro Gln Thr Pro 1895 1900 Gln Pro Pro Ala Gln Pro Gln Pro Ser Pro Val Asn Met Ser Pro Ala 1910 1915 Gly Phe Pro Asn Val Ala Arg Thr Gln Pro Pro Thr Ile Val Ser Ala 1930 1925 Gly Lys Pro Thr Asn Gln Val Pro Ala Pro Pro Pro Pro Ala Gln Pro 1945 1950 Pro Pro Ala Ala Val Glu Ala Ala Arg Gln Ile Glu Arg Glu Ala Gln 1960 1965 . 1955 Gln Gln Gln His Leu Tyr Arg Ala Asn Ile Asn Asn Gly Met Pro Pro 1975 1970 1980 Gly Arg Asp Gly Met Gly Thr Pro Gly Ser Gln Met Thr Pro Val Gly 1990 1995 2000 Leu Asn Val Pro Arg Pro Asn Gln Val Ser Gly Pro Val Met Ser Ser 2010 2015 2005 Met Pro Pro Gly Gln Trp Gln Gln Ala Pro Ile Pro Gln Gln Gln Pro 2020 2025 2030 Met Pro Gly Met Pro Arg Pro Val Met Ser Met Gln Ala Gln Ala Ala 2040 2045 Val Ala Gly Pro Arg Met Pro Asn Val Gln Pro Asn Arg Ser Ile Ser 2055 -2060





Pro Ser Ala Leu Gln Asp Leu Leu Arg Thr Leu Lys Ser Pro Ser Ser 2070 2075 2080 065 Pro Gln Gln Gln Gln Val Leu Asn Ile Leu Lys Ser Asn Pro Gln 2095 2085 2090 Leu Met Ala Ala Phe Ile Lys Gln Arg Thr Ala Lys Tyr Val Ala Asn 2100 2105 2110 Gln Pro Gly Met Gln Pro Gln Pro Gly Leu Gln Ser Gln Pro Gly Met 2115 2120 2125 Gln Pro Gln Pro Gly Met His Gln Gln Pro Ser Leu Gln Asn Leu Asn 2130 2135 2140 Ala Met Gln Ala Gly Val Pro Arg Pro Gly Val Pro Pro Pro Gln Pro 145 2150 2155 2160 Ala Met Gly Gly Leu Asn Pro Gln Gly Gln Ala Leu Asn Ile Met Asn 2165 2170 2175 Pro Gly His Asn Pro Asn Met Thr Asn Met Asn Pro Gln Tyr Arg Glu 2180 2185 2190 Met Val Arg Arg Gln Leu Leu Gln His Gln Gln Gln Gln Gln Gln 2195 2200 2205 Gln Gln Gln Gln Gln Gln Gln Asn Ser Ala Ser Leu Ala Gly Gly 2210 2215 2220 Met Ala Gly His Ser Gln Phe Gln Gln Pro Gln Gly Pro Gly Gly Tyr 225 2230 2235 Ala Pro Ala Met Gln Gln Gln Arg Met Gln Gln His Leu Pro Ile Gln 2245 2250 2255 Gly Ser Ser Met Gly Gln Met Ala Ala Pro Met Gly Gln Leu Gly Gln 2265 2270 2260 Met Gly Gln Pro Gly Leu Gly Ala Asp Ser Thr Pro Asn Ile Gln Gln 2275 2280 2285 Ala Leu Gln Gln Arg Ile Leu Gln Gln Gln Gln Met Lys Gln Gln Ile 2290 2295 2300 Gly Ser Pro Gly Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu 305 2310 2315 2320 Ser Gly Gln Pro Gln Ala Ser His Leu Pro Gly Gln Gln Ile Ala Thr 2325 2330 2335 Ser Leu Ser Asn Gln Val Arg Ser Pro Ala Pro Val Gln Ser Pro Arg 2340 2345 2350 Pro Gln Ser Gln Pro Pro His Ser Ser Pro Ser Pro Arg Ile Gln Pro 2355 2360 2365 Gln Pro Ser Pro His His Val Ser Pro Gln Thr Gly Thr Pro His Pro 2375 2380 Gly Leu Ala Val Thr Met Ala Ser Ser Met Asp Gln Gly His Leu Gly 385 2390 2395 2400 Asn Pro Glu Gln Ser Ala Met Leu Pro Gln Leu Asn Thr Pro Asn Arg 2405 2410 2415 Ser Ala Leu Ser Ser Glu Leu Ser Leu Val Gly Asp Thr Thr Gly Asp 2430 2420 . 2425 Thr Leu Glu Lys Phe Val Glu Gly Leu 2435 2440

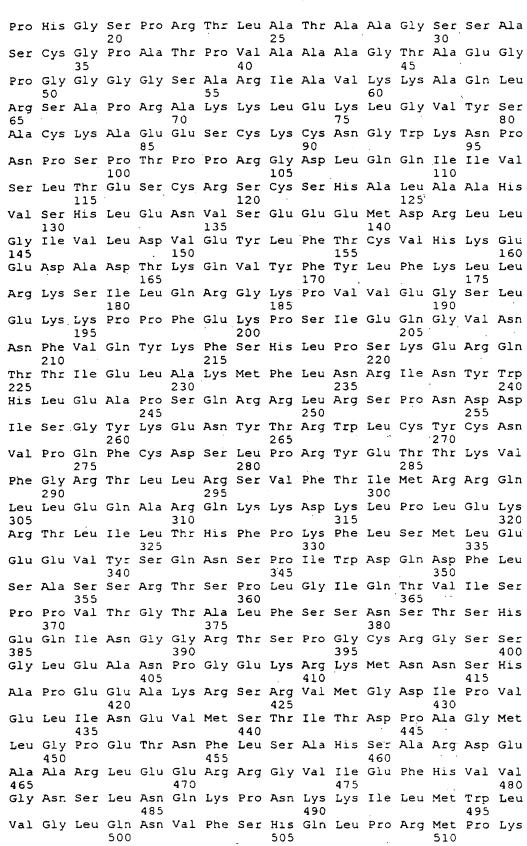
86

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 813 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear ...
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ala Glu Ala Gly Gly Ala Gly Ser Pro Ala Leu Pro Pro Ala Pro 1 5 10 15





Glu Tyr Ile Thr Arg Leu Val Phe Asp Pro Lys His Lys Thr Leu Ala 520 525 Leu Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe 540 535 Pro Ser Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val Thr Ser Asn 550 555 Glu Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His Leu Lys Glu 570 565 Tyr His Ile Lys His Glu Ile Leu Asn Phe Leu Thr Tyr Ala Asp Glu 590 . 580 : 585 Tyr Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Glu Ile Lys 600 Ile Pro Lys Thr Lys Tyr Val Gly Tyr Ile Lys Asp Tyr Glu Gly Ala . 615 620 . Thr Leu Met Gly Cys Glu Leu Asn Pro Gln Ile Pro Tyr Thr Glu Phe 635 630 Ser Val Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu 650 645 Arg Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe 670 665 660 Lys Asp Gly Val Arg Gln Ile Pro Ile Glu Ser Ile Pro Gly Ile Arg 680 675 Glu Thr Gly Trp Lys Pro Ser Gly Lys Glu Lys Ser Lys Glu Pro Lys 695 700 Asp Pro Glu His Val Tyr Ser Thr Leu Lys Asn Ile Leu Gln Gln Val 715 710 Lys Asn His Pro Asn Ala Trp Pro Phe Met Glu Pro Val Lys Arg Thr 735 730 725 Glu Ala Pro Gly Tyr Tyr Glu Val Ile Arg Phe Pro Met Asp Leu Lys 750 745 740 Thr Met Ser Glu Arg Leu Arg Asn Arg Tyr Tyr Val Ser Lys Lys Leu 765. 760 Phe Met Ala Asp Leu Gln Arg Val Phe Thr Asn Cys Lys Glu Tyr Asn 780 775 Pro Pro Glu Ser Glu Tyr Tyr Lys Cys Ala Ser Ile Leu Glu Lys Phe 795 790 Phe Phe Ser Lys Ile Lys Glu Ala Gly Leu Ile Asp Lys 805

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 50 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

His Thr Lys Gly Cys Lys Arg Lys Thr Asn Gly Gly Cys Pro Val Cys 1 10 15

Lys Gln Leu Ile Ala Leu Cys Cys Tyr His Ala Lys His Cys Gln Glu 20 25 30.

Asn Lys Cys Pro Val Pro Phe Cys Leu Asn Ile Lys His Asn Val Arg 35 40 45

- (2) INFORMATION FOR SEQ ID NO:10:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2204 base pairs





(B) TYPE: nucleic acid (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ACCCACTCCC CCCAGAGCCG ACCTGCAGCA AATAATTGTC AGTCTAACAG AATCCTGTCG . 60 GAGTTGTAGC CATGCCCTAG CTGCTCATGT TTCCCACCTG GAGAATGTGT CAGAGGAAGA 120 AATGAACAGA CTCCTGGGAA TAGTATTGGA TGTGGAATAT CTCTTTACCT GTGTCCACAA 180 GGAAGAAGAT GCAGATACCA AACAAGTTTA TTTCTATCTA TTTAAGCTCT TGAGAAAGTC 240 TATTTTACAA AGAGGAAAAC CTGTGGTTGG AAGGCTCTTT GGAAAAGAAA CCCCCATTTG 300 AAAAACCTAG CATTGAACAG GGTGTGAATA ACTTTGTGCA GTACAAATTT AGTCACCTGC 360 CAGCAAAAAG AAAGGCAAAC CAATAGTTGA GTTGGCAAAA ATGTTCCTAA ACCGCATCAC CTATTGGCAT CTGGAGGCAC CATCTCAACG AGACTGCGAT CTCCAATGAT GATATTCTGG ATACAAAGAG AACTACACAA GGTGGCTGTG TTACTGCAAC GTGCCACAGT TCTGCGACAG TCTACCTCGG TACGAAACCA CACAGGTGTT TGGGAGAACA TCGTTCGCTC GGTCTTCACT 600 GTTATGAGGC GACAACTCCT GGAACAAGCA AGACAGGAAA AAGATAAACT GCCTCTTGAA
AAACGAACTC TAATCCTCAC TCATTTCCCA AAATTTCTGT CCATGCTAGA AGAAGAAGTA
TATAGTCAAA ACTCTCCCAT CTGGGATCAC CATTTTCTCT CAGCCTCTTC CAGAACCAGC 720 780 CAGCTAGGCA TCCAAACAGT TATCAATCAC CTCCTGTGGC TGGGACAATT TCATACAATT 840 CAACCTCATC TTCCCTTGAG CAGCCAAACG CAGGGAGCAG CAGTCCTGCC TGCAAAGCCT 900 CTTCTGGACT TGAGGCAAAC CCAGGAGAAA AGAGGAAAAT GACTGATTCT CATGTTCTGG AGGAGGCCAA GAAACCCCGA GTTATGGGGG ATATTCCGAT GGAATTAATC AACGAGGTTA 1020 TGTCTACCAT CACGGACCCT GCAGCAATGC TTGGACCAGA GACCAATTTT CTGTCAGCAC 1080 ACTCGGCCAG GGATGAGGCG GCAAGGTTGG AAGAGCGCAG GGGTGTAATT GAATTTCACG 1140 TEGTTEGCAA TTCCCTCAAC CAGAAACCAA ACAAGAAGAT CCTGATETGG CTGGTTGGCC 1200 TACAGAACGT TTTCTCCCAC CAGCTGCCCC GAATGCCAAA AGAATACATC ACACGGCTCG TCTTTGACCC GAAACACAAA ACCCTTGCTT TAATTAAAGA TGGCCGTGTT ATTGGTGGTA 1320 TCTGTTTCCG TATGTTCCCA TCTCAAGGAT TCACAGAGAT TGTCTTCTGT GCTGTAACCT 1380 CAAATGAGCA AGTCAAGGGC TATGGAACAC ACCTGATGAA TCATTTGAAA GAATATCACA 1440 TAAAGCATGA CATCCTGAAC TTCCTCACAT ATGCAGATGA ATATGCAATT GGATACTTTA 1500 AGAAACAGGG TTTCTCCAAA GAAATTAAAA TACCTAAAAC CAAATATGTT GGCTATATCA 1560 AGGATTATGA AGGAGCCACT TTAATGGGAT GTGAGCTAAA TCCACGGATC CCGTACACAG 1620 AATTTTCTGT CATCATTAAA AAGCAGAAGG AGATAATTAA AAAACTGATT GAAAGAAAAC 1680 AGGCACAAAT TCGAAAAGTT TACCCTGGAC TTTCATGTTT TAAAGATGGA GTTCGACAGA 1740 TTCCTATAGA AAGCATTCCT GGAATTAGAG AGACAGGCTG GAAACCGAGT GGAAAAGAGA AAAGTAAAGA GCCCAGAGAC CCTGACCAGC TTTACAGCAC GCTCAAGAGC ATCCTCCAGC 1860 AGGTGAAGAG CCATCAAAGC GCTTGGCCCT TCATGGAACC TGTGAAGAGA ACAGAAGCTC 1920 CAGGATATTA TGAAGTTATA AGGTCCCCCA TGGATCTCAA AACCATGAGT GAACGCCTCA 1980 AGAATAGGTA CTACGTGTCT AAGAAATTAT TCATGGCAGA CTTACAGCGA GTCTTTACCA 2040 ATTGCAAAGA GTACAACGCC CCTGAGAGTG AATACTACAA ATGTGCCAAT ATCCTGGAGA 2100 AATTCTTCTT CAGTAAAATT AAGGAAGCTG GATTAATTGA CAAGTGATTT TTTTTCCCCC 2160 TCTGCTTCTT AGAAACTCAC CAAGCAGTGT GCCTAAAGCA AGGT

89

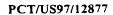
(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2093 base pairs
- (B) TYPE: nucleic acid (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GAATTCCGGC	GAAACCACTC	ATGTCTTTGG	GCGAAGCCTT	CTCCGGTCCA	TTTTCACCGT	60
TACCCGCCGG	CAGCTGCTGG	AAAAGTTCCG	AGTGGAGAAG	GACAAATTGG	TGCCCGAGAA	120
GAGGACCCTC	ATCCTCACTC	ACTTCCCCAA	GTAAGGCTCC	TTCTGGCCTA	CCAGGATTTG	180
GCCCCAAGTT	CACATCCTCC	CTGTTGTCCC	CTTTTTTCCA	GGAAGGCTTC	CTGGATTGGT	240
CCCTCCTCTC	CCTCCATGGG	CCTTTTGGGA	TCTGGGCGTC	TACCTGGCAG	ACTTGCCCAT	300
GGCCCAGAAG	CAACTTGCTA	GTACTAGTCT	GGGGATGGCA	GATTCCTGTC	CATGCTGGAG	360
GAGGAGATCT	ATGGGGCAAA	CTCTCCAATC	TGGGAGTCAG	GCTTCACCAT	GCCACCCTCA	420









GAGGGGACAC	AGCTGGTTCC	CCGGCCAGCT			TCCCAGCACC	480
CCCATCTTCA				CCCTGAGTCT	GGATTCTGCA	540
GGGGCCGAGC	CTATGCCAGG	CGAGAAGAGG	ACGCTCCCAG		CCTGGAGGAT	600
GCCAAGCGGC	TCCGTGTGAT	GGGTGACATC	CCCATGGAGC	TGGTCAATGA	GGTCATGCTG	660
ACCATCACTG	ACCCTGCTGC	CATGCTGGGG	CCTGAGACGA	GCCTGCTTTC	GGCCAATGCG	720
	AGACAGCCCG	CCTGGAGGAG	CGCCGCGGCA	TCATCGAGTT	CCATGTCATC	780
GGCAACTCAC	TGACGCCCAA	GGCCAACCGG	CGGGTGTTGC	TGTGGCTCGT	GGGGCTGCAG	840
AATGTCTTTT	CCCACCAGCT	GCCGCGCATG	CCTAAGGAGT	ATATCGCCCG		900
GACCCGAAGC	ACAAGACTCT	GGCCTTGATC	AAGGATGGGC	GGGTCATCGG		960
TTCCGCATGT	TTCCCACCCA	GGGCTTCACG	GAGATTGTCT	TCTGTGCTGT	CACCTCGAAT	1020
	AGGGTTATGG	GACCCACCTG	ATGAACCACC	TGAAGGAGTA	TCACATCAAG	1080
CACAACATTC	TCTACTTCCT	CACCTACGCC	GACGAGTACG	CCATCGGCTA	CTTCAAAAAG	1140
CAGGGTTTCT	CCAAGGACAT	CAAGGTGCCC	AAGAGCCGCT	ACCTGGGCTA		1200
TACGAGGGAG	CGACGCTGAT	GGAGTGTGAG	CTGAATCCCC	GCATCCCCTA	CACGGAGCTG	1260
TCCCACATCA	TCAAGAAGCA	GAAAGAGATC	ATCAAGAAGC	TGATTGAGCG	CAAACAGGCC	1320
CAGATCCGCA	AGGTCTACCC		TGCTTCAAGG	AGGGCGTGAG	GCAGATCCCT	1380
GTGGAGAGCG	TTCCTGGCAT	TCGAGAGACA	GGCTGGAAGC	CATTGGGGAA		1440
	AGGACCCCGA	CCAGCTCTAC	ACAACCCTCA	AAAACCTGCT	GGCCCAAATC.	1500
AAGTCTCACC		GCCCTTCATG	GAGCCTGTGA	AGAAGTCGGA	GGCCCCTGAC	1560
TACTACGAGG		CCCCATTGAC				1620
CGCTACTACG		GCTCTTTGTG	GCCGACCTGC	AGCGGGTCAT	CGCCAACTGT	1680
	ACCCCCGGA	CAGCGAGTAC	TGCCGCTGTG		GGAGAAGTTC	1740
TTCTACTTCA	AGCTCAAGGA	GGGAGGCCTC	ATTGACAAGT	AGGCCCATCT	TTGGGCCGCA	1800
GCCCTGACCT				TCCTTAGGGG		1860
CCCACGGACC		TGAGACACTC	CAGCCAAGGG	TCCTCCGGAC		1920
AGCTCTTTCT		GCACCCCAA	GCGTGCAGCT			1980
TGTGAGAGGT	CTCCTGGGTT	GGGGCCCAGC				2040
ACCTTGCCCA		CCCCAGGCC	TGGTCCCCAA	GAGCCCGGAA	TTC	2093
.,						

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9046 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CCTTGTTTGT	GTGCTAGGCT	GGGGGGGAGA	GAGGGCGAGA	0	GAGAGTGGGC	60
AAGCAGGACG	CCGGGCTGAG	TGCTAACTGC	GGGACGCAGA	GAGTGCGGAG	GGGAGTCGGG	120
	GCGGCAGGGG	CCAGAACAGT	GGCAGGGGGC	CCGGGGCGCA	CGGGCTGAGG	180
TCGGAGAGAG		-	000110000	GGTCCAGCAG	CCGGGCCGGC	240
CGACCCCCAG	CCCCCTCCCG	TCCGCACACA	00000	CCGTCTTCTC	CCGCCGCCGC	300
GTCGACGCTA	GGGGGGACCA	TTACATAACC	00000000			360
GGCGCCCGAA	CTGAGCCCGG	GGCGGGCGCT	CCAGCACTGG	CCGCCGGCGT	GGGGCGTAGC	-
AGCGGCCGTA	TTATTATTTC	GCGGAAAGGA	AGGCGAAGGA	GGGGAGCGCC	GGCGCGAGGA	420
GGGGCCGCCT	GCGCCCGCCG	CCGGAGCGGG	GCCTCCTCGG	TGGGCTCCGC	GTCGGCGCGG	480
	GGCGCTGCTC	GGCCCGGCCC	CCTCGGCCCT	CTGGTCCGGC	CAGCTCCGCT	540
GCGTGCGGGC		TCCGCCGGCC	GCCGCGCGAT	GTGAGGCGGC	GGCGCCAGCC	600
CCCGGCGTCC	TTGCCGCGCC	_	CCATTAGGGG	CCGGTGCGGC		660
TGGCTCTCGG	CTCGGGCGAG	TTCTCTGCGG	••••		GGGGCACCGG	720
AGCGCGGCGG	CAGGAGGAGG	GTTCGGAGGG	TGGGGGCGCA			
GAGGAGGTGA	GTGTCTCTTG	TCGCCTCCTC	CTCTCCCCCC	TTTTCGCCCC	CGCCTCCTTG	780
TGGCGATGAG	AAGGAGGAGG	ACAGCGCCGA	GGAGGAAGAG	GTTGATGGCG	GCGGCGGAGC	840
TCCGAGAGAC			CCGTGGCGGG	CCGGGGACTG	CGCCTCTAGA	900
• •				GGCGAATTTG	TGCTCTTGTG	960
GCCGCGAGTT				CCCTTACCTT	TTCTATCGAG	1020
CCCTCCTCCG					CCCCCACCCC	1080
TCCGCATCCC						1140
CTCGGGTGCC	GTCGGAGCCC	CCCAGCCCAC		GGCGCGGGGA		
AAGAAGAGAT	TTCCTGAGGA	. TTCTGGTTTT		ATCTCCGAAA		1200
TGGCCGAGAA		CCGGGGCCGC	CTTCAGCCAA	GCGGCCTAAA		1260
CGGCCCTCTC			CAGATTTTGG	CTCTCTATTT	GACTTGGAGC	1320
					GGTGGTGATA	1380
ACGACTTACO	, AGALGAALIA	, WICHWOICIU				





TTAATCAGCT TCAGACAAGT CTTGGCATGG TACAAGATGC AGCTTCTAAA CATAAACAGC 1440 TGTCAGAATT GCTGCGATCT GGTAGTTCCC CTAACCTCAA TATGGGAGTT GGTGGCCCAG-GTCAAGTCAT GGCCAGCCAG GCCCAACAGA GCAGTCCTGG ATTAGGTTTG ATAAATAGCA 1560 TGGTCAAAAG CCCAATGACA CAGGCAGGCT TGACTTCTCC CAACATGGGG ATGGGCACTA GTGGACCAAA TCAGGGTCCT ACGCAGTCAA CAGGTATGAT GAACAGTCCA GTAAATCAGC CTGCCATGGG AATGAACACA GGGACGAATG CGGGCATGAA TCCTGGAATG TTGGCTGCAG 1740 GCAATGGACA AGGGATAATG CCTAATCAAG TCATGAACGG TTCAATTGGA GCAGGCCGAG 1800 GGCGACAGGA TATGCAGTAC CCAAACCCAG GCATGGGAAG TGCTGGCAAC TTACTGACTG 1860 AGCCTCTTCA GCAGGGCTCT CCCCAGATGG GAGGACAAAC AGGATTGAGA GGCCCCCAGC 1920 CTCTTAAGAT GGGAATGATG AACAACCCCA ATCCTTATGG TTCACCATAT ACTCAGAATC CTGGACAGCA GATTGGAGCC AGTGGCCTTG GTCTCCAGAT TCAGACAAAA ACTGTACTAT 2040 CAAATAACTT ATCTCCATTT GCTATGGACA AAAAGGCAGT TCCTGGTGGA GGAATGCCCA 2100 ACATGGGTCA ACAGCCAGCC CCGCAGGTCC AGCAGCCAGG TCTGGTGACT CCAGTTGCCC AAGGGATGGG TTCTGGAGCA CATACAGCTG ATCCAGAGAA GCGCAAGCTC ATCCAGCAGC AGCTTGTTCT CCTTTTGCAT GCTCACAAGT GCCAGCGCCG GGAACAGGCC AATGGGGAAG 2280 TGAGGCAGTG CAACCTTCCC CACTGTCGCA CAATGAAGAA TGTCCTAAAC CACATGACAC ACTGCCAGTC AGGCAAGTCT TGCCAAGTGG CACACTGTGC ATCTTCTCGA CAAATCATTT 2340 2400 CACACTGGAA GAATTGTACA AGACATGATT GTCCTGTGTG TCTCCCCCTC AAAAATGCTG 2460 GTGATAAGAG AAATCAACAG CCAATTTTGA CTGGAGCACC CGTTGGACTT GGAAATCCTA 2520 GCTCTCTAGG GGTGGGTCAA CAGTCTGCCC CCAACCTAAG CACTGTTAGT CAGATTGATC 2580 CCAGCTCCAT AGAAAGAGCC TATGCAGCTC TTGGACTACC CTATCAAGTA AATCAGATGC 2640 CGACACAACC CCAGGTGCAA GCAAAGAACC AGCAGAATCA GCAGCCTGGG CAGTCTCCCC 2700 AAGGCATGCG GCCCATGAGC AACATGAGTG CTAGTCCTAT GGGAGTAAAT GGAGGTGTAG 2760 GAGTTCAAAC GCCGAGTCTT CTTTCTGACT CAATGTTGCA TTCAGCCATA AATTCTCAAA 2820 ACCCAATGAT GAGTGAAAAT GCCAGTGTGC CCTCCCTGGG TCCTATGCCA ACAGCAGCTC 2880 AACCATCCAC TACTGGAATT CGGAAACAGT GGCACGAAGA TATTACTCAG GATCTTCGAA ATCATCTTGT TCACAAACTC GTCCAAGCCA TATTTCCTAC GCCGGATCCT GCTGCTTTAA 2940 3000 AAGACAGACG GATGGAAAAC CTAGTTGCAT ATGCTCGGAA AGTTGAAGGG GACATGTATG 3060 AATCTGCAAA CAATCGAGCG GAATACTACC ACCTTCTAGC TGAGAAAATC TATAAGATCC AGAAAGAACT AGAAGAAAAA CGAAGGACCA GACTACAGAA GCAGAACATG CTACCAAATG CTGCAGGCAT GGTTCCAGTT TCCATGAATC CAGGGCCTAA CATGGGACAG CCGCAACCAG GAATGACTTC TAATGGCCCT CTACCTGACC CAAGTATGAT CCGTGGCAGT GTGCCAAACC 3300 AGATGATGCC TCGAATAACT CCACAATCTG GTTTGAATCA ATTTGGCCAG ATGAGCATGG CCCAGCCCC TATTGTACCC CGGCAAACCC CTCCTCTTCA GCACCATGGA CAGTTGGCTC 3420 AACCTGGAGC TCTCAACCCG CCTATGGGCT ATGGGCCTCG TATGCAACAG CCTTCCAACC AGGGCCAGTT CCTTCCTCAG ACTCAGTTCC CATCACAGGG AATGAATGTA ACAAATATCC 3480 3540 CTTTGGCTCC GTCCAGCGGT CAAGCTCCAG TGTCTCAAGC ACAAATGTCT AGTTCTTCCT 3600 GCCCGGTGAA CTCTCCTATA ATGCCTCCAG GGTCTCAGGG GAGCCACATT CACTGTCCCC AGCTTCCTCA ACCAGCTCTT CATCAGAATT CACCCTCGCC TGTACCTAGT CGTACCCCCA CCCCTCACCA TACTCCCCCA AGCATAGGGG CTCAGCAGCC ACCAGCAACA ACAATTCCAG 3780 CCCCTGTTCC TACACCACCA GCCATGCCAC CTGGGCCACA GTCCCAGGCT CTACATCCCC 3840 CTCCAAGGCA GACACCTACA CCACCAACAA CACAACTTCC CCAACAAGTG CAGCCTTCAC 3900
TTCCTGCTGC ACCTTCTGCT GACCAGCCC AGCAGCAGCC TCGCTCACAG CAGAGCACAG 3960
CAGCGTCTGT TCCTACCCCA AACGCACCGC TGCTTCCTCC GCAGCCTGCA ACTCCACTTT 4020
CCCAGCCAGC TGTAAGCATT GAAGGACAGG TATCAAATCC TCCATCTACT AGTAGCACAG 4080 AAGTGAATTC TCAGGCCATT GCTGAGAAGC AGCCTTCCCA GGAAGTGAAG ATGGAGGCCA 4140 AAATGGAAGT GGATCAACCA GAACCAGCAG ATACGCAGCC GGAGGATATT TCAGAGTCTA 4200 AAGTGGAAGA CTGTAAAATG GAATCTACCG AAACAGAAGA GAGAAGCACT GAGTTAAAAA 4260 CTGAAATAAA AGAGGAGGAA GACCAGCCAA GTACTTCAGC TACCCAGTCA TCTCCGGCTC CAGGACAGTC AAAGAAAAAG ATTTTCAAAC CAGAAGAACT ACGACAGGCA CTGATGCCAA 4380 CATTGGAGGC ACTTTACCGT CAGGATCCAG AATCCCTTCC CTTTCGTCAA CCTGTGGACC CTCAGCTTTT AGGAATCCCT GATTACTTTG ATATTGTGAA GAGCCCCATG GATCTTTCTA 4500 CCATTAAGAG GAAGTTAGAC ACTGGACAGT ATCAGGAGCC CTGGCAGTAT GTCGATGATA 4560 TTTGGCTTAT GTTCAATAAT GCCTGGTTAT ATAACCGGAA AACATCACGG GTATACAAAT 4620 ACTGCTCCAA GCTCTCTGAG GTCTTTGAAC AAGAAATTGA CCCAGTGATG CAAAGCCTTG 4680 GATACTGTTG TGGCAGAAG TTGGAGTTCT CTCCACAGAC ACTGTGTTGC TACGGCAAAC 4740 AGTTGTGCAC AATACCTCGT GATGCCACTT ATTACAGTTA CCAGAACAGG TATCATTTCT GTGAGAAGTG TTTCAATGAG ATCCAAGGGG AGAGCGTTTC TTTGGGGGGAT GACCCTTCCC 4860 AGCCTCAAAC TACAATAAAT AAAGAACAAT TTTCCAAGAG AAAAAATGAC ACACTGGATC 4920 CTGAACTGTT TGTTGAATGT ACAGAGTGCG GAAGAAAGAT GCATCAGATC TGTGTCCTTC ACCATGAGAT CATCTGGCCT GCTGGATTCG TCTGTGATGG CTGTTTAAAG AAAAGTGCAC GAACTAGGAA AGAAAATAAG TTTTCTGCTA AAAGGTTGCC ATCTACCAGA CTTGGCACCT 4980 5100 TTCTAGAGAA TCGTGTGAAT GACTTTCTGA GGCGACAGAA TCACCCTGAG TCAGGAGAGG 5160





телететта	AGTAGTTCAT	GCTTCTGACA	AAACCGTGGA	AGTAAAACCA	GGCATGAAAG	5220
CARCETTTET	GGACAGTGGA	GAGATGGCAG	AATCCTTTCC	ATACCGAACC	AAAGCCCTCT	5280
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NECCET TOX	CTGCCCTCCA	CCCAACCAGA	GGAGAGTATA	CATATCTTAC	CTCGATAGTG	5400
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TEGETTGCAT!	T CATCAAGCAG	CGGGCTGCCA	AGTATGCCAP	CTCTAATCCA	CAACCCATCC	7500
CTEGGCAGC	TGGCATGCCC	CAGGGGCAGC	CAGGGCTACA	L GCCACCTACO	: ATGCCAGGTC	7560
AGCAGGGG	T CCACTCCAAT	CCAGCCATGC	: AGAACATGAA	A TCCAATGCAG	GCGGGCGTTC	7620
ACAGGGCTG	G CCTGCCCCAG	CAGCAACCAC	: AGCAGCAAC1	CCAGCCACCC	: ATGGGAGGGA	7680
TGAGCCCCC	A GGCTCAGCAG	ATGAACATGA	ACCACAACAC	CATGCCTTCA	CAATTCCGAG	7740
ACATCTTGA	G ACGACAGCAA	ATGATGCAAC	: AGCAGCAGCA	ACAGGGAGCA	GGGCCAGGAA	7800
TACALCITUA TACACCCCTA	G AATGGCCAAC	CATAACCAGT	TCCAGCAAC	CCAAGGAGTT	GGCTACCCAC	7860
CACACCCCC	A GCAGCGGATG	CAGCATCACA	TGCAACAGA	r gcaacaagg	AATATGGGAC	7920
ACAMACGCC	A GCTTCCCCAG	GCCTTGGGA	CAGAGGCAG	TGCCAGTCT	CAGGCCTATC	7980
AGATAGGCC	T CCTTCAGCAL	CAGATGGGG	י ככככייפיייכו	GCCCAACCC	ATGAGCCCCC	8040
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ATTCTCTCT	C CAATCAAGIC	B MCCCCAACC	TCCACCCTC	A GCCTTCTCC	A CACCACGTTT	.8220
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CCCCACAGA	C AAGTTCCCC	CATCCTGGAC	TGGTAGCTG	C CCAGGCCAA	C CCCATGGAAC	
AAGGGCATT	T TGCCAGCCC	GACCAGAAT"	CAATGCTTT	C TOMOGRACIA	r AGCAATCCAG	8400
GCATGGCAA	A CCTCCATGG	GCAAGCGCC	A CGGACCTGG	ACTUAGUAU	C GATAACTCAG	8460
ACTTGAATT	C AAACCTCTC	CAGAGTACA	TAGACATAC	A CTAGAGACA	C CTTGTATTTT	
GGGAGCAAA	TTTATTAAA A	r ctcttaaca	A GACTTTTTG	T ACTGAAAAC	A ATTTTTTTGA	8520
ATCTTTCGT	A GCCTAAAAG	A CAATTTTCC	T TGGAACACA	T AAGAACTGT	G CAGTAGCCGT	8580
TTGTGGTT	TA AAGCAAACA	r gcaagatga	A CCTGAGGGA	T GATAGAATA	C AAAGAATATA	8640
TTTTTGTTA	T GGGCTGGTT.	A-CCACCAGCC	T TTCTTCCCC	T TTGTGTGTG	T GGTTCAAGTG	8700
TGCACTGG	SA GGAGGCTGA	G GCCTGTGAA	G CCAAACAAT	A TGCTCCTGC	C TTGCACCTCC	8760
AATAGGTTT	ጥጥጥጥልጥጥል ጥባ	AATTAAATTAA	T GAACATATG	TAATATAAT	G AACATATGTA	8820
ותממדדמדה	TTATTATT	A CTGGTGCAG	A TGGTTGACA	TTTTCCCTA	T TTTCCTCACT	8880
TTATGGAA	GA GTTAAAACA	T TTCTAAACC	A GAGGACAAA	A GGGGTTAAT	G TTACTTTGAA	8940





ATTACATTCT ATATATAT AAATATATA AAATATATA TAAAATACCA GTTTTTTTC 9000 TCTGGGTGCA AAGATGTTCA TTCTTTTAAA AAATGTTTAA AAAAAA 9046

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(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7326 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

		_			-		
	ATGGCCGAGA	ACTTGCTGGA	CGGACCGCCC	AACCCCAAAC	GAGCCAAACT	CAGCTCGCCC	60
	GGCTTCTCCG	CGAATGACAA	CACAGATTTT	GGATCATTGT	TTGACTTGGA	AAATGACCTT	120
			TGGAGAATTA			· -	180
			ACAACTGTCA				240
						CCTTGGTGGC	300
			CAGTACAAAC				360
			ATCAACACCC				420
			CCAAGCACTG			_	480
			ATCACAGACT				540
			TCTCAATAGT				600
			CATGAATGGA				660
			TCCAGCCATG				720
			ACAAATGGCT				780
			AATGACTGGT				840
			GGGAGCCACT				900
			TGCTTTTCCT				960
			GCAAACATCA				1020
			TGAAAAACGC				1080
			GAGACGAGAG				1140
			GAAAAACGTT				1200
			TTGTGCATCT				1260
			TGTTTGCCTC				1320
			TCCAGCTAGT				1320
			TTCCTTAAGT				1440
			AGGACTCCCC				1500
			ACCAGCACAG				1560
			CATGAGTGTC				1620
			ATCAGCTCTT				1680
			TGGTAACATT				1740
			TCGAAAAGGC				1800
			CGTTCAAGCC				1860
			CCTGGTTGCC				1920
			TGAATACTAT				1980
			GCGGAGGACA				2040
*			TGGGGCTCAG				2100
			GCCTTTGCCA			and the second s	2160
	AATTCATTTA	ACCCAATGTC	CCTGGGAAAC	GTCCAGTTGC	CACAGGCACC	CATGGGACCT	2220
			CCACTCTGTG				2280
	ATGGCCATTT	CTCCTTCACG	GATGCCTCAG	CCTCCAAATA	TGATGGGCAC	TCATGCCAAC	2340
	AACATTATGG	CCCAGGCACC	TACTCAGAAC	CAGTTTCTGC	CACAGAACCA	GTTTCCATCA	2400
	TCCAGTGGGG	CAATGAGTGT	GAACAGTGTG	GGCATGGGGC	AACCAGCAGC	CCAGGCAGGT	2460
			TGGAGCTGCT				2520
			CCCACCAGTG				2580
			CATGCCCTCT				2640
			CACTCAGCCA				2700
		•	GCCCAGCGCT				2760
			GACTCCACAG				2820
			GCAGCAACCA				2880
			CAGCATTGAT				2940





AGTGCTGAAA CCAGTTCCCA GCAGCCAGGA CCCGATGTGC CCATGCTGGA AATGAAGACA	3000
GAGGTGCAGA CAGATGATGC TGAGCCTGAA CCTACTGAAT CCAAGGGGGA ACCTCGGTCT	3060
GAGATGATGG AAGAGGATTT ACAAGGTTCT TCCCAAGTAA AAGAAGAGAC AGATACGACA	3120
GAGCAGAAGT CAGAGCCAAT GGAAGTAGAA GAAAAGAAAC CTGAAGTAAA AGTGGAAGCT	3180
AAAGAGGAAG AAGAGAACAG TTCGAACGAC ACAGCCTCAC AATCAACATC TCCTTCCCAG	3240
CCACGCAAAA AAATCTTTAA ACCCGAGGAG CTACGCCAGG CACTTATGCC AACTCTAGAA	3300
GCACGCAAAA AAATCITTAA ACCCGAGGAG CTACGCCAGG CACTTTTGGT AGCCTGTAGA TCCTCAGCTC	3360
GCACTCTATC GACAGGACCC AGAGTCTTG CCTTTCGTC AGGCCCTTTC TACCATCALA	3420
CTAGGAATCC CAGATTATTT TGATATAGTG AAGAATCCTA TGGACCTTTC TACCATCAAA	3480
CGAAAGCTGG ACACAGGGCA ATATCAAGAA CCCTGGCAGT ATGTGGATGA TGTCAGGCTT	3540
ATGTTCAACA ATGCGTGGCT ATATAATCGT AAAACGTCCC GTGTATATAA ATTTTGCAGT	3600
AAACTTGCAG AGGTCTTTGA ACAAGAAATT GACCCTGTCA TGCAGTCTCT TGGATATTGC	
TGTGGACGAA AGTATGAGTT CTCCCCACAG ACTTTGTGCT GTTACGGAAA GCAGCTGTGT	3660
ACAATTCCTC GTGATGCAGC CTACTACAGC TATCAGAATA GGTATCATTT CTGTGGGAAG	3720
TGTTTCACAG AGATCCAGGG CGAGAATGTG ACCCTGGGTG ACGACCCTTC CCAACCTCAG	3780
ACCACATTT CCAAGGATCA ATTTGAAAAG AAGAAAAATG ATACCTTAGA TCCTGAACCT	3840
TTTGTTGACT GCAAAGAGTG TGGCCGGAAG ATGCATCAGA TTTGTGTTCT ACACTATGAC	3900
ATCATTIGGC CTTCAGGTTT TGTGTGTGAC AACTGTTTGA AGAAAACTGG CAGACCTCGG	3960
ADDEDDDCD DATTCAGTGC TAAGAGGCTG CAGACCACAC GATTGGGAAA CCACTTAGAA	4020
GACAGAGTGA ATAAGTTTTT GCGGCGCCAG AATCACCCTG AAGCTGGGGA GGTTTTTGTC	4080
AGAGTGGTGG CCAGCTCAGA CAAGACTGTG GAGGTCAAGC CGGGAATGAA GTCAAGGTTT	4140
GTGGATTCTG GAGAGATGTC GGAATCTTTC CCATATCGTA CCAAAGCACT CTTTGCTTTT	4200
GAGGAGATCG ATGGAGTCGA TGTGTGCTTT TTTGGGATGC ATGTGCAAGA TACGGCTCTG	4260
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TATGTGAAGA AATTGGTGTA TGTGACAGCA CATATTTGGG CCTGTCCCCC AAGTGAAGGA	4440
GATGACTATA TCTTTCATTG CCACCCCCT GACCAGAAAA TCCCCAAACC AAAACGACTA	4500
CAGGAGTGGT ACAAGAAGAT GCTGGACAAG GCGTTTGCAG AGAGGATCAT TAACGACTAT	4560
CAGGAGTGGT ACAAGAAGAT GCTGGACAAG GCGTTTGCAG AGAGGATCAT TATGGACTAT AAGGACATCT TCAAACAAGC GAACGAAGAC AGGCTCACGA GTGCCCAAGGA GTTGCCCTAT	4620
AAGGACATCT TCAAACAAGG GAACGAAGAC AGGCCACCA GIGCCAAGA GIGCCAAGA GIGCCAAGA GAACGAAGAC AGGCCAAGA GIGCCAAGA GIGCCAAGA GAACGAAGAC AGGCCAAGA GIGCCAAGA GIGCCAAGA GIGCCAAGA GAACGAAGAC AGGCCAAGA GIGCCAAGA GAACGAAGA GIGCCAAGA GICCAAGA G	4680
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GAAGAAAA GGAAAAAAGA AGAGAGTACT GCAGCGAGTG AGACTCCTGA GGGCAGTCAG	4800
GTGACAGCA AAAATGCGAA GAAAAAGAAC AACAAGAAGA CCAACAAAAA CAAAAGCAGC	4860
ATTAGCCGCG CCAACAAGAA GAAGCCCAGC ATGCCCAATG TTTCCAACGA CCTGTCGCAG	4920
AAGCTGTATG CCACCATGGA GAAGCACAAG GAGGTATTCT TTGTGATTCA TCTGCATGCT	4980
GGGCCTGTTA TCAGCACTCA GCCCCCCATC GTGGACCCTG ATCCTCTGCT TAGCTGTGAC	
CTCATGGATG GGCGAGATGC CTTCCTCACC CTGGCCAGAG ACAAGCACTG GGAATTCTCT	5040
TCCTTACGCC GCTCCAAATG GTCCACTCTG TGCATGCTGG TGGAGCTGCA CACACAGGGC	5100
CAGGAGCGCT TTGTTTATAC CTGCAATGAG TGCAAACACC ATGTGGAAAC ACGCTGGCAC	5160
TGCACTGTGT GTGAGGACTA TGACCTTTGT ATCAATTGCT ACAACACAAA GAGCCACACC	5220
CATARGATGG TGARGTGGGG GCTAGGCCTA GATGATGAGG GCAGCAGTCA GGGTGAGCCA	5280
CAGTCCARGA GCCCCCAGGA ATCCCGGCGT CTCAGCATCC AGCGCTGCAT CCAGTCCCTG	5340
GTGCATGCCT GCCAGTGTCG CAATGCCAAC TGCTCACTGC CGTCTTGCCA GAAGATGAAG	5400
CGAGTCGTGC AGCACACCAA GGGCTGCAAG CGCAAGACTA ATGGAGGATG CCCAGTGTGC	5460
ARGORGOTOR TEGOTOTTE CEGOTACOAC GCCAAACACE GCCAAGAAAA TAAAEGCCCE	552 0
GTGCCTTCT GCCTCAACAT CAAACATAAC GTCCGCCAGC AGCAGATCCA GCACTGCCTG	5580
CACCAGGCTC ACCTCATGCG CCGGCGAATG GCAACCATGA ACACCCGCAA TGTGCCTCAG	5640
CAGAGTTTGC CTTCTCCTAC CTCAGCACCA CCCGGGACTC CTACACAGCA GCCCAGCACA	5700
CCCCAAACAC CACAGCCCCC AGCCCAGCCT CAGCCTTCAC CTGTTAACAT GTCACCAGCA	5760
GGCTTCCCTA ATGTAGCCCG GACTCAGCCC CCAACAATAG TGTCTGCTGG GAAGCCTACC	5820
AACCAGGTGC CAGCTCCCC ACCCCCTGCC CAGCCCCCAC CTGCAGCAGT AGAAGCAGCC	5880
CGGCAAATTG AACGTGAGGC CCAGCAGCAG CAGCACCTAT ACCGAGCAAA CATCAACAAT	5940
GGCAAATTG AACGTGAGGC CCAGCAGGA CCCCAGGAA GCCAAATGAC TCCTGTGGGC	6000
CTGAATGTGC CCCGTCCCAA CCAAGTCAGT GGGCCTGTCA TGTCTAGTAT GCCACCTGGG	6060
CTGAATGTGC CCCGTCCCAA CCAAGTCAGT GGGCCTGTCA TGTCTAGTAT GCCACCTGTA CAGTGGCAGC AGGCACCCAT CCCTCAGCAG CAGCCGATGC CAGGCATGCC CAGGCCTGTA	6120
CAGTGGCAGC AGGCACCAT CCCTCAGCAG CAGCCGATGC CAGCCATTGCCCAAACC	6180
ATGTCCATGC AGGCCCAGGC AGCAGTGGCT GGGCCACGGA TGCCCAATGT GCAGCCAAAC	6240
AGGAGCATCT CGCCAAGTGC CCTGCAAGAC CTGCTACGGA CCCTAAAGTC ACCCAGCTCT	6300
CCTCAGCAGC AGCAGCAGGT GCTGAACATC CTTAAATCAA ACCCACAGCT AATGGCAGCT	6360
TTCATCAAAC AGCGCACAGC CAAGTATGTG GCCAATCAGC CTGGCATGCA GCCCCAGCCC	6420
GGACTTCAAT CCCAGCCTGG TATGCAGCCC CAGCCTGGCA TGCACCAGCA GCCTAGTTTG	
CAAAACCTGA ACGCAATGCA AGCTGGTGTG CCACGGCCTG GTGTGCCTCC ACCACAACCA	6480
GCAATGGGAG GCCTGAATCC CCAGGGACAA GCTCTGAACA TCATGAACCC AGGACACAAC	6540
CCCAACATGA CAAACATGAA TCCACAGTAC CGAGAAATGG TGAGGAGACA GCTGCTACAG	6600
CACCAGCAGC AGCAGCAGCA ACAGCAGCAG CAGCAGCAGC AACAACAAAA TAGTGCCAGC	6660
TTGGCCGGGG GCATGGCGGG ACACAGCCAG TTCCAGCAGC CACAAGGACC TGGAGGTTAT	6720





GCCCCAGCCA	TGCAGCAGCA	ACGCATGCAA	CAGCACCTCC	CCATCCAGGG	CAGCTCCATG	6780
GGCCAGATGG	CTGCTCCAAT	GGGACAACTT	GGCCAGATGG	GGCAGCCTGG	GCTAGGGGCA	6840
GACAGCACCC	CTAATATCCA	GCAGGCCCTG	CAGCAACGGA	TTCTGCAGCA	GCAGCAGATG	6900
AAGCAACAAA	TTGGGTCACC	AGGCCAGCCG	AACCCCATGA	GCCCCCAGCA	GCACATGCTC	6960
TCAGGACAGC	CACAGGCCTC	ACATCTCCCT	GGCCAGCAGA	TCGCCACATC	CCTTAGTAAC	7020
CAGGTGCGAT	CTCCAGCCCC	TGTGCAGTCT	CCACGGCCCC	AATCCCAACC	TCCACATTCC	7080
AGCCCGTCAC	CACGGATACA	ACCCCAGCCT	TCACCACACC	ATGTTTCACC	CCAGACTGGA	7140
ACCCCTCACC	CTGGACTCGC	AGTCACCATG	GCCAGCTCCA	TGGATCAGGG	ACACCTGGGG	7200
AACCCTGAAC	AGAGTGCAAT	GCTCCCCCAG	CTGAATACCC	CCAACAGGAG	CGCACTGTCC	7260
AGTGAACTGT	CCCTGGTTGG	TGATACCACG	GGAGACACAC	TAGAAAAGTT	TGTGGAGGGT	7320
TTGTAG						7326

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(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2499 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TCACTTGTCA ATTAATCCAG CTTCCTTAAT TTTACTGAAG AAGAATTTCT CCAGGATATT GGCACATTTG TAGTATTCAC TCTCAGGGGC GTTGTACTCT TTGCAATTGG TAAAGACTCG 120 CTGTAAGTCT GCCATGAATA ATTTCTTAGA CACGTAGTAC CTATTCTTGA GGCGTTCACT CATGGTTTTG AGATCCATGG GGGACCTTAT AACTTCATAA TATCCTGGAG CTTCTGTTCT 240 CTTCACAGGT TCCATGAAGG GCCAAGCGCT TTGATGGCTC TTCACCTGCT GGAGGATGCT CTTGAGCGTG CTGTAAAGCT GGTCAGGGTC TCTGGGCTCT TTACTTTTCT CTTTTCCACT CGGTTTCCAG CCTGTCTCTC TAATTCCAGG AATGCTTTCT ATAGGAATCT GTCGAACTCC ATCTTTAAAA CATGAAAGTC CAGGGTAAAC TTTTCGAATT TGTGCCTGTT TTCTTTCAAT 480 540 CAGTTTTTTA ATTATCTCCT TCTGCTTTTT AATGATGACA GAAAATTCTG TGTACGGGAT CCGTGGATTT AGCTCACATC CCATTAAAGT GGCTCCTTCA TAATCCTTGA TATAGCCAAC ATATTTGGTT TTAGGTATTT TAATTTCTTT GGAGAAACCC TGTTTCTTAA AGTATCCAAT 660 TGCATATTCA TCTGCATATG TGAGGAAGTT CAGGATGTCA TGCTTTATGT GATATTCTTT 720 CARATGATTC ATCAGGTGTG TTCCATAGCC CTTGACTTGC TCATTTGAGG TTACAGCACA 780 GAAGACAATC TCTGTGAATC CTTGAGATGG GAACATACGG AAACAGATAC CACCAATAAC ACGGCCATCT TTAATTAAAG CAAGGGTTTT GTGTTTCGGG TCAAAGACGA GCCGTGTGAT GTATTCTTTT GGCATTCGGG GCAGCTGGTG GGAGAAAACG TTCTGTAGGC CAACCAGCCA CATCAGGATC TTCTTGTTTG GTTTCTGGTT GAGGGAATTG CCAACCACGT GAAATTCAAT 1020 TACACCCCTG CGCTCTTCCA ACCTTGCCGC CTCATCCCTG GCCGAGTGTG CTGACAGAAA 1080 ATTGGTCTCT GGTCCAAGCA TTGCTGCAGG GTCCGTGATG GTAGACATAA CCTCGTTGAT 1140 TAATTCCATC GGAATATCCC CCATAACTCG GGGTTTCTTG GCCTCCTCCA GAACATGAGA 1200 ATCAGTCATT TTCCTCTTTT CTCCTGGGTT TGCCTCAAGT CCAGAAGAGG CTTTGCAGGC 1260 AGGACTGCTG CTCCCTGCGT TTGGCTGCTC AAGGGAAGAT GAGGTTGAAT TGTATGAAAT 1320 TGTCCCAGCC ACAGGAGGTG GATTGATAAC TGTTTGGATG CCTAGCTGGC TGGTTCTGGA 1380 AGAGGCTGAG AGAAAATCCT GATCCCAGAT GGGAGAGTTT TGACTATATA CTTCTTCTTC 1440 TAGCATGGAC AGAAATTTTG GGAAATGAGT GAGGATTAGA GTTCGTTTTT CAAGAGGCAG 1500 TTTATCTTTT TCCTGTCTTG CTTGTTCCAG GAGTTGTCGC CTCATAACAG TGAAGACCGA 1560 GCGAAGCAAT GTTCTCCCAA ACACCTGTGT GGTTTCGTAC CGAGGTAGAC TGTCGCAGAA 1620 CTGTGGCACG TTGCAGTAAC ACAGCCACCT TGTGTAGTTC TCTTTGTATC CAGAAATATC ATCATTGGGA GATCGCAGTC TTCGTTGAGA TGGTGCCTCC AGATGCCAAT AGTTGATGCG 1740 GTTTAGGAAC ATTTTTGCCA ACTCAACTAT TGTTTGCCTT TCTTTTGCTG GCAGGTGACT 1800 AAATTTGTAC TGCACAAAGT TATTCACACC CTGTTCAATG CTAGGTTTTT CAAATGGGGG 1860 TTTCTTTTCC AAAGAGCCTT CAACCACAGG TTTTCCTCTT TGTAAAATAG ACTTTCTCAA 1920 GAGCTTAAAT AGATAGAAAT AAACTTGTTT GGTATCTGCA TCTTCTTCCT TGTGGACACA 1980 GGTAAAGAGA TATTCCACAT CCAATACTAT TCCCAGGAGT CTGTTCATTT CTTCCTCTGA 2040 CACATTCTCC AGGTGGGAAA CATGAGCAGC TAGGGCATGG CTACAACTCC GACAGGATTC 2100 TGTTAGACTG ACAATTATTT GCTGCAGGTC GGCTCTGGGG GGAGTGGGTG AGGGGTTAGG 2160 GTTTTTCCAG CCATTACATT TACAAGACTC CTCGGCCTTG CAGGCGGAGT ACACTCCGAG 2220 TTTCTCCAGT TTCTTGGCCC GCGGAGCGGA GCGTAGTTGC GCTTTCTTCA CGGCGATTCG 2280 GGCCGAGCCA CCGCCTCCCG GTCCTTCGGC CGTGCCCGCT GCAGCCACTG CCGTCGCCGG 2340 ACCGCAGGCG CCCGAGCCCC CGGCGGCAGC GGCGCAGGGG GAGCCCTGCG GGGGCGCGGG 2400





CGGAAGCGCC GCAGGCTGCG GGGGCAGCGC CCCGGGCCCG GCCCCTGCCC CGGCTCCTGC 2460 2499 CCCGCAGCCG CCCGGCCAGC CTCGGACAT

96

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2442 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TCACTTGTCA	ATCAACCCTG	CTTCCTTAAT	TTTACTGAAG		CCAGGATGCT	60
GGCGCATTTG	TAGTACTCGC	TCTCGGGAGG	GTTGTACTCC		TGAACACTCG	120
TTGCAAGTCC	GCCATGAATA	ACTTCTTAGA	CACATAGTAC	CTGTTCCTGA		180
CATGGTTTTC	AGATCCATGG	GGAACCTTAT	AACTTCATAA	TATCCCGGAG	CTTCTGTTCT	240
CTTCACTGGT	TCCATGAAAG	GCCAAGCATT	TGGATGGTTC	TTCACCTGCT	GCAGGATGTT	300
CTTGAGGGTG	CTGTAAACGT	GCTCAGGGTC	TTTGGGCTCT	TTACTTTTCT	CTTTTCCACT	360
TGGTTTCCAG	CCTGTCTCTC	TGATTCCAGG	AATGCTTTCT	ATAGGAATCT	GCCGAACTCC	420
ATCTTTGAAA		CAGGGTAGAC	TTTTCGAATC	TGGGCTTGTT	TTCTTTCTAT	480
	ATGATCTCCT	TCTGCTTTTT	AATGATGACA	GAGAACTCTG	TGTATGGGAT	540
CTGAGGGTTC	AGCTCACATC	CCATCAAAGT	GGCCCCTTCA	TAATCCTTGA	TGTAGCCAAC	600
ATATTTGGTT	TTAGGTATTT	TGATTTCTTT	GGAGAAACCC	TGCTTCTTGA		660
GGCATACTCA	TCTGCATATG	TGAGGAAGTT	GAGGATCTCG	TGCTTTATGT	GGTATTCTTT	720
GAGATGGTTC	ATCAGGTGGG	TTCCATAGCC	CTTGACTTGT	TCATTTGAGG	TTACTGCACA	780
GAAAACAATC	TCTGTGAATC	CCTGGGATGG	AAACATCCGG		CACCAATGAC	840
ACGGCCATCT	TTAATTAAAG	CAAGGGTTTT	GTGTTTCGGG	TCAAAGACGA	GCCGTGTGAT	900
GTACTCTTTG	GGCATTCTGG		GGAAAACACA	TTCTGGAGGC	CCACGAGCCA	960
CATCAGGATC	TTCTTGTTTG	GTTTCTGGTT	CAGGGAGTTG	CCCACCACGT	GGAATTCAAT	1020
GACACCCCTG	CGTTCTTCCA	GCCGTGCCGC	CTCATCTCTG	GCCGAATGGG	CTGACAGAAA	1080
ATTGGTCTCT	GGTCCAAGCA			GTAGACATGA		1140
CAATTCCACG	GGAATATCCC	CCATCACTCG			GAGCATGAGA	1200
GTTGTTCATT	TTCCTCTTTT	CTCCCGGGTT	TGCTTCAAGC	CCAGAAGAGC	CTCTGCATCC	1260
AGGACTTGTT	CTCCCTCCAT			GAATTTGAAC	TGAACAATGC	1320
	ACAGGAGGAC		TTGGATTCCT	AGCGGGCTGG	TTCTGGAAGA	1380
GGCTGAGAGA	AAATCCTGAT	CCCAGATAGG			CTTCTTCCAA	1440 1500
CATGGACAGA		. AATGTGTGAG				1560
GTCTTTTTTC				ATGATGGTGA		1620
AAGCAATGTT		CCTTTGTGGT			CACAGAACTG	1680
CGGTACATTG		ACCACCTTGT			AGATGTCATC TGATGCGGTT	1740
ATTGGGAGAC			AGCCTCCAGA			1800
CAGAAACATC					AGTGACTAAA ATGGCGGCTT	1860
CTTGTACTGC					TTCTCAAGAG	1920
CTTCTCCAAC						1980
CTTGAATAGO						2040
GAAGAGGTAC						2100
ATTCTCCAAC			GGCATGGCTA			2160
	A ATTATCTGC				CGCCGAGTTT	2220
CTTCCAGCC					CGATCCGGGC	2280
CTCCAGCTT						2340
CGAGCCGCC'						2400
	A GAGCTCCCG	G CAGCGGTGG				2442
GGGAGGCAG'	r GCTGGGGAC	c ceecccec	AGCCTCGGC	_ Ar		2442

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear





(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

97

CCCGCCAGCC TCGGACATGC

20

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CCCGCCAGCC TCGGCCATGC

20

(2) INFORMATION FOR SEQ ID NO:18:

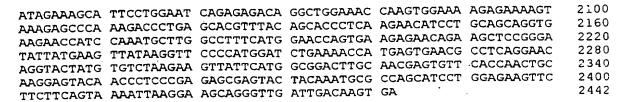
(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2442 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

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CCCCGGACCC	TGGCCACCGC	TGCCGGGAGC	TCTGCTTCCT	GCGGGCCAGC	GACGCCGGTG	120
	GCACCGCCGA					180
AAGGCGCAGT	TGCGCTCTGC	TCCGCGGGCG	AAGAAGCTGG	AGAAACTCGG	CGTGTACTCC	240
GCCTGCAAGG	CAGAGGAGTC	CTGTAAATGC	AATGGCTGGA	AGAACCCTAA	CCCCTCTCCT	300
ACTCCACCAA	GAGGAGACCT	CCAGCAGATA	ATTGTCAGTT	TGACAGAATC	CTGTCGAAGC	360
TGTAGCCATG	CCCTTGCTGC	TCACGTTTCT	CACTTGGAGA	ATGTGTCAGA	GGAAGAGATG	420
GACAGACTCC	TGGGAATTGT	GTTGGATGTG	GAGTACCTCT	TCACCTGCGT	CCACAAAGAA	480
GAAGATGCAG	ATACCAAACA	AGTGTACTTC	TACCTATTCA	AGCTCTTGAG	AAAGTCAATT	540
TTACAAAGAG	GAAAACCTGT	GGTTGAAGGC	TCCTTGGAGA	AGAAGCCGCC	ATTTGAGAAG	600
CCCAGTATTG	AACAGGGTGT	GAACAACTTC	GTGCAGTACA	AGTTTAGTCA	CTTGCCATCG	660
AAAGAGAGGC	AGACAACGAT	CGAGCTGGCC	AAGATGTTTC	TGAACCGCAT	CAACTACTGG	720
CATCTGGAGG	CTCCATCTCA	GCGGAGACTA	CGGTCTCCCA	ATGATGACAT	CTCTGGATAC	780
AAGGAAAACT	ACACAAGGTG	GTTGTGCTAC	TGCAATGTAC	CGCAGTTCTG	TGACAGCTTA	840
CCTCGGTACG	AAACCACAAA	GGTGTTTGGG	AGAACATTGC	TTCGCTCGGT	CTTCACCATC	900
ATGAGACGAC	AGCTCTTGGA	ACAAGCCAGA	CAGAAAAAAG	ACAAACTGCC	TCTTGAGAAA	960
CGCACGCTTA	TCCTCACACA	TTTCCCAAAG	TTTCTGTCCA	TGTTGGAAGA	AGAAGTGTAT	1020
	CTCCTATCTG					1080
	AAACAGTAAT					1140
	ATGAGCAGAT					1200
	CAAACCCGGG					1260
GCCAAGAGAT	CTCGAGTGAT	GGGGGATATT	CCCGTGGAAT	TGATCAATGA	GGTCATGTCT	1320
	ACCCTGCAGG				•	1380
	AGGCGGCACG					1440
GGCAACTCCC	TGAACCAGAA					1500
AATGTGTTTT	CCCACCAGCT			ACATCACACG		1560
	ACAAAACCCT			GTGTCATTGG		1620
	TTCCATCCCA					1680
0. 2 .0. 2 .0	AGGGCTATGG					1740
CACGAGATCC	TCAACTTCCT					1800
0000				ATGTTGGCTA		1860
	CCACTTTGAT					1920
	TTAAAAAGCA					1980
CAGATTCGAA	AAGTCTACCC	TGGACTTTCG	TGTTTCAAAG	ATGGAGTTCG	GCAGATTCCT	2040





1.

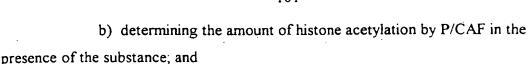


What is claimed is:

- 1. A purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones.
- 2. The protein of claim 1 consisting of the amino acid sequence of SEQ ID NO:1.
- 3. The protein of claim 1 comprising the amino acid sequence of SEQ ID NO:2.
- 4. The protein of claim 1, which also binds to the amino acid sequence of SEQ ID NO:3 on a p300 cellular protein and to amino acid residues 1805-1854 of a CBP cellular protein (SEQ ID NO:9).
- 5. A fragment of the protein of claim 1 having histone acetyltransferase activity.
- 6. A polypeptide consisting of the amino acid sequence of SEQ ID NO: 2.
- 7. A fragment of the protein of claim 1 which binds to the amino acid sequence of SEQ ID NO: 3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- 8. A polypeptide consisting of the amino acid sequence of SEQ ID NO:4.
- 9. A nucleic acid consisting of the nucleotide sequence of SEQ ID NO:10.
- 10. A nucleic acid having a nucleotide sequence which encodes the protein of claim

- A nucleic acid having a nucleotide sequence which encodes the protein of claim
- 12. A nucleic acid having a nucleotide sequence which encodes the protein of claim
 3.
- 13. A nucleic acid consisting of the nucleotide sequence which encodes the protein of claim 4.
- 14. A nucleic acid complementary to and which selectively hybridizes with the nucleic acid of claim 11 under stringent hybridization conditions.
- 15. A fragment of the nucleic acid of claim 9, which encodes a polypeptide that acetylates histones.
- A fragment of the nucleic acid of claim 9, which encodes a polypeptide which binds to the amino acid sequence of SEQ ID NO:3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- 17. A purified antibody which specifically binds the protein of claim 1.
- 18 A purified antibody which specifically binds the protein of claim 2.
- 19. A purified antibody which specifically binds the protein of claim 3.
- A purified antibody which specifically binds the protein of claim 4.
- An assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of P/CAF comprising:
- a) contacting the substance with a system in which histone acetylation by P/CAF can be determined,

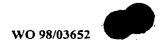




- c) comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.
- 22. An assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising.
- a) contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined,
- b) determining the amount of P/CAF binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP.
- 23. The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the p300 protein comprising amino acid residues 1767-1816 (SEQ ID NO:3) and the protein of claim 4.
- The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising amino acid residues 1805-1854 (SEQ ID NO:9) and the protein of claim 4.
- The method of claim 22, wherein the system consists of a cell extract produced from cells producing both p300 and P/CAF.



- An assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP comprising:
- a) contacting the substance with a system in which histone acetylation by p300/CBP can be determined;
- b) determining the amount of histone acetylation by p300/CBP in the presence of the substance; and
- c) comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP.
- An assay for screening substances for the ability to inhibit binding of a DNA-binding transcription factor to p300/CBP comprising:
- a) contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined;
- b) determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.
- 28. The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a DNA-binding transcription factor and p300/CBP.
- 29. The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising a DNA-binding transcription factor and p300/CBP.



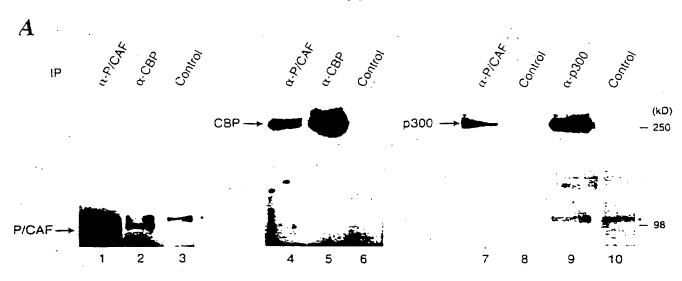


- The method of claim 27, wherein the system consists of a cell extract produced from cells producing both a DNA-binding transcription factor and p300/CBP
- The method of claim 27, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1
- 32. A method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.
- 33. The method of claim 32, wherein the substance can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP.
- A method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier
- -35. The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by promoting the binding of P/CAF to p300/CBP.
 - 36. The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by stimulating the histone acetlytransferase activity of P/CAF.
 - A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier





- 104
- The method of claim 37, wherein the substance can inhibit the transcription modulating activity of p300/CBP by preventing the binding of a DNA-binding transcription factor to p300/CBP
- The method of claim 38, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1.
- The method of claim 37, wherein the substance is an antibody which binds p300/CBP.
- A method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.
- The method of claim 41, wherein the substance can stimulate the histone acetyltransferase activity of p300/CBP by promoting the binding of a DNA-binding transcription factor to p300/CBP.



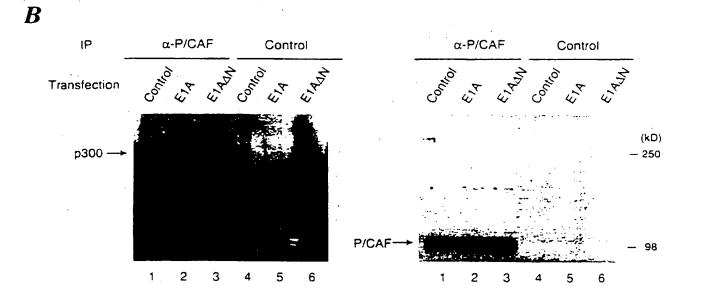


Fig. 1

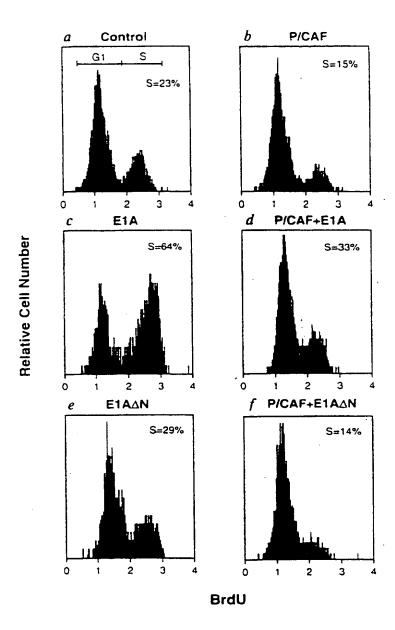


Fig. 2

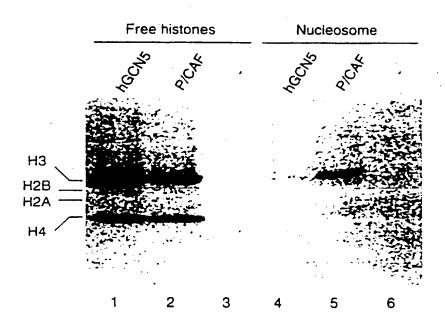


Fig. 3

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(74) Agents: MILLER, Mary, L. et al., Needle & Rosenberg, Suite 1200, 127 Peachtree Street, N.E., Atlanta, GA 30303 (US).

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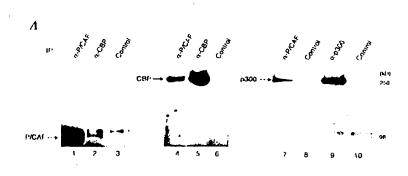
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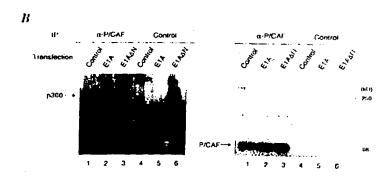
26 February 1998 (26.02.98)

(54) Title: P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

(57) Abstract

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein. The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided. Also provided are methods of screening for compounds that inhibit or stimulate the transcription modulating and histone acetyltransferase activity of P/CAF and p300/CBP.





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A. CLASS IPC 6	ification of subject matter C12N15/12 C07K14/47 G01N33	/50 A61K38/17	
According t	o International Patent Classification (IPC) or to both national classifi	cation and IPC	
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Minimum de IPC 6	ocumentation searched (classification system followed by classifica C07K	tian symbols)	
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C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the re	levant passages	Relevant to claim No.
A	EMBL EST, Accession numberN39522 Sequence no yv27b08.sl Homo sapi clone 243927 3' 25 January 1996 XP002050402 see the whole document		1
A	GEORGAKOPOULOS, T. & THIREOS, G. EMBO JOURNAL., vol. 11, 1992, EYNSHAM, OXFORD pages 4145-4152, XP002050399 see the whole document	: GB,	1
X Furth	er documents are listed in the continuation of box C.	Patent family members are listed in	n annex.
"A" document consider to consider the consideration that t	nt which may throw doubts on priority claim(s) or a cited to establish the publication date of another or other special reason (as specified) nt referring to an oral disclosure, use, exhibition or	"T" later document published after the inter- or priority date and not in conflict with cited to understand the principle or the invention "X" document of particular relevance; the c- cannot be considered novel or cannot involve an inventive step when the do- "Y" document of particular relevance; the c- cannot be considered to involve an involve an involve an involve an involve an involve an involve and involv	the application but early underlying the laimed invention be considered to current is taken alone laimed invention rentive step when the re other such docurs to a person skilled lamily
17	December 1997	1 4. 01. 98	
Name and ma	ailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer Chambonnet, F	

Form PCT/ISA/210 (second sheet) (July 1992)

		PC1/US	9//128//
C.(Continu	ation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.
Р,Х	YANG, X.Y. ET AL.: "A p300-CBP-associated factor that competes with the adenoviral oncoprotein E1A" NATURE., vol. 382, no. 8589, 25 July 1996, LONDON GB, pages 319-324, XP002050400 see the whole document		1
P,X	OGRYZKO, V.V. ET AL.: "The transcriptional coactivators p300 and CBP are histone acetyltransferases" CELL, vol. 87, no. 5, November 1996, NA US, pages 953-959, XP002050401 see the whole document		1
P,X	EMBL EST, Accession number U57316, Sequence reference human GCN5 (hGCN5) complete cds. 26 august 1996 XP002050403 see the whole document		1



INTERNATIONAL SEARCH REPORT

Box	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
	see FURTHER INFORMATION sheet PCT/ISA/210
2.	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such
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3	Claims Nos. because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This Inte	ernational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
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4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.
Form PCT	/ISA/210 (continuation of first sheet (1)) (July 1992)

International Application No PCT/US 97 /12877

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Remark: Although claims 32 to 42 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the composition.







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(74) Agents: MILLER, Mary, L. et al.; Needle & Rosenberg, Suite 1200, 127 Peachtree Street, N.E., Atlanta, GA 30303 (US).

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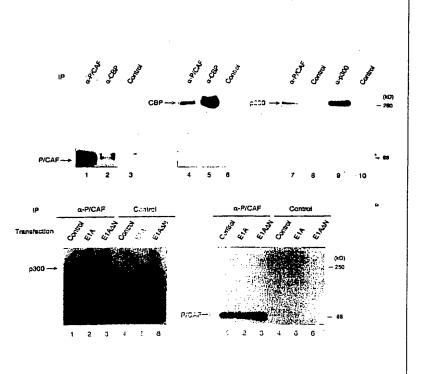
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P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

BACKGROUND OF THE INVENTION

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Field of the Invention

The present invention provides a transcriptional co-factor, p300/CBP-associated factor (P/CAF), which modulates transcription through binding to the cellular transcription co-factors p300 and CBP and through acetylation of histones. Also provided are methods for screening for the presence of P/CAF and for substances which alter the transcription modulating effect and growth regulatory activity of P/CAF.

Background Art

Cellular proteins p300 and CBP are global transcriptional coactivators that are involved in the regulation of various DNA-binding transcriptional factors (Janknecht and Hunter, 1996). Recently, p300 was found to be very closely related to CBP, a factor that binds selectively to the protein kinase A-phosphorylated form of CREB (3-5). Cellular factors p300 and CBP exhibit strong amino acid sequence similarity and share the capacity to bind both CREB and E1A (6-8). Although neither p300 nor CBP by itself binds to DNA, each can be recruited to promoter elements via interaction with sequence-specific activators and functions to be a transcriptional adaptor. For simplicity, p300 and CBP will be termed p300/CBP in the context of discussing their shared functional properties.

p300/CBP is a large protein consisting of over 2,400 amino acids, known to interact with a variety of DNA-binding transcriptional factors including nuclear hormone receptors (13,57), CREB (3,4, 7), c-Jun/v-Jun (9,11), YY1 (10), c-Myb/v-Myb (12,58), Sap-1a (59), c-Fos (11) and MyoD (60). DNA-binding factors recruit p300/CBP not only by direct but also indirect interactions through cofactors; for example, nuclear hormone receptors recruit p300/CBP directly as well as through indirect interactions, via SRC-1, which stimulates transcription by binding to various nuclear hormone receptors (13,61).



PCT/US97/12877

The transforming proteins encoded by adenovirus and several other small DNA tumor viruses disturb host cell growth control by interacting with cellular factors that normally function to repress cell proliferation. One of the most intensively studied of these viral proteins, the product of the adenovirus E1A gene, is itself sufficient for transformation (1). E1A transforming activity resides in two distinct domains, the targets of which include p300/CBP and products of the retinoblastoma (RB) susceptibility gene family (1,2). Interactions of E1A with p300/CBP and RB are thought to influence functionally distinct growth regulatory pathways, allowing the two domains to contribute additively to transformation (1).

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The paradigm for how E1A and functionally related viral proteins perturb cell growth regulation derives in large part from studies on their interactions with RB (1,2). The molecular function of E1A is based on its capacity to interfere with cellular protein-protein interactions. Since both E1A and various cellular targets bind to a site in RB termed the pocket domain (2), E1A can competitively disrupt the complex formation between RB and its cellular targets.

The second cellular factor implicated in E1A-dependent transformation, p300, is believed to inhibit G0/G1 exit, to activate certain enhancers, and to stimulate differentiation (1,2). E1A inhibits the p300/CBP-mediated transcriptional activation of many promoters (14). In one case that has been examined, the complex of p300 and YY1, E1A inhibits transcription without disrupting the complex (10).

The present invention provides a cellular protein designated P/CAF which binds to p300/CBP and plays an important role in both transcription and cell cycle regulation associated with a histone acetyltransferase activity. The present invention also provides a histone acetyltransferase activity in the p300/CBP cellular protein, thus providing targets for modulating transcription and cell cycle regulation in cells.

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SUMMARY OF THE INVENTION



The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein.

The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided.

In addition, also provided is a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF and/or histone acetyltransferase activity, comprising contacting the substance with a system in which histone acetylation by P/CAF can be determined, determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

Furthermore, the present invention provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF comprising contacting the substance with a system in which the p300 binding of P/CAF can be determined; determining the amount of p300 binding of P/CAF in the presence of the substance, and comparing the amount of p300 binding of P/CAF in the presence of the substance with the amount of p300 binding of P/CAF in the absence of the substance, a decreased amount of p300 binding of P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

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Also provided is a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:3 under conditions whereby a P/CAF/p300 complex can be formed; and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample.

The present invention additionally provides a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/antibody complex can be formed; and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample.

Also provided herein is an assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of P/CAF, comprising: contacting the substance with a system in which histone acetylation by P/CAF can be determined, determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.

The present invention further provides an assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising: contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined; determining the amount of P/CAF binding of p300/CBP in the presence of the substance; and comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP.

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In addition, an assay is provided for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP, comprising: contacting the substance with a system in which histone acetylation by p300/CBP can be determined; determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP.

Furthermore, the present invention provides an assay for screening substances for the ability to inhibit binding of a DNA-binding transcription factor to p300/CBP comprising: contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined, determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.

A method is also provided for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

Also provided in the present invention is a method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

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Furthermore, the present invention provides a method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

Finally, the present invention additionally provides a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

BRIEF DESCRIPTION OF THE FIGURES

Figs. 1A-B. Fig 1A. P/CAF-p300/CBP interaction *in vivo*. Cell extract was immunoprecipitated with rabbit anti-P/CAF (lanes 1, 4, and 7), rabbit anti-CBP (lanes 2 and 5), and mouse anti-p300 (lane 9) antibodies. For controls, cell extract was precipitated with rabbit control IgG (lanes 3, 6, and 8) or mouse anti-HA monoclonal antibody (lane 10). The precipitates were analyzed by immunoblotting with anti-P/CAF (lanes 1-3), anti-CBP (lanes 4-6), and anti-p300 (lanes 7-10) antibodies. The positions of non-specific bands are indicated by asterisks. Fig. 1B. E1A inhibits the P/CAF-p300 interaction *in vivo*. Osteosarcoma cells were transfected with either control vector (lanes 1 and 4) or E1A- (lanes 2 and 5) or E1AΔN- (lanes 3 and 6) expression vectors. Extract from the transfected subpopulation was immunoprecipitated with anti-P/CAF (lanes 1-3) or control (lanes 4-6) IgG. The precipitates were analyzed by immunoblotting with anti-p300 and anti-P/CAF.

Figs. 2A-F. P/CAF and E1A mediate antagonistic effects on cell cycle progression. HeLa cells (ATCC accession number CCL 2) were transfected by electroporation with 7 μ g of P/CAF-expression plasmid and/or 3 μ g of the full-length or the N-terminally deleted ($\Delta 2$ -36) E1A 12S-expression plasmid as indicated in the figure. These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into



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pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1 μ g of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11 μ g. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 hours and subsequently labeled in medium containing 10 μ M bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32). Histograms show percentages of cells in G1 and S phases. Abscissa values represent fluorescence intensity of bound anti-BrdU antibodies in log scale.

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Fig. 3. Histone acetyltransferase activity of P/CAF. Activity of hGCN5 (lanes 1 and 4) and P/CAF (lanes 2 and 5) that acetylates free histones (lanes 1-3) or histones in the nucleosome core particle (35) (lanes 4-6) was measured as described (36). Each reaction contains 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. Note that the histone octamer dissociates into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE. The bands corresponding to acetylated histones H3 and H4 are indicated by arrows.

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DETAILED DESCRIPTION OF THE INVENTION

As used in the specification and in the claims, "a" can mean one or more, depending upon the context in which it is used.

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P/CAF protein and fragments

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones. The P/CAF protein can also bind to the amino acid region of SEQ ID NO 3

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(amino acid (aa) residues 1753 - 1966) of the cellular transcriptional factor, p300 (which has the complete amino acid sequence of SEQ ID NO:6 and the nucleotide sequence of SEQ ID NO:12), and the amino acid region of SEQ ID NO:6 (amino acid residues 1805 - 1854) of the cellular transcriptional factor, CBP (which has the complete amino acid sequence of SEQ ID NO:7 and the nucleotide sequence of SEQ ID NO:13). The P/CAF protein can be defined by any one or more of the typically used parameters. Examples of these parameters include, but are not limited to molecular weight (calculated or empirically determined), isoelectric focusing point, specific epitope(s), complete amino acid sequence, sequence of a specific region (e.g., N-terminus) of the amino acid sequence and the like.

For example, The P/CAF protein can consist of the amino acid sequence of SEQ ID NO:1 or the P/CAF protein can comprise the amino acid sequence of SEQ ID NO:2 which represents the carboxy terminal end of the P/CAF protein and contains the histone acetyltransferase activity, or the amino acid sequence of SEQ ID NO:4, which represents the amino terminal end of the P/CAF protein, containing the binding site for p300/CBP. Because the amino-terminal region is specific for P/CAF it can be used to define and identify P/CAF.

As used herein, "purified" refers to a protein (polypeptide, peptide, etc.) that is sufficiently free of contaminants or cell components with which it normally occurs to distinguish it from the contaminants or other components of its natural environment. The purified protein need not be homogeneous, but must be sufficiently free of contaminants to be useful in a clinical or research setting, for example, in an assay for detecting antibodies to the protein. Greater levels of purity can be obtained using methods derived from well known protocols. Specific methods for purifying P/CAF proteins are known in the art.

As will be appreciated by those skilled in the art, the invention also includes
those P/CAF polypeptides having slight variations in amino acid sequence which yield
polypeptides equivalent to the P/CAF protein defined herein. Such variations may arise

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naturally as allelic variations (e.g., due to genetic polymorphism) or may be produced by human intervention (e.g., by mutagenesis of cloned DNA sequences), such as induced point, deletion, insertion and substitution mutants. Minor changes in amino acid sequence are generally preferred, such as conservative amino acid replacements, small internal deletions or insertions, and additions or deletions at the ends of the molecules. Substitutions may be designed based on, for example, the model of Dayhoff, et al. (37). These modifications can result in changes in the amino acid sequence, provide silent mutations, modify a restriction site, or provide other specific mutations.

Modifications to any of the P/CAF proteins or fragments can be made, while preserving the specificity and activity (function) of the native protein or fragment thereof. As used herein, "native" describes a protein that occurs in nature. The modifications contemplated herein can be conservative amino acid substitutions, for example, the substitution of a basic amino acid for a different basic amino acid. Modifications can also include creation of fusion proteins with epitope tags or known recombinant proteins or genes encoding them created by subcloning into commercial or non-commercial vectors (e.g., polyhistidine tags, flag tags, myc tag, glutathione-S-transferase [GST] fusion protein, *xylE* fusion reporter construct). Furthermore, the modifications can be such as do not affect the function of the protein or the way the protein accomplishes that function (e.g., its secondary structure or the ultimate result of the protein's activity). These products are equivalent to the P/CAF protein. The means for determining the function, way and result parameters are well known.

enables the purification of P/CAF homologs from other species and allelic variants from individuals within a species. For example, an antibody raised against the exemplary human P/CAF protein can be used routinely to screen preparations from different humans for allelic variants of the P/CAF protein that react with the P/CAF protein-specific antibody. Similarly, an antibody raised against an epitope, for example, from a conserved amino acid region of the human P/CAF protein can be used to routinely screen for homologs of the P/CAF protein in other species. A P/CAF protein can be

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routinely identified in and obtained from other species and from individuals within a species using the methods taught herein and others known in the art. For example, given the present sequence, the DNA encoding a conserved amino acid sequence can be used to probe genomic DNA or DNA libraries of an organism to predictably obtain the P/CAF gene for that organism. The gene can then be cloned and expressed as the P/CAF protein and purified according to any of a number of routine, predictable methods. An example of the routine protein purification methods available in the art can be found in Pei et al. (38).

A purified polypeptide fragment of the P/CAF protein is also provided. The term "fragment" as used herein regarding a P/CAF protein, means a molecule of at least five contiguous amino acids of P/CAF protein that has at least one function shared by P/CAF protein or a region thereof. These functions can include antigenicity, binding capacity, acetyltransferase activity and structural roles, among others. The P/CAF fragment can be specific for a recited source. As used herein to describe an amino acid sequence (protein, polypeptide, peptide, etc.), "specific" means that the amino acid sequence is not found identically in any other source. The determination of specificity is made routine by the availability of computerized amino acid sequence databases and sequence comparison programs, wherein an amino acid sequence of almost any length can be quickly and reliably checked for the existence of identical sequences. If an identical sequence is not found, the protein is "specific" for the recited source. For example, a P/CAF fragment can be species-specific (e.g., found in the P/CAF protein of humans, but not of other species).

A fragment of the P/CAF protein having histone acetyltransferase activity can consist of the amino acid sequence of SEQ ID NO:2. A fragment of the P/CAF protein which binds to the amino acid sequence of SEQ ID NO:3 on p300 and the amino acid sequence of SEQ ID NO:9 on CBP can consist of the amino acid sequence of SEQ ID NO:4. To the extent that these fragments are specific for P/CAF, they can be used to identify and define P/CAF.

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methods.

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An antigenic fragment of P/CAF protein is provided. An antigenic fragment has an amino acid sequence of at least about five consecutive amino acids of a P/CAF protein amino acid sequence and binds an antibody or elicits an immune response in an animal. An antigenic fragment can be selected by applying the routine technique of epitope mapping to P/CAF protein to determine the regions of the proteins that contain epitopes reactive with antibodies or are capable of eliciting an immune response in an animal. Once the epitope is selected, an antigenic polypeptide containing the epitope can be synthesized directly, or produced recombinantly by cloning nucleic acids encoding the antigenic polypeptide in an expression system, according to standard

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Alternatively, an antigenic fragment of the antigen can be isolated from the whole P/CAF protein or a larger fragment of the P/CAF protein by chemical or mechanical disruption. Fragments can also be randomly chosen from a known P/CAF protein sequence and synthesized. The purified fragments thus obtained can be tested to determine their antigenicity and specificity by routine methods.

Nucleic Acids Encoding P/CAF Protein

An isolated nucleic acid that encodes a P/CAF protein is also provided. As used herein, the term "isolated" means a nucleic acid separated or substantially free from at least some of the other components of the naturally occurring organism, for example, the cell structural components commonly found associated with nucleic acids in a cellular environment and/or other nucleic acids. The isolation of nucleic acids can therefore be accomplished by techniques such as cell lysis followed by phenol plus chloroform extraction, followed by ethanol precipitation of the nucleic acids (39). It is not contemplated that the isolated nucleic acids are necessarily totally free of all nonnucleic acid components or all other nucleic acids, but that the isolated nucleic acids are isolated to a degree of purification to be useful in clinical, diagnostic, experimental, or other procedures such as, for example, gel electrophoresis, Southern, Northern or dot blot hybridization, or polymerase chain reaction (PCR).

A skilled artisan in the field will readily appreciate that there are a multitude of procedures which may be used to isolate the nucleic acids prior to their use in other procedures. These include, but are not limited to, lysis of the cell followed by gel filtration or anion exchange chromatography, binding DNA to silica in the form of glass beads, filters or diatoms in the presence of high concentrations of chaotropic salts, or ethanol precipitation of the nucleic acids.

The nucleic acids of the present invention can include positive and negative strand RNA as well as DNA and can include genomic and subgenomic nucleic acids found in the naturally occurring organism. The nucleic acids contemplated by the present invention include double stranded and single stranded DNA of the genome, complementary positive stranded cRNA and mRNA, and complementary cDNA produced therefrom and any nucleic acid which can selectively or specifically hybridize to the isolated nucleic acids provided herein. Stringent conditions (further described below) are used to distinguish selectively or specifically hybridizing nucleic acids from non-selectively and non-specifically hybridizing nucleic acids.

An isolated nucleic acid that encodes a P/CAF protein can be species-specific (i.e., does not encode the P/CAF protein of other species and does not occur in other species). Examples of the nucleic acids contemplated herein include the nucleic acid of SEQ ID NO:10 as well as the nucleic acids that encode each of the P/CAF proteins or fragments thereof described herein. P/CAF proteins and protein fragments can be routinely obtained as described herein and their structure (sequence) determined by routine means including the methods as used herein.

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P/CAF protein-encoding nucleic acids can be isolated from an organism in which they are normally found (e.g., humans), using any of the routine techniques. For example, a genomic DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest using one of the present P/CAF protein-encoding nucleic acids as a probe. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are

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commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers, which contain restriction sites, to the termini of the nucleic acid (See, for example, ref. 39).

P/CAF protein-encoding nucleic acids can also be synthesized. For example, a method of obtaining a DNA molecule encoding a specific P/CAF protein is to synthesize a recombinant DNA molecule which encodes the P/CAF protein. For example, nucleic acid synthesis procedures are routine in the art and oligonucleotides coding for a particular protein region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector.

Oligonucleotides complementary to or identical with the P/CAF proteinencoding nucleic acid sequence can be synthesized as primers for amplification reactions, such as PCR, or as probes to detect P/CAF protein encoding nucleic acids by various hybridization protocols (e.g., Northern blot; Southern blot; dot blot, colony screening, etc.)

Double-stranded molecules coding for relatively large proteins can readily be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein, followed by ligating these DNA molecules together. For example, Cunningham, et al. (40), have constructed a synthetic gene encoding the human growth hormone by first constructing overlapping and complementary synthetic oligonucleotides and ligating these fragments together. See also, Ferretti, et al. (41), wherein synthesis of a 1057 base pair synthetic bovine rhodopsin gene from synthetic oligonucleotides is disclosed.

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By constructing a P/CAF protein-encoding nucleic acid in this manner, one skilled in the art can readily obtain any particular P/CAF protein with modifications at any particular position or positions. See also, U.S. Patent No. 5,503,995 which describes an enzyme template reaction method of making synthetic genes. Techniques such as this are routine in the art and are well documented. DNA encoding the P/CAF protein or P/CAF protein fragments can then be expressed *in vivo* or *in vitro*.

The nucleic acid encoding the P/CAF protein can be any nucleic acid that functionally encodes the P/CAF protein. To functionally encode the protein (i.e., allow the nucleic acid to be expressed), the nucleic acid can include, but is not limited to, expression control sequences, such as an origin of replication, a promoter, regions upstream or downstream of the promoter, such as enhancers that may regulate the transcriptional activity of the promoter, appropriate restriction sites to facilitate cloning of inserts adjacent to the promoter, antibiotic resistance genes or other markers which can serve to select for cells containing the vector or the vector containing the insert, and necessary information processing sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites and transcription termination sequences as well as any other sequence which may facilitate the expression of the inserted nucleic acid.

Preferred expression control sequences are promoters derived from metallothionine genes, actin genes, immunoglobulin genes, CMV, SV40, adenovirus, bovine papilloma virus, etc. A nucleic acid encoding a P/CAF protein can readily be determined based upon the genetic code for the amino acid sequence of the P/CAF protein and many nucleic acid sequences will encode a P/CAF protein. Modifications in the nucleic acid sequence encoding the P/CAF protein are also contemplated. Modifications that can be useful are modifications to the sequences controlling expression of the P/CAF protein to make production of P/CAF protein inducible or repressible as controlled by the appropriate inducer or repressor. Such means are standard in the art (see, e.g., ref. 39). The nucleic acids can be generated by means standard in the art, such as by recombinant nucleic acid techniques, as exemplified in the examples herein, and by synthetic nucleic acid synthesis or *in vitro* enzymatic synthesis.

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After a nucleic acid encoding a particular P/CAF protein of interest, or a region of that nucleic acid, is constructed, modified, or isolated, that nucleic acid can then be cloned into an appropriate vector, which can direct the *in vivo* or *in vitro* synthesis of that wild-type and/or modified P/CAF protein. The vector is contemplated to have the necessary functional elements that direct and regulate transcription of the inserted nucleic acid, as described above. The vector containing the P/CAF nucleic acid or nucleic acid fragment can be in a host (e.g., cell or transgenic animal) for expressing the nucleic acid. The P/CAF protein or fragment thereof can thus be produced in a host system containing the expression vector and its functional activity as described herein can be demonstrated according to methods well known in the art.

There are numerous *E. coli* (*Escherichia coli*) expression vectors known to one of ordinary skill in the art useful for the expression of proteins. Other microbial hosts suitable for use include bacilli, such as *Bacillus subtilis*, and other enterobacteria, such as *Salmonella*, *Serratia*, as well as various *Pseudomonas* species. These prokaryotic hosts can support expression vectors which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (Trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence and have ribosome binding site sequences, for example, for initiating and completing transcription and translation. If necessary, an amino terminal methionine can be provided by insertion of a Met codon 5' and in-frame with the gene sequence. Also, the carboxy-terminal extension of the protein can be removed using standard oligonucleotide mutagenesis procedures.

Additionally, yeast expression can be used. There are several advantages to yeast expression systems. First, evidence exists that proteins produced in yeast secretion systems exhibit correct disulfide pairing. Second, post-translational glycosylation is efficiently carried out by yeast secretory systems. The *Saccharomyces cerevisiae* prepro-alpha-factor leader region (encoded by the $MF\alpha$ -I gene) is routinely used to direct

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protein secretion from yeast (42). The leader region of pre-pro-alpha-factor contains a signal peptide and a pro-segment which includes a recognition sequence for a yeast protease encoded by the KEX2 gene. This enzyme cleaves the precursor protein on the carboxyl side of a Lys-Arg dipeptide cleavage-signal sequence. The polypeptide coding sequence can be fused in-frame to the pre-pro-alpha-factor leader region. This construct is then put under the control of a strong transcription promoter, such as the alcohol dehydrogenase I promoter or a glycolytic promoter. The protein coding sequence is followed by a translation termination codon which is followed by transcription termination signals. Alternatively, the polypeptide encoding sequence of interest can be fused to a second protein coding sequence, such as Sj26 or β -galactosidase, used to facilitate purification of the resultant fusion protein by affinity chromatography. The

Efficient post-translational glycosylation and expression of recombinant proteins can also be achieved in *Baculovirus* expression systems in insect cells.

insertion of protease cleavage sites to separate the components of the fusion protein is

applicable to constructs used for expression in yeast.

Mammalian cells permit the expression of proteins in an environment that favors important post-translational modifications such as folding and cysteine pairing, addition of complex carbohydrate structures and secretion of active protein. Vectors useful for the expression of proteins in mammalian cells are characterized by insertion of the protein encoding sequence between a strong viral promoter and a polyadenylation signal. The vectors can contain genes conferring either gentamicin or methotrexate resistance for use as selectable markers. For example, the antigen and immunoreactive fragment coding sequence can be introduced into a Chinese hamster ovary (CHO) cell line using a methotrexate resistance-encoding vector. Presence of the vector RNA in transformed cells can be confirmed by Northern blot analysis and production of a cDNA or opposite strand RNA corresponding to the protein encoding sequence can be confirmed by Southern and Northern blot analysis, respectively. A number of other suitable host cell lines capable of secreting intact proteins have been developed in the art and include the CHO cell lines, HeLa cells, myeloma cell lines, Jurkat cells, and the like

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Expression vectors for these cells can include expression control sequences, as described above. The vectors containing the nucleic acid sequences of interest can be transferred into the host cell by well-known methods, which vary depending on the type of cell host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cell hosts.

Alternative vectors for the expression of protein in mammalian cells, similar to those developed for the expression of human gamma-interferon, tissue plasminogen activator, clotting Factor VIII, hepatitis B virus surface antigen, protease Nexin I, and eosinophil major basic protein, can be employed. Further, the vector can include CMV promoter sequences and a polyadenylation signal available for expression of inserted nucleic acid in mammalian cells (such as COS7).

The nucleic acid sequences can be expressed in hosts after the sequences have been positioned to ensure the functioning of an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors can contain selection markers, e.g., tetracycline resistance or hygromycin resistance, to permit detection and/or selection of those cells transformed with the desired nucleic acid sequences (see, e.g., U.S. Patent 4,704,362).

The nucleic acids produced as described above can also be expressed in a host which is a non-human animal to create a transgenic animal, containing, in a germ or somatic cell, a nucleic acid comprising the coding sequence for all or a portion of the P/CAF protein, as well as all of the other regulatory elements required for expression of the P/CAF protein-encoding sequence. The animal will express the P/CAF gene or portion thereof to produce the P/CAF protein or protein fragment and such expression can be detected by determination of a particular phenotype unique to the transgenic animal expressing the transferred nucleic acid.

The nucleic acid can be the nucleic acid of SEQ ID NO:10, a nucleic acid having a nucleotide sequence which encodes the P/CAF protein, a nucleic acid having a nucleotide sequence which encodes the protein of SEQ ID NO:1, as well as the nucleic acids that encode the proteins comprising the fragments of SEQ ID NOS:2 and 4.

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The nucleic acids of the invention can contain substitutions or deletions which provide a particular phenotype of interest. For example, various deletions or base substitutions can be introduced into the nucleic acid encoding the P/CAF protein for the purpose of studying the effects of these particular deletions or substitutions on the transcription modulation activity of the P/CAF protein. These effects can be monitored by observation of such characteristics as growth and development of the animal, the ability to develop tumors, survival rates and the like. The gene construct introduced into the animal cells to produce the transgenic animal can contain any of the regulatory elements described above to modulate expression of the foreign genes. As used herein, the term "phenotype" includes morphology, biochemical profiles, changes in tumor formation and other parameters that are affected by the presence of the P/CAF protein.

The transgenic animals of the invention can also be used in a method for determining the effectiveness of administering a nucleic acid encoding a functional P/CAF protein to a subject in need of a functional P/CAF protein. First, a nucleic acid encoding a nonfunctional P/CAF protein can be introduced into the animal's cells and expressed to yield a characteristic phenotype. Then, using standard gene therapy techniques, a nucleic acid encoding a functional P/CAF protein can be introduced into the animal's cells and the effects on the animal's phenotypic characteristics can be determined.

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Having provided and taught how to obtain a nucleic acid that encodes a P/CAF protein, an isolated nucleic acid that encodes a fragment of P/CAF protein is also provided. The nucleic acid encoding the fragment can be obtained using any of the methods applicable to the nucleic acid encoding the entire P/CAF protein. The nucleic acid fragment can encode a species-specific P/CAF protein fragment (e.g., found in the

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P/CAF protein of humans, but not in the P/CAF proteins of other species). Nucleic acids encoding species-specific fragments of P/CAF protein are themselves species-specific or allele-specific fragments of the P/CAF gene.

Examples of fragments of a nucleic acid encoding a fragment of the P/CAF protein can include the nucleic acid sequences which encode the amino acid sequences of the fragments of SEQ ID NOS:2 or 4. The same routine computer analyses used to select these examples of fragments can be routinely used to obtain others. Fragments of P/CAF-encoding nucleic acids can be primers for PCR or probes, which can be species-specific, gene-specific or allele-specific. P/CAF-encoding nucleic acid fragments can encode antigenic or immunogenic fragments of P/CAF protein that can be used in therapeutic assays or screening protocols. P/CAF gene fragments can encode fragments of P/CAF protein having histone acetylase activity and/or p300/CBP binding activity as described above, as well as other uses that may become apparent.

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An isolated nucleic acid of at least ten nucleotides that selectively hybridizes with the nucleic acid of SEQ ID NO:10 under selected conditions is provided. For example, the conditions can be PCR amplification conditions and the hybridizing nucleic acid can be a primer consisting of a specific fragment of the reference sequence or a nearly identical nucleic acid that hybridizes only to the exemplified P/CAF-encoding nucleic acid or allelic variants thereof.

The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF-encoding nucleic acid sequence of SEQ ID NO:10 under stringent conditions. The hybridizing nucleic acid can be a probe that hybridizes only to the exemplified P/CAF-encoding nucleic acid sequence. Thus, the hybridizing nucleic acid can be a naturally occurring species-specific allelic variant of the exemplified P/CAF gene. The hybridizing nucleic acid can also include insubstantial base substitutions that do not prevent hybridization under the stated stringent conditions or affect either the function of the encoded protein, the way the protein accomplishes that function (e.g., its

secondary structure) or the ultimate result of the protein's activity. The means for determining these parameters are well known.

As used herein to describe nucleic acids, the term "selectively hybridizes" excludes the occasional randomly hybridizing nucleic acids as well as nucleic acids that encode other known homologs of the P/CAF protein. The selectively hybridizing nucleic acids of the invention can have at least 70%, 73%, 78%; 80%, 85%, 88%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% complementarity with the segment and strand of the sequence to which it hybridizes. This list is not intended to exclude percent complementarity values between these values. The nucleic acids can be at least 10, 15, 16, 17, 18, 20, 21, 23, 24, 25, 30, 35; 40, 50, 100, 150, 200, 300, 500, 550, 750, 900, 950, or 1000 nucleotides in length or any intervening length, depending on whether the nucleic acid is to be used as a primer, probe or for protein expression. The hybridizing nucleic acid can comprise a region of at least ten nucleotides (up to full length) that is completely complementary to a unique region of the nucleic acid to which it hybridizes.

The nucleic acid can be an alternative coding sequence for the P/CAF protein, or can be used as a probe or primer for detecting the presence of or obtaining the P/CAF protein. If used as primers, the invention provides compositions including at least two nucleic acids which selectively hybridize with different regions of the nucleic acid so as to amplify a desired region. Depending on the length of the probe or primer, it can range between 70% complementary bases and full complementarity and still hybridize under stringent conditions.

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For example, for the purpose of obtaining or determining the presence of a nucleic acid encoding the P/CAF protein, the degree of complementarity between the hybridizing nucleic acid (probe or primer) and the sequence to which it hybridizes (P/CAF DNA in a sample) should be at least enough to exclude hybridization with a nucleic acid from another species. The invention provides examples of these nucleic acids of P/CAF, so that the degree of complementarity required to distinguish selectively



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hybridizing from nonselectively hybridizing nucleic acids under stringent conditions can be clearly determined for each nucleic acid. It should also be clear that the hybridizing nucleic acids of the invention will not hybridize with nucleic acids encoding unrelated proteins (hybridization is selective) under stringent conditions.

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"Stringent conditions" refers to the washing conditions used in a hybridization protocol. In general, the washing conditions should be a combination of temperature and salt concentration chosen so that the denaturation temperature is approximately 5-20°C below the calculated T_m of the nucleic acid hybrid under study. The temperature and salt conditions are readily determined empirically in preliminary experiments in which samples of reference DNA immobilized on filters are hybridized to the probe or protein encoding nucleic acid of interest and then washed under conditions of different stringencies. For example, the nucleic acid sequence of SEQ ID NO 10 was used as a specific radiolabeled probe for the detection of messenger RNA transcribed from the P/CAF gene by performing hybridizations under stringent conditions. The T_m of such an oligonucleotide can be estimated by allowing 2°C for each A or T nucleotide, and 4°C for each G or C. For example, an 18 nucleotide probe of 50% G+C would, therefore, have an approximate T_m of 54°C.

The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF gene shown in the sequence set forth as SEQ ID NO:10 under stringent conditions. The invention further provides an isolated nucleic acid complementary to the nucleotide sequence set forth in SEQ ID NO:10.

25 Antibodies to the P/CAF protein

A purified antibody and an antiserum containing polyclonal antibodies that specifically bind the P/CAF protein or antigenic fragment are also provided. The term "bind" means the well understood antigen/antibody binding as well as other nonrandom association with an antigen. "Specifically bind" as used herein describes an antibody or other ligand that does not cross react substantially with any antigen other than the one specified, in this case, an antigen of the P/CAF protein. Antibodies can be made as

described in Harlow and Lane (33). Briefly, purified P/CAF protein or an antigenic fragment thereof can be injected into an animal in an amount and in intervals sufficient to elicit a humoral immune response. Serum polyclonal antibodies can be purified directly, or spleen cells from the animal can be fused with an immortal cell line and screened for monoclonal antibody secretion, according to procedures well known in the art. Purified monospecific polyclonal antibodies that specifically bind the P/CAF antigen are also within the scope of the present invention. The antibodies of the present invention can bind the protein of claim 1, the protein of claim 2, the protein of claim 3 and/or the protein of claim 4, as well as any other proteins of the present invention.

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A ligand that specifically binds the antigen is also contemplated. The ligand can be a fragment of an antibody, such as, for example, an Fab fragment which retains P/CAF binding activity, or a smaller molecule designed to bind an epitope of the P/CAF antigen. The antibody or ligand can be bound to a substrate or labeled with a detectable moiety or both bound and labeled. The detectable moieties contemplated within the compositions of the present invention include those listed above in the description of the diagnostic methods, including fluorescent, enzymatic and radioactive markers

The antibody can be bound to a solid support substrate or conjugated with a

detectable moiety or therapeutic compound or both bound and conjugated. Such
conjugation techniques are well known in the art. For example, conjugation of
fluorescent, radioactive or enzymatic moieties can be performed as described in the art
(33,43). The detectable moieties contemplated in the present invention can include
fluorescent, radioactive and enzymatic markers and the like. Therapeutic drugs
contemplated with the present invention can include cytotoxic moieties such as ricin A
chain, diphtheria toxin, pseudomonas exotoxin and other chemotherapeutic compounds.

It is well understood by one of skill in the art that all of the above discussion regarding antibodies to P/CAF can also be applied with regard to production, characterization and use of antibodies which bind the p300/CBP protein or any of the DNA-binding transcription factors of this invention.

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Measuring the P/CAF protein in a sample

The present invention also provides a method for determining the presence and thus the amount of P/CAF protein in a biological sample. As used herein, a biological sample includes any tissue or cell which would contain the P/CAF protein. Examples of cells include tissues taken from surgical biopsies, isolated from a body fluid or prepared in an *in vitro* tissue culture environment.

One example of determining the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:3 under conditions whereby a P/CAF/p300 complex can be formed; and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/p300 complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the p300 protein or by the detection of an antibody which binds p300 or the P/CAF protein, as taught in the Examples herein. Antibodies which bind p300 or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/p300 complexes by the detection of the binding of antibodies reactive with p300 or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as described below.

Alternatively, determination of the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:9 under conditions whereby a P/CAF/CBP complex can be formed, and determining the amount of the P/CAF/CBP complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/CBP complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the CBP protein or by the detection of an antibody which binds either CBP or the P/CAF protein, as taught in the Examples

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herein. Antibodies which bind CBP or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/CBP complexes by the detection of the binding of antibodies reactive with CBP or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as described below.

Another example of determining the amount of P/CAF in a biological sample comprises contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/ antibody complex can be formed and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample. Antibodies which bind P/CAF can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Determination of P/CAF/antibody complexes can be accomplished using various immunoassays as are available in the art, as described below.

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Immunoassays such as immunofluorescence assays, radioimmunoassays (RIA), immunoblotting and enzyme linked immunosorbent assays (ELISA) can be readily adapted for detection and measurement of P/CAF in a biological sample. Both polyclonal and monoclonal antibodies can be used in the assays. Available immunoassays are well known in the art and are extensively described in the patent scientific literature. See, for example, U.S. Patent Nos. 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; and 4,098,876.

25 Screening assays for P/CAF

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur, determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the

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amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.

The present invention also provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF, comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur, determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and cell cycle progression suppressing activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the transcription modulating activity of P/CAF by interfering with the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.



Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 and P/CAF can occur; determining the amount of p300 binding to P/CAF in the presence of the substance; and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, a decreased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO:3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

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Additionally provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur; determining the amount of CBP binding to P/CAF in the presence of the substance; and comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, a decreased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid sequence of SEQ ID NO:9 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further contemplates a bioassay for screening substances for the ability to stimulate the transcription modulating activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the transcription modulating activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples.

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Thus, the compound to be tested for the ability to stimulate the transcription modulating activity of P/CAF by increasing the histone acetyltransferase activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 to P/CAF can occur; determining the amount of p300 binding to P/CAF in the presence of the substance; and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, an increased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO:3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

Additionally provided in the present invention is a bioassay for screening substances for the ability to stimulate the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur, determining the amount of CBP binding to P/CAF in the presence of the substance; and 25 comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, an increased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid 30 sequence of SEQ ID NO:9 and P/CAF. Alternatively, the system can comprise a cell



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extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

Transcription modulating activity of P/CAF

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The present invention contemplates a method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. For example, the substance can be identified according to the protocols provided herein as one that can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP or by inhibiting the histone acetyltransferase activity of P/CAF as well as by any other inhibitory mechanism as identified by the protocols provided herein. Inhibition of the transcription modulating activity of P/CAF in a subject is desirable, for example, to inhibit HIV TAT-mediated transcription and therefore, the method of the present invention can be used to treat HIV-infected subjects.

The substance can be in a pharmaceutically acceptable carrier. By "pharmaceutically acceptable" is meant a material that is not biologically or otherwise undesirable, i.e., the material may be administered to a subject, along with the substance, without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the pharmaceutical composition in which it is contained. The carrier would naturally be selected to minimize any degradation of the active ingredient and to minimize any adverse side effects in the subject.

The transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the P/CAF binding site or a substance which binds the P/CAF protein at the p300/CBP binding site, the ultimate result being that P/CAF and p300/CBP do not bind with one another and P/CAF cannot exert its transcription modulating and/or histone acetyltransferase effect. The substance can be a protein, such as an antibody which binds the P/CAF protein binding site at or near the p300/CBP

binding site, thereby preventing its binding or an antibody which binds the p300/CBP protein at or near the P/CAF binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on P/CAF or at the acetylation site on the histone, thereby preventing acetylation by P/CAF.

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The substance which binds p300/CBP, the P/CAF protein or the histone and has the net effect of inhibiting the transcription modulating effect and or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by mechanisms well known in the art.

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Alternatively, a nucleic acid encoding a protein which binds either to p300/CBP or the P/CAF protein and has the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors, as described below.

The substance which inhibits the transcription modulating effect and/or histone acetyltransferase activity of P/CAF can be an antisense RNA or an antisense DNA which binds the RNA or DNA of P/CAF, thereby preventing translation or transcription of the RNA or DNA encoding P/CAF and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF by inhibiting P/CAF production. The antisense RNA of the present invention can be generated from the nucleic acid of SEQ ID NO:14 (human) or SEQ ID NO:15 (mouse). Furthermore, the 25 antisense DNA can be a phosphorothioate oligodeoxyribonucleotide having the nucleotide sequence of SEQ ID NO:16 (human) or of SEQ ID NO:17 (mouse). The mouse antisense RNA can be used to inhibit the activity of mouse P/CAF, having the nucleotide sequence of SEQ ID NO:18 and the amino acid sequence of SEQ ID NO:8. The present invention also contemplates an antisense nucleic acid sequence which can bind the DNA or RNA of any of the transcription factors or other proteins now known or later identified to bind P/CAF, thereby inhibiting expression of the gene products of



these proteins and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF.

The antisense nucleic acid can comprise a typical nucleic acid, but the antisense nucleic acid can also be a modified nucleic acid or a derivative of a nucleic acid such as a phosphorothioate analogue of a nucleic acid. The composition can comprise, for example, an antisense RNA that specifically binds an RNA encoded by the gene encoding the serum protein. Antisense RNAs can be synthesized and used by standard methods (62).

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Antisense RNA can inhibit gene expression by forming an RNA/RNA duplex between the antisense RNA and the RNA transcribed from the target gene. The precise mechanism by which this duplex formation decreases the production of the protein encoded by the endogenous gene probably involves binding of complementary regions of the normal sense mRNA and the antisense RNA strand with duplex formation in a manner that blocks RNA processing and translation. Alternative mechanisms include the formation of a triplex between the antisense RNA and duplex DNA or the formation of an DNA-RNA duplex with subsequent degradation of DNA-RNA hybrids by RNAse H. Furthermore, an antigene effect can result from certain DNA-based oligonucleotides via triple-helix formation between the oligomer and double-stranded DNA which results in the repression of gene transcription. Regardless of the specific molecular mechanism, the present invention results in inhibition of expression of the P/CAF gene by the introduced and replicated DNA resulting in inhibition of the transcription modulating and/or histone acetyltransferase activity of P/CAF, by a reduction in the expression of the nucleic acid to which the antisense nucleic acid is hybridized, and therefore a reduction of the gene product from the targeted gene.

The antisense nucleic acid may be obtained by any number of techniques known to one skilled in the art. One method of constructing an antisense nucleic acid is to synthesize a recombinant antisense DNA molecule. For example, oligonucleotide synthesis procedures are routine in the art and oligonucleotides coding for a particular

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routine in the art and are well documented.

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protein or regulatory region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector. Double-stranded molecules coding for relatively large proteins or regulatory regions can be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein or regulatory region, followed by ligating these DNA molecules together. Once the appropriate DNA molecule is synthesized, this DNA can be cloned downstream of a promoter in an antisense orientation. Techniques such as this are

An example of another method of obtaining an antisense nucleic acid is to isolate that nucleic acid from the organism in which it is found and clone it in an antisense orientation. For example, a DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector in an antisense orientation, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers which contain restriction sites to the termini of the nucleic acid. General methods are set forth in Sambrook et al. (39).

The DNA that is introduced into the cell is in an expression orientation that is antisense to a corresponding endogenous DNA or RNA of the cells. For example, where an endogenous DNA comprises a gene which encodes for a particular protein, the introduced DNA is in an expression orientation opposite the expression of the endogenous DNA; that is the DNA operatively linked to a promoter is in an antisense expression orientation relative to the corresponding endogenous gene. The introduced DNA may be homologous to the entire transcribed gene or homologous to only part of

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the transcribed gene. Alternatively, the sequence of the introduced DNA may be divergent to that of the endogenous DNA but only divergent to the extent that hybridization of the nucleic acids occurs, thereby preventing transcription. One skilled in the art can determine the maximum extent of this divergence by routine screening of antisense DNAs corresponding to an endogenous DNA of the cell. In this manner, one skilled in the art can readily determine which fragments, or alternatively the extent of homology of the fragments or the entire gene that is necessary to inhibit gene expression.

The antisense nucleic acids of the present invention can be made according to protocols standard in the art, as well as described in the Examples provided herein. The antisense nucleic acids can be administered to a subject according to the gene transduction protocols standard in the art, as described below.

The present invention also contemplates a method for stimulating the transcription modulating activity and/or histone acetyltransferase activity of P/CAF in a subject comprising administering to the subject a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a stimulatory affect on the transcription modulating and/or histone acetyltransferase activity of P/CAF. The substance can be one which has been identified, according to the protocols provided herein, to stimulate histone acetyltransferase activity in P/CAF or promote binding of P/CAF to p300/CBP. The stimulation of the transcription modulation activity and/or histone acetyltransferase activity of P/CAF in a subject is desirable, for example, to activate tumor suppressor p53 (which promotes apoptosis) or to activate the muscle differentiation factor, MyoD. Thus, the method of the present invention can be employed to treat cancer and to promote muscle differentiation in conditions where muscle differentiation is desired. The substance can be delivered to a cell in the subject by mechanisms well known in the art.

Further contemplated in the present invention is a method for promoting binding of P/CAF to p300/CBP in a subject, comprising administering to the subject a substance

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identified by the methods provided herein to promote binding of P/CAF to either p300 or CBP.

Additionally, a nucleic acid encoding a protein which stimulates the transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

Also provided in the present invention is a method of inhibiting the cell cycle progression inducing effect of an oncoprotein which binds p300/CBP in a subject 10 comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein, inducing expression of the nucleic acid in the cell to produce the P/CAF in an amount which will allow the P/CAF gene product to replace the oncoprotein bound to p300/CBP, whereby the replacement of the oncoprotein bound to p300/CBP by the P/CAF gene product inhibits the cell cycle progression inducing effect of the oncoprotein. The oncoprotein which binds p300/CBP in the cell can be the adenovirus E1A oncoprotein.

A method for providing a functional P/CAF protein to a subject in need of the functional P/CAF protein is also provided, comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein and inducing expression of the nucleic acid to produce the functional P/CAF protein in the cell, thereby providing the functional P/CAF protein to the subject. The transduction of the vector nucleic acid into the subject's cells can be carried out according to standard gene therapy protocols well known in the art (see, for example, U.S. Patent No. 5,339,346).

Screening assays for p300/CBP

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance under conditions whereby histone acetylation by p300/CBP can occur;

determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for acetyltransferase inhibiting ability.

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Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of a transcriptional factor to p300/CBP, comprising contacting a system in which the binding of a transcriptional factor to p300/CBP can be determined, with the substance under conditions whereby the binding of the transcriptional factor and p300/CBP can occur; determining the amount of transcriptional factor binding to p300/CBP in the presence of the substance, and comparing the amount of transcriptional factor binding to p300/CBP in the presence of the substance with the amount of transcriptional factor binding to p300/CBP in the absence of the substance, a decreased amount of transcriptional factor binding to p300/CBP in the presence of the substance indicating a substance that can inhibit the binding of a transcriptional factor to p300/CBP. The binding of a transcriptional factor to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising a transcriptional factor which binds p300/CBP and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both a transcriptional factor which binds p300/CBP and p300/CBP. The transcriptional factor which binds p300/CBP can be selected from, but is not limited to



the group consisting of nuclear hormone receptors, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YYI, Sap-1a, c-Fos, MyoD and SRC-1, as well as any other transcriptional factor now known or later identified to bind p300/CBP. The screening assay of the present invention can also be used to identify substances which inhibit the binding of p300/CBP to other components to which it is known to bind, for example, P/CAF, pp90_{RSK}, TFIIB, E1A, SV40 large T antigen, as well as any other substances now known or later identified to bind p300/CBP. Determination of the binding of a transcriptional factor or other substance to p300/CBP can be carried out as taught in the Examples herein as well as by protocols described in the literature.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance, determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, an increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the p300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for stimulating ability

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of a component, which binds p300/CBP, to p300/CBP, comprising contacting a system in which the binding of the component to p300/CBP can



be determined, with the substance under conditions whereby the binding of the component to p300/CBP can occur; determining the amount of component binding to p300/CBP in the presence of the substance, and comparing the amount of component binding to p300/CBP in the presence of the substance with the amount of component binding to p300/CBP in the absence of the substance, an increased amount of component binding to p300/CBP in the presence of the substance indicating a substance that can stimulate the binding of the component to p300/CBP. The binding of the component to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising the component and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both the component and p300/CBP. The component which binds p300/CBP can be any of the transcriptional factors or other proteins which are known or are identified in the future to bind p300/CBP, as set forth above. Determination of the binding of the component to p300/CBP can be carried out as taught in the Examples provided herein and according to protocols available in the literature.

Histone acetyltransferase activity of p300/CBP

A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject is provided in the present invention, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. The mechanism of the inhibitory action of the substance can be the inhibition of the binding of a DNA-binding transcription factor, such as, for example, a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD or SRC-1, to p300/CBP.

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The histone acetyltransferase activity of p300/CBP can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the transcription factor binding site or a substance which binds the transcription factor protein at the p300/CBP binding site, the ultimate result being that the transcription factor and p300/CBP do not bind with one another and p300/CBP cannot acetylate histones.

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The substance which binds either to the transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be identified according to the screening methods provided herein and delivered to a cell in the subject by mechanisms well known in the art. The substance can be a protein, such as an antibody which binds the p300/CBP protein binding site at or near the DNA-binding transcription factor binding site, thereby preventing its binding or an antibody which binds the DNA-binding transcription factor at or near the p300/CBP binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on p300/CBP (aa 1195-1673 on p300 or aa 1174-1850 on CBP) or at the acetylation site on the histone, thereby preventing acetylation by p300/CBP.

Additionally, the substance can be a nucleic acid which can be expressed in the cell to produce a protein which inhibits the histone acetyltransferase activity of p300/CBP. For example, a nucleic acid encoding a protein which binds either to a transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors, as described below.

The substance which inhibits the histone acetyltransferase activity of p300/CBP can be an antisense RNA or an antisense DNA which binds the RNA or DNA of p300/CBP thereby preventing translation or transcription of the RNA or DNA encoding p300/CBP and having the net effect of inhibiting the histone acetyltransferase activity of P/CAF by inhibiting p300/CBP production. The antisense RNA or DNA of the present invention can be produced and introduced into cells according to the same methods as set forth above for P/CAF antisense nucleic acids.

The present invention also contemplates a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject comprising administering to the

subject a histone acetyltransferase activity stimulating amount of a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a stimulatory affect on the histone acetyltransferase activity of p300/CBP. The substance can exert a stimulatory effect by promoting the binding of a DNA-binding transcription factor of the present invention to p300/CBP. The substance can be delivered to a cell in the subject by mechanisms well known in the art. A nucleic acid encoding a protein which stimulates the transcription modulating activity of p300/CBP can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

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Gene transduction

In the methods described above which include gene transduction into cells (i.e., addition of exogenous DNA into cells), the nucleic acids of the present invention can be in a vector for delivering the nucleic acids to the site for expression of the P/CAF protein. The vector can be one of the commercially available preparations, such as the pGM plasmid (Promega). Vector delivery can be by liposome, using commercially available liposome preparations or newly developed liposomes having the features of the present liposomes. Additionally, vector delivery can be via a viral system, including, but not limited to, retroviral, adenoviral and adeno-associated viral systems. Other delivery methods can be adopted and routinely tested according to the methods taught herein.

The modes of administration of the liposome will vary predictably according to the disease being treated and the tissue being targeted. For example, for treating cancer in either the lung or the liver, which are both sinks for liposomes, intravenous delivery is reasonable. For other localized cancers, as well as precancerous conditions, catheterization of an artery upstream from the target organ is a preferred mode of delivery, because it avoids significant clearance of the liposome by the lung and liver. For cancerous lesions at a number of other sites (e.g., skin cancer, localized dysplasias), topical delivery is expected to be effective and may be preferred, because of its convenience.

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Leukemias and other disorders involving dysregulated proliferation of certain isolatable cell populations may be more readily treated by ex vivo administration of the nucleic acid.

The liposomes may be administered topically, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally or the like, although intravenous or topical administration is typically preferred. The exact amount of the liposomes required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the severity of the disease being treated, the particular compound used, its mode of administration and the like. Thus, it is not possible to specify an exact amount. However, an appropriate amount may be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein.

Parenteral administration, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant level of dosage is maintained. See, e.g., U.S. Patent No. 3,610,795, which is incorporated by reference herein.

Topical administration can be by creams, gels, suppositories and the like. Ex vivo (extracorporeal) delivery can be as typically used in other contexts.

The present invention is more particularly described in the following examples which are intended as illustrative only since numerous modifications and variations

therein will be apparent to those skilled in the art.

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EXAMPLES

I. P/CAF studies.

5 Cloning and characterization of P/CAF protein.

In human cells, CBP binds to c-Jun in a phosphorylation-dependent manner in association with stimulation of transcription (9). In yeast, GCN4 is believed to be a c-Jun counterpart on the basis of similarities in DNA recognition (15) as well as the participation of both proteins in UV signaling pathways (16). Yeast genetic screening has led to the isolation of various cofactors for GCN4, including GCN5 (yGCN5), ADA2 (yADA2) and ADA3 (yADA3) (17-19). These factors are considered to function as a complex (or in a common pathway) based on genetic and protein-protein interaction studies (18-22). Finally, p300/CBP and yADA2 exhibit significant sequence similarity within a 50 amino acid region including a Zn²⁺ finger motif (3). Human counterparts to yGCN5, yADA2, or yADA3 that interact with p300/CBP to mediate transcriptional activation by c-Jun were searched for in various nucleotide sequence databases.

Comparison of the yGCN5 protein sequence with various databases (23) revealed significant similarities with the two randomly sequenced human cDNAs, ETS05039 (24) (P=4.0x10⁻¹⁵) and NIB2000-5R (P=6.5x10⁻⁹). Given that these cDNAs were truncated, human fetal liver and fetal brain cDNA libraries (Clontech) were screened with ETS05039 and NIB2000-5R, respectively and complete clones were isolated from the human fetal liver cDNA library. The complete sequences revealed that the ETS05039- and NIB2000-5R-derived clones are encoded by distinct genes but are highly related within the protein coding regions (68% identity at the DNA level, 75% identity and 86% similarity at the protein level). The former encodes an N-terminal region with no sequence similarity to any proteins in the databases besides the yGCN5-related C-terminal region, whereas the latter encodes only the yGCN5-related region. Given that p300/CBP-binding activity was observed in the former polypeptide as shown below, it was designated p300/CBP-associated factor (P/CAF), having the amino acid

sequence of SEQ ID NO:1 and the nucleotide sequence of SEQ ID NO:10 and the latter was named human GCN5 (hGCN5), having the amino acid sequence of SEQ ID NO:5 and the nucleotide sequence of SEQ ID NO:11.

Additionally, an RNA blot (Clontech) was hybridized with a random-primed probe made from the cDNA encoding P/CAF. RNA blotting indicated that transcripts detected by the P/CAF and hGCN5 cDNAs are ubiquitously expressed, but the former is most abundant in heart and skeletal muscle, whereas the latter is most abundant in pancreas and skeletal muscle.

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P/CAF-p300/CBP interaction in vitro

The P/CAF binding site was presumed to reside in the C terminal one third of CBP (residues 1,678-2,442) because it was observed that this region, when fused to a DNA binding domain, activates transcription (4) in a manner repressed by coexpression of 12S E1A. This region was divided into 6 overlapping fragments and each was expressed in *E. coli* as a glutathione-S-transferase (GST) fusion protein. GST-CBP fusions were incubated with recombinant P/CAF protein and, subsequently, purified using glutathione-Sepharose. Co-purified P/CAF was detected by immunoblotting analysis.

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To construct GST-fusions, various regions of CBP and p300 were amplified by PCR. A series of deletions of the CBP segment B was created by site-directed *in vitro* mutagenesis (30). These fragments were subcloned into pGEX-2T (Pharmacia). GST-fusions were expressed in *E. coli* and extracted with buffer B [20 mM Tris-HCl (pH 8.0), 5 mM MgCl₂, 10% glycerol, 1 mM AEBSF, 0.1% NP40, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 1 mM DTT] containing 0.1 M KCl for these experiments. GST-CBP-segment B was purified by glutathione-Sepharose and phenyl-Sepharose chromatographic steps, P/CAF, hGCN5, and E1A were expressed as FLAG-fusions in Sf9 cells via baculovirus vectors and affinity-purified with M2-agarose (ref. 30, Kodak-IBI). For interaction, a crude *E. coli* extract containing 20 pmol of GST-fusion was incubated with 40-60 pmol of P/CAF or E1A in a total volume of 50 μl of

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buffer B with 0.1 M KCl on ice for 10 min. Samples were further incubated with 10 µl (packed volume) of glutathione-Sepharose at 4°C for 30 min, washed four times with 200 µl of buffer B containing 0.1 M KCl, and eluted with 20 µl of buffer E [50 mM Tris-HCl (pH 8.0), 0.2 M KCl, 20 mM glutathione] for 60 min. Interacting proteins were detected by anti-FLAG immunoblotting or silver staining.

For p300 interactions, the segment spanning residues 1763-1966 (segment B') of p300, which is analogous to the CBP segment-B, was used. Twenty percent of the P/CAF and hGCN5 inputs and 100% of the E1A input were also analyzed. In the GST precipitation assays, almost identical amounts of the GST fusions were recovered in all samples. Interaction between P/CAF and CBP (segment B) was determined in the absence and in the presence of E1A. Control reactions with GST-CBP alone and without GST-CBP were also performed. Input proteins were analyzed.

Two CBP segments, A and B, interacted specifically with P/CAF. The stronger interaction was observed in the latter segment, which does not include the yADA2-like Zn²⁺ finger. Given that the CBP segment-B is well conserved in p300 (66% identity, 75% similarity), the binding of P/CAF to p300 *in vitro* was also analyzed. For this experiment, the p300 segment spanning residues 1763-1966, termed segment B', which is analogous to the CBP segment-B, was used. Like CBP, p300 interacted specifically with P/CAF. These studies demonstrated that P/CAF binds specifically to both p300 and CBP *in vitro*. In contrast to P/CAF, hGCN5 did not bind to CBP or p300.

These studies also demonstrated that the Zn²⁺ finger region of p300/CBP, which shares sequence similarity with yADA2, is not essential for the interaction with P/CAF. Cloning of a human structural homolog of yADA2, termed hADA2 (25) has revealed that, unlike the sequence similarity between p300/CBP and yADA2, which is restricted to a 50 amino acid region, hADA2 shares extensive similarity (30% identity, 52% similarity) to yADA2 over the entire protein sequence. Moreover, a computer search of the complete genomic sequence of *Saccharomyces cerevisiae* revealed that yeast does



not have counterparts of p300/CBP or P/CAF. Thus, the p300/CBP-P/CAF pathway may have been acquired during metazoan evolution.

Action of E1A in vitro 5

Previous reports indicated that E1A binds to both the p300 segment spanning residues 1767-1816 and the CBP segment spanning residues 1805-1854 (7). These interactions were reconfirmed in the present system; thus, both p300 and CBP segments covering the previously identified regions interacted with E1A.

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For further mapping, a series of deletions was introduced within the CBP segment-B and tested for interactions with P/CAF and E1A. Deletions of residues 1801-1825 or 1824-1851 markedly reduced interactions with both P/CAF and E1A, whereas deletion of residues 1850-1878 did not affect these interactions. Furthermore, deletion of residues 1801-1851 completely abolished interactions with both P/CAF and E1A. These data indicate that residues 1801-1851 of CBP are critical for interaction with both P/CAF and E1A. Taken together with the evidence that CBP segment A (aa residues 1,678-1,880) also binds to these factors, the above findings demonstrate that P/CAF and E1A bind to the same or very closely spaced sites on CBP.

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Evidence that both P/CAF and E1A recognize the same p300/CBP segments raises the possibility of direct competition between P/CAF and E1A for binding to p300/CBP. To test this possibility, a competition experiment was performed with the use of affinity purified recombinant proteins. The interaction of P/CAF with the CBPsegment B was progressively inhibited by the addition of increasing amounts of E1A. In contrast, no inhibition was caused by an E1A mutant which does not bind to p300/CBP (E1AΔN). Similar results were obtained with the p300-segment B', leading to the conclusion that P/CAF and E1A compete for the same binding sites in p300/CBP.

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P/CAF-p300/CBP interaction in vivo

The *in vivo* interaction between P/CAF and p300/CBP was established by co-immunoprecipitation from a human osteosarcoma cell extract. Proteins in this extract were immunoprecipitated with rabbit anti-P/CAF, rabbit anti-CBP and anti-p300 antibodies. For controls, cell extract was precipitated with rabbit control IgG or mouse anti-HA monoclonal antibody. The precipitates were analyzed by immunoblotting with anti-P/CAF, anti-CBP and anti-p300 antibodies.

Osteosarcoma cells were transfected with either control vector or E1A- or

E1AΔN-expression vectors. Extract from the transfected subpopulation was immunoprecipitated with anti-P/CAF or control IgG. The precipitates were analyzed by immunoblotting with anti-p300 and anti-P/CAF antibodies.

Rabbit anti-P/CAF antibody was raised to the P/CAF segment spanning residues 125-397 and purified by immunoaffinity chromatography (33). A mixture of 15 monoclonal antibodies raised to the human p300 segment spanning residues 1572-2371 (5) and rabbit polyclonal antibodies raised to the mouse CBP segment spanning residues 2-23 (for immunoprecipitation) and 1736-2179 (immunoblotting) were purchased from Upstate Biotechnology. Approximately 2 x 10⁷ human osteosarcoma U-2 OS cells (ATCC accession number HTB 96) were extracted with 10 ml of lysis buffer [25 mM 20 HEPES-KOH (pH 7.2), 150 mM potassium acetate, 2 mM EDTA, 1 mM DTT, 1 mM AEBSF, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 20 mM sodium fluoride, 0.1% NP40]. Two to 10 ml of extract were incubated with 2 µg of the respective antibody for four hours at 4°C. Fifty µl (packed volume) of protein-A Trisacryl (Pierce) were added and incubation was continued for two hours. The matrix was washed four times with 1 ml of the lysis buffer, then boiled in 2x SDS sample buffer. Human osteosarcoma U-2 OS cells were transfected with 20 µg of the indicated plasmid and 1 µg of sorting plasmid (pCMV-IL2R) (3.1). The transfected subpopulation was purified by magnetic affinity cell sorting (32). Extract from approximately 2 x 10⁵ sorted cells was immunoprecipitated as described. 30

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Anti-P/CAF antibody specifically detected a 95 kDa protein, which is very close to the calculated value for the full-length P/CAF, in the immunoprecipitates. Anti-P/CAF antibody co-immunoprecipitated both CBP and p300. Similarly, anti-CBP antibody also co-immunoprecipitated P/CAF. However, anti-p300 antibody did not co-immunoprecipitate P/CAF. This is most likely due to steric interference since the anti-p300 antibody was raised to the p300 segment spanning residues 1572-2371 which includes the P/CAF binding region. These data demonstrate that P/CAF forms complexes with both p300 and CBP *in vivo*.

10 Action of E1A in vivo

The *in vitro* experiments described herein indicate that P/CAF and E1A compete for the binding sites in p300/CBP. Thus, a study was conducted to determine whether E1A targets the endogenous interaction between P/CAF and p300. An E1A-expression vector was transiently transfected into human osteosarcoma cells and the transfected subpopulation was purified by cell sorting. Then, the interaction between P/CAF and p300 in transfected cells was examined by co-immunoprecipitation with anti-P/CAF antibody. The endogenous interaction of P/CAF with p300 was drastically inhibited by expression of E1A. On the other hand, no inhibition was observed by the E1A mutant lacking the p300 binding domain (E1AΔN), indicating that E1A disrupts the P/CAF-p300 complex *in vivo* through an interaction with p300.

'Cell cycle regulation by P/CAF

Given that binding of P/CAF to p300/CBP is inhibited by E1A, experiments were performed to evaluate whether P/CAF, by binding to and forming a functional complex with p300, is involved in the regulation of entry into S phase. This possibility was addressed by examining whether transient expression of P/CAF would affect the rate of G1/S transit in HeLa cells. P/CAF negatively affected the distribution of cells between G1 and S phases in this assay.

HeLa cells were transfected by electroporation with 7 μg of P/CAF-expression plasmid and/or 3 μg of the full-length or the N-terminally deleted (Δ2-36) E1A 12S-



expression plasmid as indicated. These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1 µg of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11 µg. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 h, and subsequently labeled in medium containing 10 µM bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32).

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The fraction of cells accumulating in S phase in control cultures was 23%, compared to 15% in P/CAF-transfected cells. This effect was reproducible in multiple independent experiments. In parallel experiments to verify the utility of this experimental protocol, plasmids encoding E2F-1, simian virus 40 small t, cyclin A or cyclin E increased the accumulation of cells in S phase, whereas plasmids encoding the cyclin-dependent kinase inhibitors p21 or p27 reduced the number of S phase cells.

On the basis of evidence that E1A and P/CAF compete for binding sites on p300, it seemed possible that cotransfection of P/CAF with E1A would oppose the mitogenic effect caused by E1A. As shown by the data herein, this is indeed the case. E1A alone has mitogenic activity in this experimental setting, while the E1A mutant lacking the p300 binding domain (E1AΔN) has very weak activity. Comparable expression levels between wild type and mutant E1A in the transfected cells were revealed by immunoblotting analysis with anti-E1A. Intriguingly, when P/CAF was cotransfected with E1A, the mitogenic activity of E1A was significantly counteracted by P/CAF. These results show that P/CAF and E1A mediate antagonistic effects on cell cycle progression.

In the course of assessing P/CAF activity, it was also revealed that p300 is able to inhibit cell cycle progression under the same assay conditions. These findings suggest

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that P/CAF and p300, perhaps by forming a complex, act in concert to suppress cell cycle progression.

Histone acetyltransferase activity in P/CAF

Acetylation of the N-terminal histone tails has been considered to play a crucial role in accessibility of transcription factors to nucleosomal templates (26-27). Recently, yGCN5 has been identified as a histone acetyltransferase (28). On the basis of this information, intrinsic histone acetyltransferase activity in P/CAF and hGCN5 was examined. As substrates, the core histones (histones H2A, H2B, H3 and H4) and the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) were used.

Activity of hGCN5 and P/CAF that acetylates free histones or histones in the nucleosome core particle (35) was measured as described (36). Each reaction contained 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. The histone octamer dissociated into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE.

P/CAF and hGCN5 acetylated the core histones with almost the same efficiency. Both factors acetylated histones H3 and H4, but preferentially H3. In contrast, very weak or no acetylation by hGCN5 was detected in the nucleosome core particles. Remarkably, significant acetylation by P/CAF was observed in a nucleosomal context. Although all core histones are acetylated in the nucleus, P/CAF and hGCN5 did not acetylate histones H2A and H2B *in vitro*.

Direct function of P/CAF is likely to involve its intrinsic histone acetyltransferase activity. Although exact molecular mechanisms by which acetylation of core histones contribute to transcription remains undefined, acetylation of the histones is considered to play an important role in transcriptional regulation (26-27). The positively charged N-terminal tails of core histones are believed to affect nucleosome structure by interacting



with DNA at or near the nucleosome-spacer junction. Acetylation of the histone tails presumably destabilizes the nucleosome and facilitates access by regulatory factors. Likewise, there is a general correlation between the level of acetylation and transcriptional activity of nucleosomal domains. The findings of the present invention provide insights into the mechanisms of targeted histone acetylation.

Cellular factor p300/CBP binds to various sequence-specific factors that are involved in cell growth and/or differentiation, including CREB (3,4), c-Jun (9), Fos (11), c-Myb (12) and nuclear receptors (13). P/CAF could stimulate the activation function of these factors via promoter-specific histone acetylation. The present invention demonstrates that E1A appears to perturb normal cellular regulation by disrupting the connection between p300/CBP and its associated histone acetyltransferase.

II. p300/CBP studies.

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Purification of E1A associated histone acetyltransferase.

FLAG-epitope tagged E1A (or ΔE1A) was expressed in Sf9 cells (ATCC accession number CRL 1711) by infecting recombinant baculovirus (43). All purification steps were carried out at 4°C. Extract was prepared from infected cells by one cycle of freeze and thaw in buffer B (20 mM Tris-HCl, pH 8.0; 5 mM MgCl₂, 10% glycerol; 1 mM PMSF; 10 mMβ-mercaptoethanol, 0.1% Tween 20) containing 0.1 M KCl and the complete protease inhibitor cocktail (Boehringer Mannheim). To prepare E1A-immobilized beads, the extract was incubated with M2 anti-FLAG antibody agarose (Kodak-IBI) for four hours with rotating and subsequently washed with the same buffer three times. The resulting beads were incubated with HeLa (ATCC accession number CCL 2) nuclear extract for four to eight hours and thereafter washed with the same buffer six times. Finally, FLAG-E1A was eluted from the beads along with associated polypeptides by incubating with the same buffer containing 0.1 mg/ml FLAG peptide.

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For further purification, eluted polypeptides were dialyzed in 0.05 M KCI-buffer B and subsequently loaded onto a SMART Mono Q column (Pharmacia) equilibrated with the same 0.05 M KCI-buffer B. After washing, the column was developed with a linear gradient of 0.05-1.0 M KCl in buffer B. Mono Q fractions were concentrated with a MICROCON spin-filter (Amicon) and consequently loaded onto a SMART Superdex

200 column (Pharmacia) equilibrated with 0.1 M KCl-buffer B.

Histone acetyltransferase assays

Filter binding assays were performed as described (80) with minor modifications. Samples were incubated at 30°C for 10-60 minutes in 30 ml of assay buffer containing 50 mM Tris-HCl, pH 8.0, 10% glycerol, 1 mM DTT, 1 mM PMSF, 10 mM sodium butyrate; 6 pinol of [³H]acetyl CoA (4.3 mCi/mmole, Amersham Life Science Inc.); and 33 mg/ml of calf thymus histones (Sigma Chemical Co.). In experiments where synthetic peptides were substituted for core histones, 50 pmol of each peptide were used. After incubation, the reaction mixture was spotted onto Whatman P-81 phosphocellulose filter paper and washed for 30 minutes with 0.2 M sodium carbonate buffer pH 9.2 at room temperature with 2-3 changes of the buffer. The dried filters were counted in a liquid scintillation counter.

PAGE analysis was done as above except that 90 pmol of [14C] acetyl CoA (55 mCi/mmole, Amersham Life Science Inc.) and 9 pmol of core histones or mononucleosomes were used. Core histones and mononucleosomes were prepared as described (35). For trypsin digestion, reaction mixtures were further incubated with various amounts of trypsin on ice for 30 minutes. The samples were analyzed on one dimensional SDS-PAGE gels or two dimensional gels, where the first dimension was an acid-urea-PAGE gel (44) and the second dimension was an SDS-PAGE gel.

Protein expression

For baculovirus expression, cDNAs corresponding to p300 portions of aa 1-670, aa 671-1194 and aa 1135-2414 were amplified by PCR (EXPAND High Fidelity PCR System, Boehringer Mannheim) as KpnI-NotI fragments. The resulting fragments were

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subcloned into a baculovirus transfer vector having the FLAG-tag sequence (43). The recombinant viruses were isolated using the BACULOGOLD system (Pharmingen), according to the manufacturer's protocol and were infected into Sf9 cells (ATCC accession number CRL 1711) to express FLAG-p300. Recombinant proteins were affinity purified with M2 anti-FLAG antibody-immobilized agarose (Kodak-IBI) according to the manufacturer's protocol.

For bacterial expression, cDNAs encoding the p300 portions and the CBP portion (aa 1174-1850) were first subcloned into the baculovirus transfer vector having the FLAG-tag as described above. Thereafter, the XhoI and NotI fragments encoding FLAG-p300 or FLAG-CBP fusions were resubcloned into the E. coli expression vector pET-28c (Novagene) digested with SalI and NotI. Recombinant proteins were expressed in E. coli BL21(DE3) and affinity purified with M2-antibody agarose.

15 Histone acetyltransferases that associate with E1A

Although the adenovirus E1A 12S protein (E1A) inhibits transcription in a variety of genes via direct binding to p300/CBP (45), E1A also stimulates transcription in some contexts (46). Thus, p300/CBP-bound E1A was tested to determine whether it might recruit histone acetyltransferases or deacetylases to regulate transcription. In addition, experiments were conducted as described below to determine if p300/CBP per se is a histone acetyltransferase.

Initially, recombinant FLAG-epitope tagged E1A was immobilized on anti-FLAG antibody beads. Immobilized E1A was incubated with a HeLa nuclear extract for affinity purification of E1A-associated polypeptides. FLAG-E1A was then eluted from the beads, along with E1A-associated polypeptides, by incubating with FLAG-peptide. Although E1A per se has no histone acetyltransferase activity, E1A recruited significant amounts of histone acetyltransferase activity from the nuclear extract. It is very unlikely that this activity is derived from P/CAF given that E1A and P/CAF cannot bind to p300/CBP simultaneously (43). Consistent with this, no P/CAF was detected in these fractions by immunoblotting.

The E1A N-terminus, a region that is not highly conserved among the various adenovirus serotypes, is involved in p300/CBP binding in vivo. Mutations in the N-terminal region lead to loss of the ability for p300/CBP binding without affecting RB binding (1,47). Thus, the requirement of the E1A N-terminal region for the recruitment of histone acetyltransferase activity was tested. In contrast to the wild type, the N-terminal deleted form of E1A (Δ N-E1A) recruited only a background level of acetyltransferase activity. In agreement with previous reports (47), the $\Delta N-E1A$ showed no ability to interact with p300/CBP, although it still retained the ability to 10 interact with a variety of other polypeptides, including RB.

To define the relationship between p300/CBP and histone acetyltransferase activity, affinity purified E1A-binding polypeptides were separated by Mono Q ion-exchange column. Both p300/CBP and the acetyltransferase activity were coeluted at 140 mM KCl, while most of polypeptides were eluted at 260 mM KCl. The active fraction of Mono Q column (~140 mM KCl) was further separated by Superdex-200 gel filtration column. Both p300/CBP and the acetyltransferase activity coeluted after the void volume, indicating that p300/CBP is involved in the histone acetyltransferase activity.

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p300 is a histone acetyltransferase

The data provided herein indicate that p300 per se, or a polypeptide(s) associated with p300, possesses histone acetyltransferase activity. To test the former possibility, the acetyltransferase activity of recombinant p300 was measured. p300 was divided into three fragments, each of which was expressed in and purified from Sf9 cells via a baculovirus expression vector. Histone acetyltransferase activity was readily detected in the C-terminal fragment containing amino acids 1135-2414, whereas no activity was found in the other fragments, demonstrating conclusively that p300 per se is a histone acetyltransferase:

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p300/CBP-histone acetyltransferase domain

To map the histone acetyltransferase domain of p300, a series of deletions was prepared. Given the poor conservation of the glutamine-rich region (aa 1815-2414) in the C. elegans p300/CBP homolog (6), the p300 fragment encoding aa 1135-1810 was expressed in and purified from E. coli. Importantly, this candidate region of p300 (aa 1135-1810) showed significant histone acetyltransferase activity. For further mapping within this region, a series of N-terminal deletions was constructed. Deletion of 60 residues, resulting in a fragment containing as 1195-1810, had no effect on the acetyltransferase activity, whereas the deletion of 185 residues, yielding a fragment comprising as residues 1320-1810, completely eliminated the acetyltransferase activity.

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Next, a series of C-terminal deletions was analyzed to determine the requirement of the P/CAF (or E1A) -binding domain. The p300 fragments lacking the E1A binding domain (aa 1195-1760, 1195-1706 and 1195-1673) still retained the acetyltransferase activity, whereas the further truncated mutant (aa 1195-1652) completely lost the acetyltransferase activity. Consistent with these results, the internal deletion of residues 1418-1720 showed no acetyltransferase activity. These data demonstrate that the histone acetyltransferase domain is located between the bromodomain and the E1A-binding domain. Given that the histone acetyltransferase domain is highly conserved between p300 and CBP (91% similarity), the corresponding region of CBP, aa residues 1174-1850, was expressed to confirm the acetyltransferase activity. As expected, comparable activity was detected, indicating that both p300 and CBP are histone acetyltransferases.

Among various acetyltransferases including histone acetyltransferases GCN5 and P/CAF, putative acetyl-CoA binding sites are conserved (48). However, multiple alignment analysis (49) showed that the p300/CBP histone acetyltransferase domain does not belong to this group. Moreover, comparison of the p300/CBP histone acetyltransferase domain with peptide sequence databases (23) showed no sequence similarity to any other proteins. Accordingly, this invention shows that p300/CBP represents a novel class of acetyltransferases in that it does not have the conserved motif found among previously described acetyltransferases (48).



p300 acetylates all core histones in mononucleosomes

Substrate specificity for acetylation by p300 was also examined. As substrates, histone octamers and mononucleosomes (146 base pairs of DNA wrapped around the octamer of core histones) were used. Given that the histone octamer dissociates into dimers or tetramers under physiological conditions, the histone octamer is referred to here as core histones. When core histones were used, p300 acetylated all four proteins, but preferentially H3 and H4. More importantly, in a nucleosomal context, p300 acetylated all four core histones nearly stoichiometrically. In contrast, p300 acetylated neither BSA nor lysozyme.

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Hyperacetylated histones are believed to be linked with transcriptionally active chromatin (26,27,50,51). Hyperacetylated forms are found in histones H4, H3 and H2B, which have multiple acetylation sites in vivo. Thus, the level of acetylation by p300 was also tested.

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Mononucleosomes treated with p300 were analyzed by two-dimensional gel electrophoresis. A Coomassie blue-stained gel and the corresponding autoradiogram showed that a significant amount of histones, especially H4, were hyperacetylated. Importantly, acetylation levels by p300 were very close to those of hyperacetylated 20 histones prepared from HeLa nuclei treated with sodium butyrate, a histone deacetylase inhibitor. In contrast, no acetylated forms were detected in the reaction without p300. These results indicate that p300 acetylates histones in mononucleosomes to the hyperacetylated state by targeting multiple lysine residues.

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p300 acetylates the four lysines in the histone H4 N-terminal tail in vitro which are acetylated in vivo

Lysines at positions 5, 8, 12 and 16 of histone H4 are acetylated in vivo (51). Recent studies with yeast histone acetyltransferases demonstrate the position-specific acetylation by distinct acetyltransferases, i.e., while cytoplasmic 30 acetyltransferases for histone deposition and chromatin assembly modify

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positions 5 and 12, GCN5 modifies positions 8 and 16 (52). Accordingly, the positions of acetylation by p300 were also determined. A series of synthetic peptides containing acetylated lysines at various positions was used to determine the acetylation site-specificity of p300. Consistent with the two-dimensional gel electrophoresis analysis, the experiments with peptide substrates showed that p300 acetylates all four lysines in the histone H4 that are acetylated *in vivo*. These results are consistent with the view that deposition-related diacetylated histones are deacetylated during maturation of chromatin (53).

p300 preferentially acetylates the N-terminal histone tail

Histone acetyltransferases modify specific lysine residues in the N-terminal tail of core histones but not the C-terminal globular domain in vivo (26,27,50,51). Structural models of nucleosomes (54,55,56) suggest that most of the lysine residues in the C-terminal globular domain are buried. Therefore, experiments were conducted to examine whether restricted acetylation of the N-terminal tail resulted from the substrate 15 specificity of the enzyme or inaccessibility of the enzyme to the core domain in nucleosomes. The globular domains of all core histones contain a long helix flanked on either side by a loop segment and short helix, termed the "histone fold" (54,55,56). The histone fold is involved in formation of the stable H2A-H2B and H3-H4 hetero-dimers, consisting of extensive hydrophobic contacts between the paired 20 molecules. Therefore, it is likely that a histone monomer cannot fold properly, thereby increasing access of the histone acetyltransferase to the core domain. Based on these considerations, experiments were conducted to determine whether p300 acetylates free histone H4 in a N-terminal-specific manner.

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Histone H4 was acetylated with p300 and subsequently the histone tail was removed by partial digestion with trypsin. The distributions of radioactivity between intact and core histones were compared. While the globular core histone domain was predominant at the higher trypsin concentrations, radioactivity was detected mostly in the intact histone. These data demonstrate that p300 preferentially acetylates the N-terminal tail of histone H4.

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III. P/CAF interaction with MyoD

Tissue culture and transfection experiments

 C_2C_{12} mouse cells (ATCC accession number CRL 1772) were grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 20% fetal bovine serum (FBS) until they reached confluence. Differentiation was induced by switching medium to differentiation medium (DM), consisting of DMEM containing 2% horse serum. $C_3H/10T1/2$ fibroblasts (ATCC accession number CCL 226) were grown in DMEM supplemented with 10% FBS. Cells were transfected by the calcium phosphate precipitation method. Total amounts of transfected DNA were equalized by empty vector DNA. After 12 h incubation in medium containing the precipitated DNA, the cells were washed and incubated in fresh DMEM containing 10% FBS for an additional 24 h. Afterwards, differentiation was induced by incubating in DM for 36 to 72 h. Chloramphenicol acetyltransferase (CAT) assays were performed as previously described (64,69). The quantities of cell extracts used for CAT assays were normalized to β -galactosidase activity by cotransfection of 1 mg of the β -galactosidase expression vector, pON260.

Expression vectors used for transfection experiments are as follows:

pCX-P/CAF for P/CAF (43), pCMV-bp300 for p300 (65), pCMV-p300 (1869-2414)

25 (64) and pCMV-p300 (1514-1922) (60) for p300 wild type and mutants, pE1A12S, pE1A12S R2G, pE1A12S D2-36 and pE1A12S D121-130 for E1A wild type and mutants (66,67,68); and pEMSV-MyoD for MyoD (64).

The antisense P/CAF RNA expression vector, pcDNA3 P/CAF-AS, was created as follows. The 2.5 Kb EcoRI-KpnI fragment containing the entire P/CAF open reading frame was isolated from pCX-P/CAF (43). This fragment was subcloned into the

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EcoRI-KpnI sites of plasmid pcDNA3 (Invitrogen) so that the antisense P/CAF RNA is driven under the CMV promoter. Reporter genes employed were 4RE-CAT and MCK-CAT (69). 4RE-CAT is driven by a synthetic promoter containing 4 copies of the E-box, whereas MCK-CAT is driven by the native MCK promoter (nucleotides -1256 to +7).

Microinjection and immunofluorescence

Cells were grown on small glass slides, subdivided into numbered squares of 2 mm x 2 mm and microinjected with purified and concentrated antibodies, as previously described (70). For immunofluorescence, cells were fixed in either 2% paraformaldehyde or 1:2 methanol/acetone solution, preincubated with 5% BSA/PBS and incubated with the primary antibodies for 30 min at 37° C. Subsequently, antibody was visualized by incubating with either rhodamine- or fluorescein-conjugated secondary antibody for 30 min at 37° C. Injected antibodies were stained with a rhodamine-conjugated secondary antibody and nuclei were counter-stained by DAPI as previously described (69).

Antibodies employed are as follows, rabbit polyclonal affinity purified affi-P/CAF antibody (43), rabbit polyclonal anti-p300/CBP antiserum (71), mouse monoclonal anti-MyoD antibody (clone 5.8A, kindly provided by P. Houghton), goat polyclonal anti-c-Jun affinity purified antibody (Santa Cruz) and rabbit pre-immune serum.

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Immunoprecipitation and DNA affinity purification

Cells were resuspended in lysis buffer (20 mM NaPO₄, 150 mM NaCl, 5mM MgCl₂, 0.1% NP40, 1 mM DTT, 10 mM sodium fluoride, 0.1 mM sodium vanadate, 1 mM phenylmethylsulfonyl-fluoride and 10 mg/ml each of leupeptin, aprotinin and pepstatin). After 30 min incubation on ice, samples were centrifuged at 12,000 x g for 30 min and supernatants were used as cell extracts. Extracts were pre-cleared by

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incubating with rabbit pre-immune serum and protein A/G Plus-Agarose (Santa Cruz) for 2 h at 4 C. For immunoprecipitation, the supernatants were incubated with the respective antibodies for 3 h at 4 C. Protein A/G Plus-Agarose was added, and incubation continued for 3 h. The matrix was washed with lysis buffer, then boiled in 2 X SDS sample buffer. Immunoblotting was performed by using the ECL chemiluminescent detection kit (Amersham) according to the manufacturer's protocol.

Affinity purification of E-box-bound complexes was done as previously described (69). Briefly, 100 ng of the biotinylated double stranded DNA containing the E-box were immobilized on streptavidin-conjugated magnetic beads and incubated with 500 mg of cell extracts in the presence of poly dI-dC. After extensive washing, bound proteins were eluted with SDS sample buffer and analyzed by immunoblotting.

In vitro protein-protein interaction assays

The CBP-B fragment and its deletion derivatives were expressed as 15 GST-fusions described previously (43). MyoD and E1A (43) were expressed as FLAG-fusion proteins in Sf9 cells via a baculovirus expression system and affinity-purified on M2 anti-FLAG antibody-agarose (Kodak-IBI). Crude E. coli extracts containing GST-fusions were incubated with various amounts of MyoD and/or E1A in 50 ml of buffer B (20 mM Tris-HCl, pH 8.0, 0.1 M KCl, 5 mM MgCl₂, 10% 20 glycerol, and 0:1% Nonidet P-40) on ice for 10 min. GST-precipitation was performed as described (43). MyoD and E1A were detected by immunoblotting with anti-FLAG M2 antibody. For the interaction between P/CAF and MyoD, 1.5 pmol of FLAG-P/CAF and 15 pmol of FLAG-MyoD were incubated in 50 ml of buffer B on ice for 10 min. The mixture was further incubated with 2 mg of anti-P/CAF (43) or 25 anti-hADA2 antibody for 60 min. The immunocomplexes were precipitated by incubation with the rul of protein A-Trisacryl (Pierce) and rotated for 1-4 hr at 4oC. The matrix was washed 4 times with 200 ml of buffer B and boiled in 10 ml of 2 X SDS sample buffer. The proteins were resolved on a 4%-20% gradient SDS-PAGE and subjected to immunoblotting with the anti-FLAG M2 antibody. The blot was developed 30 with the SUPERSIGNAL chemiluminescent substrates (Pierce).

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P/CAF coactivates muscle-specific transcription

P/CAF and MyoD were co-transfected into mouse C3H10T1/2 fibroblasts, and MyoD-mediated transcription was determined from reporter activity driven by the artificial (4RE) and the naturally-occurring muscle creatine kinase (MCK) promoters. Overexpression of P/CAF stimulated MyoD-dependent transcription several folds in both promoters. Similar results were obtained for the myoD activated myogenin promoter Transcriptional activation was further stimulated by co-transfecting with MyoD, P/CAF and p300 expression vectors, suggesting that P/CAF may function by forming a complex with p300/CBP. Consistent with the lack of DNA binding capacity in P/CAF, overexpression of P/CAF alone did not increase the basal transcriptional activity of either enhancer. To test whether P/CAF and p300/CBP function in the same pathway, two dominant negative forms of p300 were employed which specifically inhibit p300/CBP-mediated transcription (60,64). The p300 segment spanning residues 1514-1922 inhibits the MyoD-dependent activation via direct interaction with MyoD 15 (60), whereas the p300 segment spanning residues 1869-2414 inhibit it without direct interaction (64). Both dominant negative mutants inhibited MyoD-coactivation by P/CAF), suggesting that P/CAF and p300/CBP function in the same pathway.

20 -For further elucidation of the activation mechanism by P/CAF, the effect of E1A, which inhibits MyoD-dependent transcription and differentiation (66,72,73) via direct interaction with p300/CBP (65,78), was tested. Expression of E1A in C3H10T1/2 fibroblasts inhibited stimulation of MyoD-directed transcription by P/CAF overexpression. E1A mutants lacking p300/CBP-binding activity, E1A D2-36 and E1A R2G (67,79), had almost no effect. On the other hand, an E1A mutant retaining p300/CBP-binding activity, E1A D121-130, behaved like the wild type. Since E1A associates with p300/CBP, but not with P/CAF, these results suggest that P/CAF functions in MyoD-directed transcription via interaction with p300/CBP.

To address the role of P/CAF as a myogenic coactivator in a more relevant environment, P/CAF was overexpressed in proliferating C2C12 myoblasts which express





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endogenous myogenic bHLH factors. As observed in fibroblasts, overexpression of P/CAF stimulated muscle specific transcription. Concomitant expression of exogenous p300 increased P/CAF-mediated coactivation. The repression exerted by wild type E1A, but not mutant E1A D2-36, on P/CAF coactivation of MyoD was also observed in muscle cells.

Similar experiments were performed with myogenic cell lines that were stably transformed with wild type or mutant E1A-expressing vectors (66). Coactivation by P/CAF was inhibited by wild type E1A or the E1A mutant that retains

10 ap300/CBP-binding activity (E1AΔ121-130). In contrast, E1A mutants that lack p300/CBP-binding (E1A Δ2-36 and E1A R2G) allowed transcriptional coactivation by P/CAF. Taken together, these experiments show that P/CAF coactivates MyoD-directed transcription via interaction with p300/CBP.

15 P/CAF stimulates myogenic differentiation

Given that P/CAF potentiates MyoD-directed transcription, the ability of P/CAF to assist MyoD in promoting myogenic differentiation was investigated. To this aim, C3H10T1/2 fibroblasts were transiently transfected with P/CAF and MyoD expression vectors. An expression vector for the green fluorescent protein (GFP) was

20 co-transfected to identify transfected cells. After incubation in differentiation medium, the myogenic conversion of transfected cells was determined by simultaneous expression of the GFP and the differentiation-specific marker myosin heavy chain (MHC). Forced expression of MyoD in fibroblasts caused muscle differentiation in 12% of the transfected fibroblasts. This myogenic conversion was 20% by co-expressing MyoD and P/CAF. As observed in transcription experiments, stimulation of differentiation by P/CAF was counteracted by co-transfection with the p300 dominant negative mutant, p300 (1869-2414). Consistent with a general role for coactivators, overexpression of P/CAF alone was unable to differentiate fibroblasts.

Similar experiments were done using proliferating C2C12 myoblasts in which the differentiation program is already committed. Most of the myoblasts differentiated into

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myotubes by overexpressing P/CAF, whereas only a modest effect was observed by overexpressing p300. In contrast, differentiation was inhibited slightly by overexpressing c-Jun. This inhibitory effect presumably was caused by titration of p300/CBP, which associates directly with c-Jun (74). A similar inhibition was observed in the p300 dominant negative mutant. Consistent with the transcriptional effect, E1A almost completely inhibited differentiation. The E1A mutant RG2, lacking p300/CBP-binding capability but retaining the retinoblastoma protein (Rb)-binding capability, only partially inhibited differentiation, although this same mutant inhibited transcription as severely as the wild type. Taken together, these data show that P/CAF stimulates muscle differentiation by coactivating MyoD function via p300/CBP association.

P/CAF is essential for myogenic transcription and differentiation

To test the necessity of P/CAF for myogenic transcription, experiments were conducted whereby P/CAF synthesis was inhibited by expressing antisense P/CAF RNA. A vector from which the P/CAF mRNA is transcribed in the antisense orientation (P/CAF-AS) was transfected with P/CAF and MyoD expression vectors into fibroblasts and MyoD-dependent transcription was examined. Cotransfection of the antisense expression vector strongly inhibited MyoD-dependent transcription below the level of induction elucidated by MyoD alone, demonstrating that expression of P/CAF antisense RNA inhibits not only the coactivation exerted by exogenous P/CAF but also that of endogenous P/CAF. These results indicate that P/CAF is essential for MyoD-dependent transcription.

Studies were also carried out to determine whether expression of P/CAF antisense RNA inhibits myogenic differentiation. C3H10T1/2 fibroblasts were transiently transfected with various expression vectors with or without the P/CAF antisense RNA expression vector. Expression of P/CAF antisense RNA reduced MyoD-mediated myogenic conversion of fibroblasts. Expression of P/CAF antisense RNA also counteracted the stimulatory effect of both P/CAF and p300 on myogenic differentiation. These data support the view that P/CAF and p300/CBP coactivate



MyoD-dependent transcription in the same pathway. More drastic inhibition was observed in C2C12 myoblasts in similar experiments. Therefore, it can be concluded that P/CAF is essential for transcription of muscle specific genes and hence differentiation into myotubes.

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To further confirm the essential role of P/CAF for myogenic differentiation, we blockage experiments by antibody microinjection were performed. Antibodies were injected into the cytoplasm of proliferating C2C12 myoblasts to prevent the nuclear transport of newly synthesized target proteins. After incubating in the differentiation medium, the degree of differentiation was determined. Microinjection of an anti-P/CAF antibody almost completely inhibited differentiation. Similar results were obtained by microinjecting anti-p300/CBP antibodies. Although microinjection of either anti-p300/CBP or P/CAF antibody was sufficient to inhibit differentiation, an even greater inhibition was observed by coinjecting both of them. Microinjection of anti-P/CAF or anti-p300/CBP antibody did not interfere with induction of p53 by DNA damaging agents, showing specificity of the inhibition by the antibodies. In contrast to anti-P/CAF or anti-p300/CBP antibodies, the injection of anti-MyoD antibody only partially inhibited differentiation, supporting the view of functional redundancy between MyoD and Myf-5 (75,76). Injection of anti-c-Jun antibody or control antibody did not interfere with muscle differentiation.

Similar experiments were performed with C3H10T1/2 fibroblasts stably expressing MyoD. In these cells, either anti-p300/CBP or anti-P/CAF antibody completely inhibited muscle differentiation. In contrast to myoblasts, anti-MyoD antibody completely blocked differentiation in the fibroblasts expressing MyoD. Anti-c-Jun and control antibodies did not interfere with differentiation. Taken together, these results demonstrate that P/CAF and p300/CBP are indispensable for activation of the myogenic program.



p300/CBP, P/CAF and MyoD form a multimeric complex in vivo

The data described above indicate that P/CAF stimulates MyoD-directed transcription via association with p300/CBP. Thus, experiments were conducted to investigate whether P/CAF, p300/CBP and MyoD could associate in a complex.

First, cellular extracts derived from C2C12 myotubes were subjected to immunoprecipitation. Both anti-MyoD and anti-p300/CBP antibodies co-precipitated P/CAF. In a complementary experiment, both anti-p300/CBP and anti-P/CAF antibodies also co-precipitated MyoD, suggesting that these factors form a multimeric protein complex in myotubes.

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Next, attempts were made to detect this complex on the E-box, the DNA binding site for MyoD. Immobilized DNA containing an E-box sequence was incubated with myotube extracts. After extensive washing, P/CAF, p300/CBP and MyoD were analyzed by immunoblotting. P/CAF, p300/CBP and MyoD were all affinity purified on the immobilized DNA, whereas they were not purified on the control DNA lacking the E-box. Given that P/CAF and p300/CBP per se cannot bind to DNA, these observations indicate that P/CAF and p300/CBP are recruited through MyoD at the E-box sites to form a multi-protein complex.

20 Complex formation is inhibited by viral transforming factors

Since the oncoviral proteins E1A and large T antigen inhibit myogenic transcription and differentiation, the effect of these factors on the formation of complexes on the E-box was tested. Importantly, very small amouts of P/CAF and p300/CBP were co-purified on the E-box from myocyte extracts which stably express E1A or large T antigen, although MyoD was detected under these conditions. The lower recovery of MyoD from E1A-expressing muscle cells could reflect the low level of MyoD in the extracts (66). These results indicate that E1A and large T antigen dissociate P/CAF and p300/CBP from MyoD without altering MyoD binding to DNA.

Consistent with the previous observations that transiently expressed E1A prevents interaction between P/CAF and p300/CBP in vivo (43), the association

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between p300/CBP and P/CAF was abolished in myoblasts stably transformed by wild type E1A but not in those clones transformed with the E1A mutant R2G unable to bind p300/CBP. Similarly, the interaction between p300/CBP and P/CAF was abolished by large T antigen but not by the mutant protein that localizes into the cytoplasm (77).

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Interaction between MyoD, P/CAF and CBP in vitro

Previous interaction experiments in vitro indicate that the CBP region spanning residues 1801 to 1850 is crucial for interaction with both P/CAF and E1A (43). While most sequence-specific factors bind to CBP sites distinct from the P/CAF/E1A binding sites, MyoD interacts with an overlapping CBP fragment called the CH3 region (60.64.65). To understand how P/CAF, p300/CBP and MyoD associate, the CBP sites important for MyoD binding were mapped more precisely. Consistent with previous reports (60,64,65), the CBP fragment spanning residues 1801-2000 (fragment B) bound MyoD. Moreover, deletion of residues 1801 to 1850 within fragment B completely abolished interaction with MyoD, which is similar to the results obtained with P/CAF and E1A. Importantly, an internal deletion of residues 1850-1878 abolished the MyoD interaction with CBP, while it did not affect binding of E1A or P/CAF (43). These results suggest that MyoD and P/CAF bind to distinct sites of p300/CBP, albeit the binding sites may overlap. Moreover, a direct interaction was observed between MyoD and P/CAF, which may contribute to stabilization of the multimeric complex.

These data show that E1A prevents not only p300/CBP-interaction with P/CAF but also that with MyoD in vivo. To obtain evidence that this inhibition is due to the direct action by E1A, competition experiments were performed in vitro. Importantly, the interaction between CBP and MyoD was strongly inhibited by addition of E1A, implicating that E1A inhibits myogenic transcription by disrupting multiple interactions.

Although the present process has been described with reference to specific details of certain embodiments thereof, it is not intended that such details should be



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The invention except as and to the extent that

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regarded as limitations upon the scope of the invention except as and to the extent that they are included in the accompanying claims.

Throughout this application various publications are referenced by numbers

within parentheses. Full citations for these publications are as follows. The disclosures
of these publications in their entireties are hereby incorporated by reference into this
application in order to more fully describe the state of the art to which this invention
pertains.

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SEQUENCE LISTING

- (1) GENERAL INFORMATION
- (i) APPLICANT: The United States of America, as repesented by the Secretary, Department of Health and Human Services, c/o National Institutes of Health, Office of Technology Transfer, 6011 Executive Boulevard, Suite 325, Rockville, Maryland 20842
- (ii) TITLE OF THE INVENTION: METHODS AND COMPOSITIONS FOR p300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF
- (iii) NUMBER OF SEQUENCES: 18
- (iv) CORRESPONDENCE ADDRESS:
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 - (B) STREET: Suite 1200, 127 Peachtree Street, NE
 - (C) CITY: Atlanta
 - (D) STATE: GA
 - (E) COUNTRY: USA
 - (F) ZIP: 30303
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: DOS
 - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE: 23-JUL-1997
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: Corresponding U.S. Serial No. 60/022,273
 - (B) FILING DATE: 23-July-1996
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Miller, Mary L
 - (B) REGISTRATION NUMBER: 39,303
 - (C) REFERENCE/DOCKET NUMBER: 14014.0238/P
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 404/688-0770
 - (B) TELEFAX: 404/688-9880
 - (C) TELEX:
 - (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 832 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None





(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Met Ser Glu Ala Gly Gly Ala Gly Pro Gly Gly Cys Gly Ala Gly Ala Gly Ala Gly Ala Gly Pro Gly Ala Leu Pro Pro Gln Pro Ala Ala Leu 20 2.5 Pro Pro Ala Pro Pro Gln Gly Ser Pro Cys Ala Ala Ala Ala Gly Gly 40 Ser Gly Ala Cys Gly Pro Ala Thr Ala Val Ala Ala Ala Gly Thr Ala 55. Glu Gly Pro Gly Gly Gly Ser Ala Arg Ile Ala Val Lys Lys Ala 70 Gln Leu Arg Ser Ala Pro Arg Ala Lys Lys Leu Glu Lys Leu Gly Val 90 Tyr Ser Ala Cys Lys Ala Glu Glu Ser Cys Lys Cys Asn Gly Trp Lys 105 100 Asn Pro Asn Pro Ser Pro Thr Pro Pro Arg Ala Asp Leu Gln Gln Ile 125 120 Ile Val Ser Leu Thr Glu Ser Cys Arg Ser Cys Ser His Ala Leu Ala 140 135 Ala His Val Ser His Leu Glu Asn Val Ser Glu Glu Glu Met Asn Arg 150 155 Leu Leu Gly Ile Val Leu Asp Val Glu Tyr Leu Phe Thr Cys Val His 170 165 Lvs Glu Glu Asp Ala Asp Thr Lys Gln Val Tyr Phe Tyr Leu Phe Lys 185 180 Leu Leu Arg Lys Ser Ile Leu Gln Arg Gly Lys Pro Val Val Glu Gly 200 205 195 Ser Leu Glu Lys Lys Pro Pro Phe Glu Lys Pro Ser Ile Glu Gln Gly 220 215 Val Asn Asn Phe Val Gln Tyr Lys Phe Ser His Leu Pro Ala Lys Glu 230 235 Arg Gln Thr Ile Val Glu Leu Ala Lys Met Phe Leu Asn Arg Ile Asn 250 Tyr Trp His Leu Glu Ala Pro Ser Gln Arg Arg Leu Arg Ser Pro Asn 265 260 Asp Asp Ile Ser Gly Tyr Lys Glu Asn Tyr Thr Arg Trp Leu Cys Tyr 285 280 Cys Asn Val Pro Gln Phe Cys Asp Ser Leu Pro Arg Tyr Glu Thr Thr 300 295 Gln Val Phe Gly Arg Thr Leu Leu Arg Ser Val Phe Thr Val Met Arg 315 310 Arg Gln Leu Leu Glu Gln Ala Arg Gln Glu Lys Asp Lys Leu Pro Leu 330 325 Glu Lys Arg Thr Leu Ile Leu Thr His Phe Pro Lys Phe Leu Ser Met 340 345 Leu Glu Glu Glu Val Tyr Ser Gln Asn Ser Pro Ile Trp Asp Gln Asp 365 360 Phe Leu Ser Ala Ser Ser Arg Thr Ser Gln Leu Gly Ile Gln Thr Val 375 Ile Asn Pro Pro Pro Val Ala Gly Thr Ile Ser Tyr Asn Ser Thr Ser 390 395 Ser Ser Leu Glu Gln Pro Asn Ala Gly Ser Ser Ser Pro Ala Cys Lys 410 405 Ala Ser Ser Gly Leu Glu Ala Asn Pro Gly Glu Lys Arg Lys Met Thr 420 425 Asp Ser His Val Leu Glu Glu Ala Lys Lys Pro Arg Val Met Gly Asp 445 440 Ile Pro Met Glu Leu Ile Asn Glu Val Met Ser Thr Ile Thr Asp Pro 460 455 Ala Ala Met Leu Gly Pro Glu Thr Asn Phe Leu Ser Ala His Ser Ala 470 475



Arg	Asp	Glu	Ala	Ala 485	Arg	Leu	Glu	Glu	Arg 490	Arg	Gly	Val	Ile	Glu 495	Phe
			500					505					510	Ile	
	-	515					520					525		Pro	
	530			•		535					540			His	
545			-		550					555				Cys	560
				565					5 70					Ala 575	
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	_	595					600					605		Thr	
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	_			645					650					Pro 655	
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		675					680					685		Gly	
	690					695					700			Ile	
705					710					715				Ser	720
				725					730		•			11e 735	
			740					745					750	Pro	
-		755					760					765		Pro	
•	770	_				775					780			Val	
785					790					795				Cys	800
	_			805					810					11e 815	
Glu	Lys	Phe	Phe 820		ser	гуѕ	TTE	Lуs 825	GΙU	Ата	σтУ	ьец	830	Asp	пуз

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 481 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Leu Glu Glu Val Tyr Ser Gln Asn Ser Pro Ile Trp Asp Gln 1 5 10 10 15 15 Asp Phe Leu Ser Ala Ser Ser Arg Thr Ser Gln Leu Gly Ile Gln Thr 20 25 30 30 Val Ile Asn Pro Pro Pro Val Ala Gly Thr Ile Ser Tyr Asn Ser Thr 35



	50					Pro 55					60	•			
65					70	Glu				75					80
	_			85		Glu			90					95	•
-			100			Ile		105	-				110		
		115			_	Pro	120					125			
	130	_				Arg 135					140				
145					150	Ser				155					160
				165		Leu			170					175	
-			180			Ile		185					190		
		195				Lys	200					205			
	210		•			Gln 215					220				
225					230	Val				235					240
		-		245		Ile			250					255	
-			260			Ile		265					270		
_		275				Lys	280					285			
_	290					Met 295					300				
305					310	Ile				315					320
-				325		Gln			330					335	
		_	340			Gly		345					350		
	_	355				Gly	360					365			
_	370					Asp 375					380				
385					390					395	•				400
				405					410					415	Pro
			420					425					430		Val
		435				Ala	440					445			
-	450	_				455					460				Ile
Leu 465 Lys	Glu	Lys	Phe	Phe	Phe 470	Ser	Lys	Ile	Lys	Glu 475	Ala	GLŸ	Leu	ile	Asp 480

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 203 amino acids

 - (B) TYPE: amino acid (C) STRANDEDNESS: single

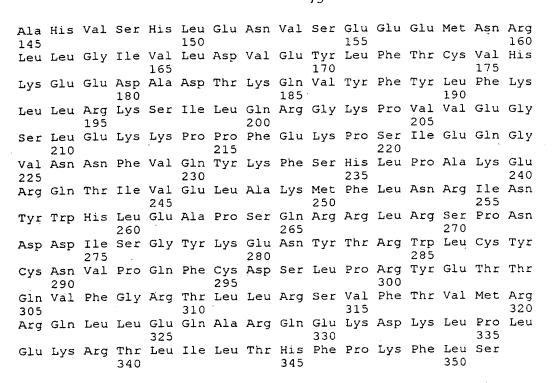
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Arg Val Val Gln His Thr Lys Gly Cys Lys Arg Lys Thr Asn Gly Gly 10 Cys Pro Ile Cys Lys Gln Leu Ile Ala Leu Cys Cys Tyr His Ala Lys 20 25 His Cys Gln Glu Asn Lys Cys Pro Val Pro Phe Cys Leu Asn Ile Lys 40 Gln Lys Leu Arg Gln Gln Gln Leu Gln His Arg Leu Gln Gln Ala Gln 5.5 Met Leu Arg Arg Arg Met Ala Ser Met Arg Thr Gly Val Val Gly Gln 75 Gln Gln Gly Leu Pro Ser Pro Thr Pro Ala Thr Pro Thr Thr Pro Thr 90 85 Gly Gln Gln Pro Thr Thr Pro Gln Thr Pro Gln Pro Thr Ser Gln Pro 105 100 Gln Pro Thr Pro Pro Asn Ser Met Pro Pro Tyr Leu Pro Arg Thr Gln 120 125 Ala Ala Gly Pro Val Ser Gln Gly Lys Ala Ala Gly Gln Val Thr Pro 140 135 Pro Thr Pro Pro Gln Thr Ala Gln Pro Pro Leu Pro Gly Pro Pro 155 150 Thr Ala Val Glu Met Ala Met Gln Ile Gln Arg Ala Ala Glu Thr Gln 170 165 Arg Gln Met Ala His Val Gln Ile Phe Gln Arg Pro Ile Gln His Gln 185 180 Met Pro Pro Met Thr Pro Met Ala Pro Met Gly 200

- (2) INFORMATION FOR SEQ ID NO:4:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 351 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ser Glu Ala Gly Gly Ala Gly Pro Gly Gly Cys Gly Ala Gly Ala 10 Gly Ala Gly Ala Gly Pro Gly Ala Leu Pro Pro Gln Pro Ala Ala Leu 20 25 Pro Pro Ala Pro Pro Gln Gly Ser Pro Cys Ala Ala Ala Gly Gly Ser Gly Ala Cys Gly Pro Ala Thr Ala Val Ala Ala Ala Gly Thr Ala ~55 Glu Gly Pro Gly Gly Gly Ser Ala Arg Ile Ala Val Lys Lys Ala 75 Gln Leu Arg Ser Ala Pro Arg Ala Lys Lys Leu Glu Lys Leu Gly Val 90 Tyr Ser Ala Cys Lys Ala Glu Glu Ser Cys Lys Cys Asn Gly Trp Lys 105 100 Asn Pro Asn Pro Ser Pro Thr Pro Pro Arg Ala Asp Leu Gln Gln Ile 120 125 Ile Val Ser Leu Thr Glu Ser Cys Arg Ser Cys Ser His Ala Leu Ala 135





(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 476 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Leu Glu Glu Glu Ile Tyr Gly Ala Asn Ser Pro Ile Trp Glu Ser 10 Gly Phe Thr Met Pro Pro Ser Glu Gly Thr Gln Leu Val Pro Arg Pro 25 Ala Ser Val Ser Ala Ala Val Val Pro Ser Thr Pro Ile Phe Ser Pro 45 40 Ser Met Gly Gly Ser Asn Ser Ser Leu Ser Leu Asp Ser Ala Gly 55 Ala Glu Pro Met Pro Gly Glu Lys Arg Thr Leu Pro Glu Asn Leu Thr 75 . 80 70 Leu Glu Asp Ala Lys Arg Leu Arg Val Met Gly Asp Ile Pro Met Glu 85 90 Leu Val Asn Glu Val Met Leu Thr Ile Thr Asp Pro Ala Ala Met Leu 105 110 Gly Pro Glu Thr Ser Leu Leu Ser Ala Asn Ala Ala Arg Asp Glu Thr 120 125 Ala Arg Leu Glu Glu Arg Arg Gly Ile Ile Glu Phe His Val Ile Gly 135 140 Asn Ser Leu Thr Pro Lys Ala Asn Arg Arg Val Leu Leu Trp Leu Val 150 155 Gly Leu Gln Asn Val Phe Ser His Gln Leu Pro Arg Met Pro Lys Glu 165 170 175 Tyr Ile Ala Arg Leu Val Phe Asp Pro Lys His Lys Thr Leu Ala Leu 185

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Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe Pro 200 195 Thr Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val Thr Ser Asn Glu 215 220 Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His Leu Lys Glu Tyr 230 235 His Ile Lys His Asn Ile Leu Tyr Phe Leu Thr Tyr Ala Asp Glu Tyr 250 245 Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Asp Ile Lys Val 265 270 Pro Lys Ser Arg Tyr Leu Gly Tyr Ile Lys Asp Tyr Glu Gly Ala Thr 285 280 275 Leu Met Glu Cys Glu Leu Asn Pro Arg Ile Pro Tyr Thr Glu Leu Ser 300 295 His Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu Arg 310 315 Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe Lys 325 330 Glu Gly Val Arg Gln Ile Pro Val Glu Ser Val Pro Gly Ile Arg Glu 345 Thr Gly Trp Lys Pro Leu Gly Lys Glu Lys Gly Lys Glu Leu Lys Asp 365 360 355 Pro Asp Gln Leu Tyr Thr Thr Leu Lys Asn Leu Leu Ala Gln Ile Lys 375 Ser His Pro Ser Ala Trp Pro Phe Met Glu Pro Val Lys Lys Ser Glu 390 395 Ala Pro Asp Tyr Tyr Glu Val Ile Arg Phe Pro Ile Asp Leu Lys Thr 405 410 Met Thr Glu Arg Leu Arg Ser Arg Tyr Tyr Val Thr Arg Lys Leu Phe 425 430 420 Val Ala Asp Leu Gln Arg Val Ile Ala Asn Cys Arg Glu Tyr Asn Pro 445 440 Pro Asp Ser Glu Tyr Cys Arg Cys Ala Ser Ala Leu Glu Lys Phe Phe 455 450 Tyr Phe Lys Leu Lys Glu Gly Gly Leu Ile Asp Lys 470

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2414 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

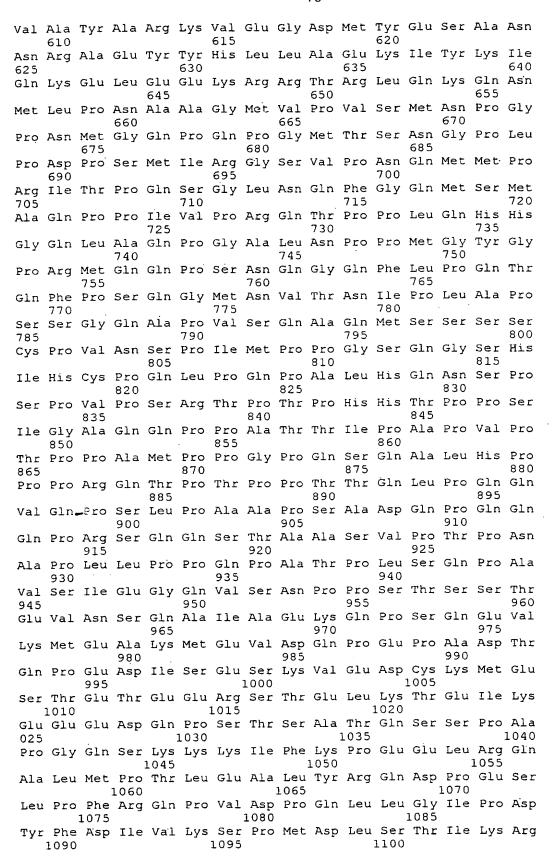
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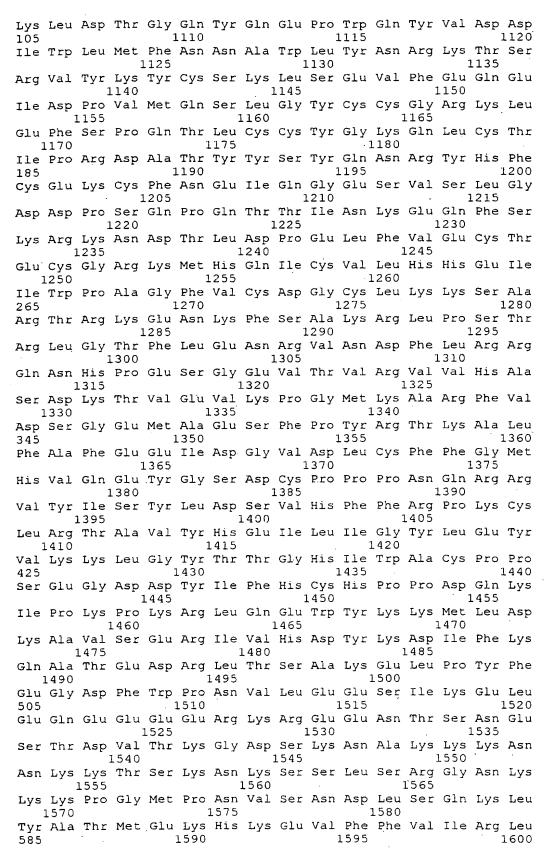


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Ala	Gly 130	Leu	Thr	Ser	Pro	Asn 135	Met	Gly	Met	Gly	Thr 140	Ser	Gly	Pro	Asn
Gln 145	Gly	Pro	Thr	Gln	Ser. 150	Thr	Gly	Met	Met	Asn 155	Ser	Pro	Val	Asn	Gln 160
Pro	Ala	Met	Gly	Met 165	Asn	Thr	Gly	Thr	Asn 170	Aļa	Gly	Met	Asn 	Pro 175	Gly
			180					185		Met			190	•	
		195					200			Gln		205			
	210		•			215				Leu	220				
225	-				230	_				Gly 235					240
		_		245					250	Asn				255	
-			260					265		Ala			270		
		275		_			280			Asn		285			
	290					295				Met Leu	300				
305					310					315 Asp					320
	_		-	325					330	His				335	
			340					345		Gln			350		٠
-	_	35 5					360			Met		365			
-	370					375				Ser	380				
385	-		_		390					395 Cys					400
				405					410	Gln				415	
	-		420	_				425		Leu			430		
		435	_				440			Ile		445			
	450					455				Tyr	460				
465	_		_		470					475 Gln					480
				485					490	Ser				495	
Pro	Met	Gly	500 Val	Asn	Gly	Gly	Val	505 Gly	Val	Gln	Thr	Pro	510 Ser	Leu	Leu
ser	Asp	515 Ser	Met	Leu	His		520 Ala	Ile	Asn	Ser		525 Asn	Pro	Met	Met
Ser	530 Glu	Asn	Ala	Ser		535 Pro	Ser	Leu	Gly	Pro	540 Met	Pro	Thr	Ala	
545 Gln	Pro	Ser	Thr		550 Gly	Ile	Arg	Lys		555 Trp	His	Glu	Asp		560 Thr
Gln	Asp	Leu	_	565 Asn	His	Leu	Val		570 Lys	Leu	Val	Glņ		575 Ile	Phe
Pro	Thr		580 Asp	Pro	Ala	Ala		585 Lys	Asp	Arg	Arg		590 Glu	Asn	Leu
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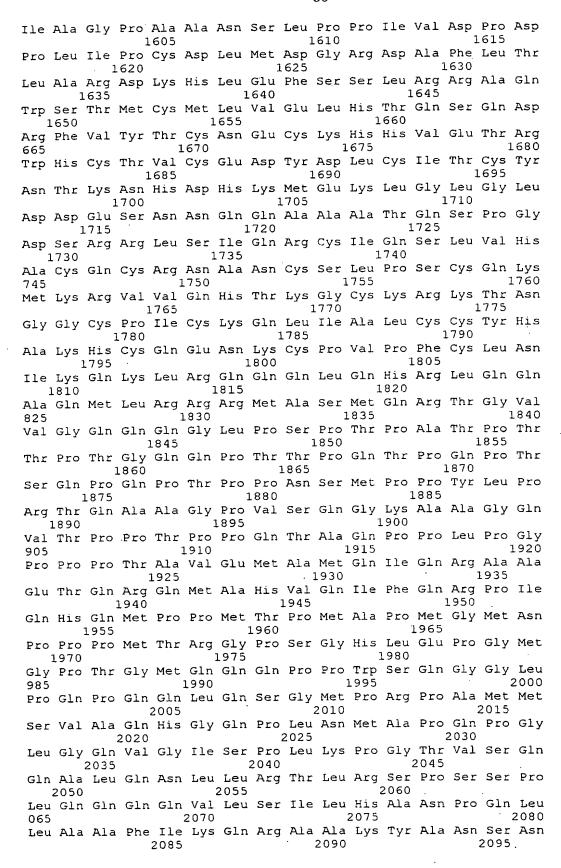
















Pro Gln Pro Ile Pro Gly Gln Pro Gly Met Pro Gln Gly Gln Pro Gly 2100 2105 Leu Gln Pro Pro Thr Met Pro Gly Gln Gln Gly Val His Ser Asn Pro 2115 2120 2125 Ala Met Gln Asn Met Asn Pro Met Gln Ala Gly Val Gln Arg Ala Gly 2130 2135 2140 Leu Pro Gln Gln Gln Pro Gln Gln Leu Gln Pro Pro Met Gly Gly 2150 2155 Met Ser Pro Gln Ala Gln Gln Met Asn Met Asn His Asn Thr Met Pro . 2165 2170 2175 Ser Gln Phe Arg Asp Ile Leu Arg Arg Gln Gln Met Met Gln Gln 2180 2185 2190 Gln Gln Gly Ala Gly Pro Gly Ile Gly Pro Gly Met Ala Asn His 2195 2200 2205 Asn Gln Phe Gln Gln Pro Gln Gly Val Gly Tyr Pro Pro Gln Pro Gln 2210 2215 2220 Gln Arg Met Gln His His Met Gln Gln Met Gln Gln Gly Asn Met Gly 2230 2235 Gln Ile Gly Gln Leu Pro Gln Ala Leu Gly Ala Glu Ala Gly Ala Ser 2245 2250 Leu Gln Ala Tyr Gln Gln Arg Leu Leu Gln Gln Met Gly Ser Pro 2260 2265 2270 Val Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu Pro Asn Gln 2275 2280 2285 Ala Gln Ser Pro His Leu Gln Gly Gln Gln Ile Pro Asn Ser Leu Ser 2290 2295 2300 Asn Gln Val Arg Ser Pro Gln Pro Val Pro Ser Pro Arg Pro Gln Ser 305 2310 2315 2320 Gln Pro Pro His Ser Ser Pro Ser Pro Arg Met Gln Pro Gln Pro Ser 2325 2330 2335 Pro His His Val Ser Pro Gln Thr Ser Ser Pro His Pro Gly Leu Val 2340 2345 2350 Ala Ala Gln Ala Asn Pro Met Glu Gln Gly His Phe Ala Ser Pro Asp 2355 2360 2365 Gln Asn Ser Met Leu Ser Gln Leu Ala Ser Asn Pro Gly Met Ala Asn 2370 2375 2380 Leu His Gly Ala Ser Ala Thr Asp Leu Gly Leu Ser Thr Asp Asn Ser 385 2390 2395 2400 Asp Leu Asn Ser Asn Leu Ser Gln Ser Thr Leu Asp Ile His

81

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

2405

- (A) LENGTH: 2441 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

 Met
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 Leu
 Leu
 Asp
 Gly
 Pro
 Pro
 Asn
 Pro
 Lys
 Arg
 Ala
 Lys

 Leu
 Ser
 Ser
 Pro
 Gly
 Phe
 Ser
 Ala
 Asn
 Asp
 Asn
 Thr
 Asp
 Phe
 Gly
 Ser

 Leu
 Phe
 Asn
 Asn
 Asp
 Leu
 Pro
 Asp
 Glu
 Leu
 Pro
 Asn
 Gly
 Asp
 Glu
 Leu
 Pro
 Asp
 Ala
 Ala
 Asp
 Asp
 Glu
 Leu
 Pro
 Asp
 Ala
 Asp
 A

2410



Ile	Asn	Pro	Gly	Ile 85	Gly	Asn	Val	Ser	Ala 90	Ser	Ser	Pro	Val	Gln 95	Gln
Gly	Leu	Gly	Gly 100	Gln	Ala	Gln	Gly	Gln 105	Pro	Asn	Ser	Thr	Asn 110	Met	Ala
Ser	Leu	Gly 115	Ala	Met	Gly	Lys	Ser 120	Pro	Leu	Asn	Gln	Gly 125	Asp	Ser	Ser
Thr	Pro 130	Asn	Leu	Pro	Lys	Gln 135	Ala	Ala	Ser	Thr	Ser 140	Gly	Pro	Thr	Pro
Pro 145	Ala	Ser	Gln	Ala	Leu 150	Asn	Pro	Gln	Ala	Gln 155	Lys	Gln	Val	Gly	Leu 160
	Thr			165					170					175	
	Ala		180					185					190		
_	His	195					200					205			
	Gly 210					215					220				
225	Pro		•		230					235					240
	Leu			245					250					255	
	Ala		260					265					270		
	Pro	275					280					285			
	Thr 290	_				295					300				
305	Leu				310					315					320
	Pro			325					330					335	
	Ala		340					345					350		
	Gln Glu	355					360					365			
_	370 Thr					375					380				
385	Ala		-		390					395					400
-	Trp			405					410					415	
	Asn	_	420					425					430		
_	Ser	435					440					445			
	450 Ala	_				455					460				
465	Arg				470					475					480
	Gln			485					490					495	
	His		500					505					510		
	Val	515					520					525			
	530 Ser	*				535					540				
545					550					555					560
		p	y	565			1		570					575	



Pro Thr Ala Ala Pro Pro Ser Ser Thr Gly Val Arg Lys Gly Trp His Glu His Val Thr Gln Asp Leu Arg Ser His Leu Val His Lys Leu Val Gln Ala Ile Phe Pro Thr Pro Asp Pro Ala Ala Leu Lys Asp Arg Arg Met Glu Asn Leu Val Ala Tyr Ala Lys Lys Val Glu Gly Asp Met Tyr. Glu Ser Ala Asn Ser Arg Asp Glu Tyr Tyr His Leu Leu Ala Glu Lys Ile Tyr Lys Ile Gln Lys Glu Leu Glu Glu Lys Arg Arg Thr Arg Leu His Lys Gln Gly Ile Leu Gly Asn Gln Pro Ala Leu Pro Ala Ser Gly Ala Gln Pro Pro Val Ile Pro Pro Ala Gln Ser Val Arg Pro Pro Asn Gly Pro Leu Pro Leu Pro Val Asn Arg Met Gln Val Ser Gln Gly Met Asn Ser Phe Asn Pro Met Ser Leu Gly Asn Val Gln Leu Pro Gln Ala Pro Met Gly Pro Arg Ala Ala Ser Pro Met Asn His Ser Val Gln Met Asn Ser Met Ala Ser Val Pro Gly Met Ala Ile Ser Pro Ser Arg Met Pro Gln Pro Pro Asn Met Met Gly Thr His Ala Asn Asn Ile Met Ala Gln Ala Pro Thr Gln Asn Gln Phe Leu Pro Gln Asn Gln Phe Pro Ser Ser Ser Gly Ala Met Ser Val Asn Ser Val Gly Met Gly Gln Pro Ala Ala Gln Ala Gly Val Ser Gln Gly Gln Glu Pro Gly Ala Ala Leu Pro Asn Pro Leu Asn Met Leu Ala Pro Gln Ala Ser Gln Leu Pro Cys Pro Pro Val Thr Gln Ser Pro Leu His Pro Thr Pro Pro Pro Ala Ser Thr Ala Ala Gly Met Pro Ser Leu Gln His Pro Thr Ala Pro Gly Met Thr Pro Pro Gln Pro Ala Ala Pro Thr Gln Pro Ser Thr Pro Val Ser Ser Gly Gln Thr Pro Thr Pro Thr Pro Gly Ser Val Pro Ser Ala Ala Gln Thr Gln Ser Thr Pro Thr Val Gln Ala Ala Gln Ala Gln Val Thr Pro Gln Pro Gln Thr Pro Val Gln Pro Pro Ser Val Ala Thr Pro Gln Ser Ser Gln Gln Pro Thr Pro Val His Thr Gln Pro Pro Gly Thr Pro Leu Ser Gln Ala Ala Ala Ser Ile Asp Asn Arg Val Pro Thr Pro Ser Thr Val Thr Ser Ala Glu Thr Ser Ser Gln Gln Pro Gly Pro Asp Val Pro Met Leu Glu Met Lys Thr Glu Val Gln Thr Asp Asp Ala Glu Pro Glu Pro Thr Glu Ser Lys Gly Glu Pro Arg Ser Glu Met Met Glu Glu Asp Leu Gln Gly Ser Ser Gln Val Lys Glu Glu Thr Asp Thr Thr Glu Gln Lys Ser Glu Pro Met Glu Val Glu Glu Lys Lys Pro Glu Val Lys Val Glu Ala Lys Glu Glu Glu Asn Ser Ser Asn Asp Thr Ala



Ser Gln Ser Thr Ser Pro Ser Gln Pro Arg Lys Lys Ile Phe Lys Pro 1075 1080 Glu Glu Leu Arg Gln Ala Leu Met Pro Thr Leu Glu Ala Leu Tyr Arg 1090 1095 1100 Gln Asp Pro Glu Ser Leu Pro Phe Arg Gln Pro Val Asp Pro Gln Leu 1110 1115 Leu Gly Ile Pro Asp Tyr Phe Asp Ile Val Lys Asn Pro Met Asp Leu 1125 1130 Ser Thr Ile Lys Arg Lys Leu Asp Thr Gly Gln Tyr Gln Glu Pro Trp 1140 1145 1150 Gln Tyr Val Asp Asp Val Arg Leu Met Phe Asn Asn Ala Trp Leu Tyr 1155 1160 1165 Asn Arg Lys Thr Ser Arg Val Tyr Lys Phe Cys Ser Lys Leu Ala Glu 1170 1175 1180 Val Phe Glu Gln Glu Ile Asp Pro Val Met Gln Ser Leu Gly Tyr Cys 1190 1195 1200 Cys Gly Arg Lys Tyr Glu Phe Ser Pro Gln Thr Leu Cys Cys Tyr Gly 1205 1210 . 1215 Lys Gln Leu Cys Thr Ile Pro Arg Asp Ala Ala Tyr Tyr Ser Tyr Gln 1220 1225 1230 Asn Arg Tyr His Phe Cys Gly Lys Cys Phe Thr Glu Ile Gln Gly Glu 1235 1240 1245 Asn Val Thr Leu Gly Asp Asp Pro Ser Gln Pro Gln Thr Thr Ile Ser 1250 1255 1260 Lys Asp Gln Phe Glu Lys Lys Lys Asn Asp Thr Leu Asp Pro Glu Pro 265 1270 1275 Phe Val Asp Cys Lys Glu Cys Gly Arg Lys Met His Gln Ile Cys Val 1285 1290 1295 1285 1290 Leu His Tyr Asp Ile Ile Trp Pro Ser Gly Phe Val Cys Asp Asn Cys 1300 1305 . 1310 Leu Lys Lys Thr Gly Arg Pro Arg Lys Glu Asn Lys Phe Ser Ala Lys 1315 1320 1325 Arg Leu Gln Thr Thr Arg Leu Gly Asn His Leu Glu Asp Arg Val Asn 1330 1335 1340 Lys Phe Leu Arg Arg Gln Asn His Pro Glu Ala Gly Glu Val Phe Val Arg Val Val Ala Ser Ser Asp Lys Thr Val Glu Val Lys Pro : 1y Met 1365 1370 1375 Lys Ser Arg Phe Val Asp Ser Gly Glu Met Ser Glu Ser Phe Pro Tyr 1380 1385 1390 Arg Thr Lys Ala Leu Phe Ala Phe Glu Glu Ile Asp Gly Val Asp Val 1395 1400 Cys Phe Phe Gly Met His Val Gln Asp Thr Ala Leu Ile Ala Pro His 1410 1415 1420 Gln Ile Gln Gly Cys Val Tyr Ile Ser Tyr Leu Asp Ser Ile His Phe 1430 1435 1440 Phe Arg Pro Arg Cys Leu Arg Thr Ala Val Tyr His Glu Ile Leu Ile 1445 1450 Gly Tyr Leu Glu Tyr Val Lys Lys Leu Val Tyr Val Thr Ala His Ile 1465 1470 1460 Trp Ala Cys Pro Pro Ser Glu Gly Asp Asp Tyr Ile Phe His Cys His 1475 1480 1485 Pro Pro Asp Gln Lys Ile Pro Lys Pro Lys Arg Leu Gln Glu Trp Tyr 1490 1495 1500 Lys Lys Met Leu Asp Lys Ala Phe Ala Glu Arg Ile Ile Asn Asp Tyr 505 1510 1515 Lys Asp Ile Phe Lys Gln Ala Asn Glu Asp Arg Leu Thr Ser Ala Lys 1530 1525 Glu Leu Pro Tyr Phe Glu Gly Asp Phe Trp Pro Asn Val Leu Glu Glu 1540 1545 1550 Ser Ile Lys Glu Leu Glu Gln Glu Glu Glu Arg Lys Lys Glu Glu 1560 1555



Ser Thr Ala Ala Ser Glu Thr Pro Glu Gly Ser Gln Gly Asp Ser Lys 1575 Asn Ala Lys Lys Lys Asn Asn Lys Lys Thr Asn Lys Asn Lys Ser Ser 1590 1595 Ile Ser Arg Ala Asn Lys Lys Lys Pro Ser Met Pro Asn Val Ser Asn 1605 1610 1615 Asp Leu Ser Gln Lys Leu Tyr Ala Thr Met Glu Lys His Lys Glu Val 1620 1625 Phe Phe Val Ile His Leu His Ala Gly Pro Val Ile Ser Thr Gln Pro 1640 1645 Pro Ile Val Asp Pro Asp Pro Leu Leu Ser Cys Asp Leu Met Asp Gly 1650 . 1655 1660 Arg Asp Ala Phe Leu Thr Leu Ala Arg Asp Lys His Trp Glu Phe Ser 1670 1675 Ser Leu Arg Arg Ser Lys Trp Ser Thr Leu Cys Met Leu Val Glu Leu 1685 1690 1695 His Thr Gln Gly Gln Asp Arg Phe Val Tyr Thr Cys Asn Glu Cys Lys 1700 1705 His His Val Glu Thr Arg Trp His Cys Thr Val Cys Glu Asp Tyr Asp 1715 1720 1725Leu Cys Ile Asn Cys Tyr Asn Thr Lys Ser His Thr His Lys Met Val 1730 1735 1740 Lys Trp Gly Leu Gly Leu Asp Asp Glu Gly Ser Ser Gln Gly Glu Pro 745 1750 1755 1760 Gln Ser Lys Ser Pro Gln Glu Ser Arg Arg Leu Ser Ile Gln Arg Cys 1765 1770 1775 Ile Gln Ser Leu Val His Ala Cys Gln Cys Arg Asn Ala Asn Cys Ser 1785 1780 Leu Pro Ser Cys Gln Lys Met Lys Arg Val Val Gln His Thr Lys Gly 1795 1800 1805 Cys Lys Arg Lys Thr Asn Gly Gly Cys Pro Val Cys Lys Gln Leu Ile 1815 1820 Ala Leu Cys Cys Tyr His Ala Lys His Cys Gln Glu Asn Lys Cys Pro 1835 1830 Val Pro Phe Cys Leu Asn Ile Lys His Asn Val Arg Gln Gln Gln Ile 1845 1850 1855 Gln His Cys Leu Gln Gln Ala Gln Leu Met Arg Arg Arg Met Ala Thr 1860 1865 1870 Met Asn Thr Arg Asn Val Pro Gln Gln Ser Leu Pro Ser Pro Thr Ser 1875 1880 1885 Ala Pro Pro Gly Thr Pro Thr Gln Gln Pro Ser Thr Pro Gln Thr Pro 1895 1900 Gln Pro Pro Ala Gln Pro Gln Pro Ser Pro Val Asn Met Ser Pro Ala 1910 1915 1920 Gly Phe Pro Asn Val Ala Arg Thr Gln Pro Pro Thr Ile Val Ser Ala 1925 1930 Gly Lys Pro Thr Asn Gln Val Pro Ala Pro Pro Pro Pro Ala Gln Pro 1945 1950 Pro Pro Ala Ala Val Glu Ala Ala Arg Gln Ile Glu Arg Glu Ala Gln 1955 1960 1965 Gln Gln His Leu Tyr Arg Ala Asn Ile Asn Asn Gly Met Pro Pro 1975 1980 Gly Arg Asp Gly Met Gly Thr Pro Gly Ser Gln Met Thr Pro Val Gly 1990 1995 2000 Leu Asn Val Pro Arg Pro Asn Gln Val Ser Gly Pro Val Met Ser Ser 2005 2010 2015 Met Pro Pro Gly Gln Trp Gln Gln Ala Pro Ile Pro Gln Gln Pro 2020 2025 2030 Met Pro Gly Met Pro Arg Pro Val Met Ser Met Gln Ala Gln Ala Ala 2035 2040 2045 Val Ala Gly Pro Arg Met Pro Asn Val Gln Pro Asn Arg Ser Ile Ser 2055 ·





Pro Ser Ala Leu Gln Asp Leu Leu Arg Thr Leu Lys Ser Pro Ser Ser 2075 2080 2070 065 Pro Gln Gln Gln Gln Val Leu Asn Ile Leu Lys Ser Asn Pro Gln 2085 2090 2095 Leu Met Ala Ala Phe Ile Lys Gln Arg Thr Ala Lys Tyr Val Ala Asn 2100 2105 2110 Gln Pro Gly Met Gln Pro Gln Pro Gly Leu Gln Ser Gln Pro Gly Met 2115 2120 2125 Gln Pro Gln Pro Gly Met His Gln Gln Pro Ser Leu Gln Asn Leu Asn 2135 2140 2130 Ala Met Gln Ala Gly Val Pro Arg Pro Gly Val Pro Pro Pro Gln Pro 145 2150 2155 2160 Ala Met Gly Gly Leu Asn Pro Gln Gly Gln Ala Leu Asn Ile Met Asn 2165 2170 2175 Pro Gly His Asn Pro Asn Met Thr Asn Met Asn Pro Gln Tyr Arg Glu 2180 2185 2190 Met Val Arg Arg Gln Leu Leu Gln His Gln Gln Gln Gln Gln Gln 2200 2195 2205 Gln Gln Gln Gln Gln Gln Gln Asn Ser Ala Ser Leu Ala Gly Gly 2210 . 2215 2220 Met Ala Gly His Ser Gln Phe Gln Gln Pro Gln Gly Pro Gly Gly Tyr 2235 2230 Ala Pro Ala Met Gln Gln Gln Arg Met Gln Gln His Leu Pro Ile Gln 2245 2250 2255 Gly Ser Ser Met Gly Gln Met Ala Ala Pro Met Gly Gln Leu Gly Gln 2260 2265 2270 Met Gly Gln Pro Gly Leu Gly Ala Asp Ser Thr Pro Asn Ile Gln Gln 2275 2280 2285 Ala Leu Gln Gln Arg Ile Leu Gln Gln Gln Gln Met Lys Gln Gln Ile 2290 2295 2300 Gly Ser Pro Gly Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu 305 2310 2315 2320 Ser Gly Gln Pro Gln Ala Ser His Leu Pro Gly Gln Gln Ile Ala Thr 2325 2330 2335 Ser Leu Ser Asn Gln Val Arg Ser Pro Ala Pro Val Gln Ser Pro Arg 2340 2345 2350 Pro Gln Ser Gln Pro Pro His Ser Ser Pro Ser Pro Arg Ile Gln Pro 2360 2365 Gln Pro Ser Pro His His Val Ser Pro Gln Thr Gly Thr Pro His Pro 2370 2375 2380 Gly Leu Ala Val Thr Met Ala Ser Ser Met Asp Gln Gly His Leu Gly 2390 2395 385 Asn Pro Glu Gln Ser Ala Met Leu Pro Gln Leu Asn Thr Pro Asn Arg 2405 2410 2415 Ser Ala Leu Ser Ser Glu Leu Ser Leu Val Gly Asp Thr Thr Gly Asp 2420 2425 Thr Leu Glu Lys Phe Val Glu Gly Leu 2435 2440

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(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 813 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ala Glu Ala Gly Gly Ala Gly Ser Pro Ala Leu Pro Pro Ala Pro 1 10 15



Pro	His	Gļy	Ser 20	Pro	Arg	Thr	Leu	Ala 25	Thr	Ala	Ala	Gly	Ser 30	Ser.	Ala
Ser	Cys	Gly 35		Ala	Thr	Pro	Val 40	Ala	Ala	Ala	Gly	Thr 45	Ala	Glu	Gly
Pro	Gly 50		Gly	Gly	Ser	Ala 55	Arg	Ile	Ala	Val	Lys 60	Lys	Ala	Gln	Leu
Arg 65		Ala	Pro	Arg	Ala 70	Lys	Lys	Leu	Glu	Lys 75	Leu	Gly.	Val	Tyr	Ser 80
	Cys	Lys	Ala	Glu 85	Glu	Ser	Cys	Lys	Cys 90	Asn	Gly	Trp	Lys	Asn 95	Pro
Asn	Pro	Ser	Pro 100	Thr	Pro	Pro	Arg	Gly 105	Asp	Leu	Gln	Gln	Ile 110	Ile	Val
Ser	Leu	Thr 115	Glu	Ser	Cys	Arg	Ser 120	Cys	Ser	His	Ala	Leu 125	Ala	Ala	His
Val	Ser 130	His	Leu	Glu	Asn	Val 135	Ser	Glu	Glu	Glu	Met 140	Asp	Arg	Leu	Leu
145			• •		150		Tyr			155					160
				165			Val		170					175	
-	_		180				Gly	185					190		
	-	195					Lys 200					205			
	210					215	Ser				220				
225					230		Met			235					240
				245			Arg		250					255	
		-	260			•	Tyr	265					270		
		275					Leu 280					285			
	290					295	Ser				300				
305					310		Lys			315	•				320
				325			Phe Ser		330					335	
			340				Pro	345					350		
		355					360 Leu					365			
	370					375	Thr				380				
385					390		Glu			395					400
•				405			Ser		410					415	
			420				Ser	425					430		
		435					440 Leu					445			
	450					455				-	460				
465	Λια	A. y	<u> </u>	C_Lu	470	• • • 9	9	~- y		475					480
	Asn	Ser	Leu	Asn 485		Lys	Pro	Asn	Lys 490	Lys	Ile	Leu	Met	Trp 495	Leu
Val	Gly	Leu	Gln 500	Asn	Val	Phe	Ser	His 505	Gln	Leu	Pro	Arg	510		Lys`







Glu Tyr Ile Thr Arg Leu Val Phe Asp Pro Lys His Lys Thr Leu Ala 520 525 515 Leu Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe 535 540 Pro Ser Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val Thr Ser Asn 555 550 Glu Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His Leu Lys Glu 570 575 565 Tyr His Ile Lys His Glu Ile Leu Asn Phe Leu Thr Tyr Ala Asp Glu 585 Tyr Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Glu Ile Lys 600 595 Ile Pro Lys Thr Lys Tyr Val Gly Tyr Ile Lys Asp Tyr Glu Gly Ala 620 615 Thr Leu Met Gly Cys Glu Leu Asn Pro Gln Ile Pro Tyr Thr Glu Phe 630 635 Ser Val Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu 650 645 Arg Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe 665 670 Lys Asp Gly Val Arg Gln Ile Pro Ile Glu Ser Ile Pro Gly Ile Arg 675 680 685 Glu Thr Gly Trp Lys Pro Ser Gly Lys Glu Lys Ser Lys Glu Pro Lys 700 695 Asp Pro Glu His Val Tyr Ser Thr Leu Lys Asn Ile Leu Gln Gln Val 710 715 Lys Asn His Pro Asn Ala Trp Pro Phe Met Glu Pro Val Lys Arg Thr 735 725 730 Glu Ala Pro Gly Tyr Tyr Glu Val Ile Arg Phe Pro Met Asp Leu Lys 745 Thr Met Ser Glu Arg Leu Arg Asn Arg Tyr Tyr Val Ser Lys Lys Leu 760 Phe Met Ala Asp Leu Gln Arg Val Phe Thr Asn Cys Lys Glu Tyr Asn 775 780 Pro Pro Glu Ser Glu Tyr Tyr Lys Cys Ala Ser Ile Leu Glu Lys Phe 790 795 Phe Phe-Ser Lys Ile Lys Glu Ala Gly Leu Ile Asp Lys 805

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(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 50 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2204 base pairs





(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ACCCACTCCC CCCAGAGCCG ACCTGCAGCA AATAATTGTC AGTCTAACAG AATCCTGTCG 60 GAGTTGTAGC CATGCCCTAG CTGCTCATGT TTCCCACCTG GAGAATGTGT CAGAGGAAGA 120 AATGAACAGA CTCCTGGGAA TAGTATTGGA TGTGGAATAT CTCTTTACCT GTGTCCACAA GGAAGAAGAT GCAGATACCA AACAAGTTTA TTTCTATCTA TTTAAGCTCT TGAGAAAGTC 240 TATTTTACAA AGAGGAAAAC CTGTGGTTGG AAGGCTCTTT GGAAAAGAAA CCCCCATTTG 300 AAAAACCTAG CATTGAACAG GGTGTGAATA ACTTTGTGCA GTACAAATTT AGTCACCTGC 360 CAGCAAAAAG AAAGGCAAAC CAATAGTTGA GTTGGCAAAA ATGTTCCTAA ACCGCATCAC CTATTGGCAT CTGGAGGCAC CATCTCAACG AGACTGCGAT CTCCAATGAT GATATTCTGG ATACAAAGAG AACTACACAA GGTGGCTGTG TTACTGCAAC GTGCCACAGT TCTGCGACAG TCTACCTCGG TACGAAACCA CACAGGTGTT TGGGAGAACA TCGTTCGCTC GGTCTTCACT GTTATGAGGC GACAACTCCT GGAACAAGCA AGACAGGAAA AAGATAAACT GCCTCTTGAA 660 AAACGAACTC TAATCCTCAC TCATTTCCCA AAATTTCTGT CCATGCTAGA AGAAGAAGTA .720 TATAGTCAAA ACTCTCCCAT CTGGGATCAC CATTTTCTCT CAGCCTCTTC CAGAACCAGC CAGCTAGGCA TCCAAACAGT TATCAATCAC CTCCTGTGGC TGGGACAATT TCATACAATT CAACCTCATC TTCCCTTGAG CAGCCAAACG CAGGGAGCAG CAGTCCTGCC TGCAAAGCCT 780 840 900 CTTCTGGACT TGAGGCAAAC CCAGGAGAAA AGAGGAAAAT GACTGATTCT CATGTTCTGG 960 AGGAGGCCAA GAAACCCCGA GTTATGGGGG ATATTCCGAT GGAATTAATC AACGAGGTTA 1020 TGTCTACCAT CACGGACCCT GCAGCAATGC TTGGACCAGA GACCAATTTT CTGTCAGCAC 1080 ACTCGGCCAG GGATGAGGCG GCAAGGTTGG AAGAGCGCAG GGGTGTAATT GAATTTCACG 1140 TGGTTGGCAA TTCCCTCAAC CAGAAACCAA ACAAGAAGAT CCTGATGTGG CTGGTTGGCC 1200 TACAGAACGT TTTCTCCCAC CAGCTGCCCC GAATGCCAAA AGAATACATC ACACGGCTCG 1260 TCTTTGACCC GAAACACAAA ACCCTTGCTT TAATTAAAGA TGGCCGTGTT ATTGGTGGTA 1320
TCTGTTTCCG TATGTTCCCA TCTCAAGGAT TCACAGAGAT TGTCTTCTGT GCTGTAACCT 1380
CAAATGAGCA AGTCAAGGGC TATGGAACAC ACCTGATGAA TCATTTGAAA GAATATCACA 1440
TAAAGCATGA CATCCTGAAC TTCCTCACAT ATGCAGATGA ATATGCAATT GGATACTTTA 1500 AGAAACAGGG TTTCTCCAAA GAAATTAAAA TACCTAAAAC CAAATATGTT GGCTATATCA 1560 AGGATTATGA AGGAGCCACT TTAATGGGAT GTGAGCTAAA TCCACGGATC CCGTACACAG 1620 AATTTTCTGT CATCATTAAA AAGCAGAAGG AGATAATTAA AAAACTGATT GAAAGAAAAC 1680 AGGCACAAAT TCGAAAAGTT TACCCTGGAC TTTCATGTTT TAAAGATGGA GTTCGACAGA 1740 TTCCTATAGA AAGCATTCCT GGAATTAGAG AGACAGGCTG GAAACCGAGT GGAAAAGAGA 1800 AAAGTAAAGA GCCCAGAGAC CCTGACCAGC TTTACAGCAC GCTCAAGAGC ATCCTCCAGC AGGTGAAGAG CCATCAAAGC GCTTGGCCCT TCATGGAACC TGTGAAGAGA ACAGAAGCTC 1920 CAGGATATTA TGAAGTTATA AGGTCCCCCA TGGATCTCAA AACCATGAGT GAACGCCTCA AGAATAGGTA CTACGTGTCT AAGAAATTAT TCATGGCAGA CTTACAGCGA GTCTTTACCA 2040 ATTGCAAAGA GTACAACGCC CCTGAGAGTG AATACTACAA ATGTGCCAAT ATCCTGGAGA 2100 AATTCTTCTT CAGTAAAATT AAGGAAGCTG GATTAATTGA CAAGTGATTT TTTTTCCCCC 2160 TCTGCTTCTT AGAAACTCAC CAAGCAGTGT GCCTAAAGCA AGGT

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(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2093 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GAATTCCGGC	GAAACCACTC	ATGTCTTTGG	GCGAAGCCTT	CTCCGGTCCA	TTTTCACCGT	60
TACCCGCCGG	CAGCTGCTGG	AAAAGTTCCG	AGTGGAGAAG	GACAAATTGG	TGCCCGAGAA	120
GAGGACCCTC	ATCCTCACTC	ACTTCCCCAA	GTAAGGCTCC	TTCTGGCCTA	CCAGGATTTG	180
GCCCCAAGTT	CACATCCTCC	CTGTTGTCCC	CTTTTTTCCA	GGAAGGCTTC	CTGGATTGGT	240
CCCTCCTCTC	CCTCCATGGG	CCTTTTGGGA	TCTGGGCGTC	TACCTGGCAG	ACTTGCCCAT	300
GGCCCAGAAG	CAACTTGCTA	GTACTAGTCT	GGGGATGGCA	GATTCCTGTC	CATGCTGGAG	360
GAGGAGATCT	ATGGGGCAAA	CTCTCCAATC	TEGERETERS	GCTTCACCAT	GCCACCCTCA	420





GAGGGGACAC AGCTGGTTCC CCGGCCAGCT TCAGTCAGTG CAGCGGTTGT TCCCAGCACC CCCATCTTCA GCCCCAGCAT GGGTGGGGGC AGCAACAGCT CCCTGAGTCT GGATTCTGCA GGGGCCGAGC CTATGCCAGG CGAGAAGAGG ACGCTCCCAG AGAACCTGAC CCTGGAGGAT 600 GCCAAGCGGC TCCGTGTGAT GGGTGACATC CCCATGGAGC TGGTCAATGA GGTCATGCTG 660 ACCATCACTG ACCCTGCTGC CATGCTGGGG CCTGAGACGA GCCTGCTTTC GGCCAATGCG 720 GCCCGGGATG AGACAGCCCG CCTGGAGGAG CGCCGCGGCA TCATCGAGTT CCATGTCATC GGCAACTCAC TGACGCCCAA GGCCAACCGG CGGGTGTTGC TGTGGCTCGT GGGGCTGCAG 840 AATGTCTTTT CCCACCAGCT GCCGCGCATG CCTAAGGAGT ATATCGCCCG CCTCGTCTTT 900 GACCCGAAGC ACAAGACTCT GGCCTTGATC AAGGATGGGC GGGTCATCGG TGGCATCTGC 960 TTCCGCATGT TTCCCACCCA GGGCTTCACG GAGATTGTCT TCTGTGCTGT CACCTCGAAT 1020 GAGCAGGTCA AGGGTTATGG GACCCACCTG ATGAACCACC TGAAGGAGTA TCACATCAAG 1080 CACAACATTC TCTACTTCCT CACCTACGCC GACGAGTACG CCATCGGCTA CTTCAAAAAG 1140 CAGGGTTTCT CCAAGGACAT CAAGGTGCCC AAGAGCCGCT ACCTGGGCTA CATCAAGGAC 1200 TACGAGGGAG CGACGCTGAT GGAGTGTGAG CTGAATCCCC GCATCCCCTA CACGGAGCTG 1260 TCCCACATCA TCAAGAAGCA GAAAGAGATC ATCAAGAAGC TGATTGAGCG CAAACAGGCC 1320 CAGATCCGCA AGGTCTACCC GGGGCTCAGC TGCTTCAAGG AGGGCGTGAG GCAGATCCCT 1380 GTGGAGAGCG TTCCTGGCAT TCGAGAGACA GGCTGGAAGC CATTGGGGAA GGAGAAGGGG 1440 AAGGAGCTGA AGGACCCCGA CCAGCTCTAC ACAACCCTCA AAAACCTGCT GGCCCAAATC AAGTCTCACC CCAGTGCCTG GCCCTTCATG GAGCCTGTGA AGAAGTCGGA GGCCCCTGAC 1560 TACTACGAGG TCATCCGCTT CCCCATTGAC CTGAAGACCA TGACTGAGCG GCTGCGAAGC 1620 CGCTACTACG TGACCCGGAA GCTCTTTGTG GCCGACCTGC AGCGGGTCAT CGCCAACTGT 1680 CGCGAGTACA ACCCCCGGA CAGCGAGTAC TGCCGCTGTG CCAGCGCCCT GGAGAAGTTC TTCTACTTCA AGCTCAAGGA GGGAGGCCTC ATTGACAAGT AGGCCCATCT TTGGGCCGCA 1800 CCCACGGACC CGACTCAGCT TGAGACACTC CAGCCAAGGG TCCTCCGGAC CCGATCCTGC 1920 AGCTCTTTCT GGACCTTCAG GCACCCCCAA GCGTGCAGCT CTGTCCCAGC CTTCACTGTG 1980 TGTGAGAGGT CTCCTGGGTT GGGGCCCAGC CCCTCTAGAG TAGCTGGTGG CCAGGGATGA 2040 ACCTTGCCCA GCCGTGGTGG CCCCCAGGCC TGGTCCCCAA GAGCCCGGAA TTC

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(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9046 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CCTTGTTTGT	GTGCTAGGCT	GGGGGGAGA	GAGGGCGAGA	GAGAGCGGGC	GAGAGTGGGC	60
AAGCAGGACG	CCGGGCTGAG	TGCTAACTGC	GGGACGCAGA	GAGTGCGGAG	GGGAGTCGGG	120
TCGGAGAGAG	GCGGCAGGGG	CCAGAACAGT	GGCAGGGGGC	CCGGGGCGCA	CGGGCTGAGG	180
CGACCCCCAG	CCCCCTCCCG	TCCGCACACA	CCCCCACCGC	GGTCCAGCAG	CCGGGCCGGC	240
GTCGACGCTA	GGGGGGACCA	TTACATAACC	CGCGCCCCGG	CCGTCTTCTC	CCGCCGCCGC	300
GGCGCCCGAA	CTGAGCCCGG	GGCGGGCGCT	CCAGCACTGG	CCGCCGGCGT	GGGGCGTAGC	360
AGCGGCCGTA	TTATTATTTC	GCGGAAAGGA	AGGCGAAGGA	GGGGAGCGCC	GGCGCGAGGA	420
GGGGCCGCCT	GCGCCGCCG	CCGGAGCGGG	GCCTCCTCGG	TGGGCTCCGC	GTCGGCGCGG	480
GCGTGCGGGC	GGCGCTGCTC	GGCCCGGCCC	CCTCGGCCCT	CTGGTCCGGC	CAGCTCCGCT	540
CCCGGCGTCC	TTGCCGCGCC	TCCGCCGGCC	GCCGCGCGAT	GTGAGGCGGC	GGCGCCAGCC	600
TGGCTCTCGG	CTCGGGCGAG	TTCTCTGCGG	CCATTAGGGG	CCGGTGCGGC	GGCGGCGG	660
AGCGCGGCGG	CAGGAGGAGG	GTTCGGAGGG	TGGGGGCGCA	GGCCCGGGAG	GGGGCACCGG	720
GAGGAGGTGA	GTGTCTCTTG	TCGCCTCCTC	CTCTCCCCCC	TTTTCGCCCC	CGCCTCCTTG	780
TGGCGATGAG	AAGGAGGAGG	ACAGCGCCGA	GGAGGAAGAG	GTTGATGGCG	GCGGCGGAGC	840
TCCGAGAGAC	CTCGGCTGGG	CAGGGGCCGG	CCGTGGCGGG	CCGGGGACTG	CGCCTCTAGA	900
GCCGCGAGTT	CTCGGGAATT	CGCCGCAGCG	GACCGGCCTC	GGCGAATTTG	TGCTCTTGTG	960
CCCTCCTCCG	GGCTTGGGCC	AGGCCGGCCC	CTCGCACTTG	CCCTTACCTT	TTCTATCGAG	1020
TCCGCATCCC	TCTCCAGCCA	CTGCGACCCG	GCGAAGAGAA	AAAGGAACTT	CCCCCACCCC	1080
CTCGGGTGCC	GTCGGAGCCC	CCCAGCCCAC	CCCTGGGTGC	GGCGCGGGGA	CCCCGGGCCG	1140
AAGAAGAGAT	TTCCTGAGGA	TTCTGGTTTT	CCTCGCTTGT	ATCTCCGAAA	GAATTAAAAA	1200
TGGCCGAGAA	TGTGGTGGAA	CCGGGGCCGC	CTTCAGCCAA	GCGGCCTAAA	CTCTCATCTC	1260
CGGCCCTCTC	GGCGTCCGCC	AGCGATGGCA	CAGATTTTGG	CTCTCTATTT	GACTTGGAGC	1320
ACGACTTACC	AGATGAATTA	ATCAACTCTA	CAGAATTGGG	ACTAACCAAT	GGTGGTGATA	1380





TTAATCAGCT TCAGACAAGT CTTGGCATGG TACAAGATGC AGCTTCTAAA CATAAACAGC TGTCAGAATT GCTGCGATCT GGTAGTTCCC CTAACCTCAA TATGGGAGTT GGTGGCCCAG GTCAAGTCAT GGCCAGCCAG GCCCAACAGA GCAGTCCTGG ATTAGGTTTG ATAAATAGCA TGGTCAAAAG CCCAATGACA CAGGCAGGCT TGACTTCTCC CAACATGGGG ATGGGCACTA GTGGACCAAA TCAGGGTCCT ACGCAGTCAA CAGGTATGAT GAACAGTCCA GTAAATCAGC 1680 CTGCCATGGG AATGAACACA GGGACGAATG CGGGCATGAA TCCTGGAATG TTGGCTGCAG . 1740 GCAATGGACA AGGGATAATG CCTAATCAAG TCATGAACGG TTCAATTGGA GCAGGCCGAG 1800 GGCGACAGGA TATGCAGTAC CCAAACCCAG GCATGGGAAG TGCTGGCAAC TTACTGACTG AGCCTCTTCA GCAGGGCTCT CCCCAGATGG GAGGACAAAC AGGATTGAGA GGCCCCCAGC CTCTTAAGAT GGGAATGATG AACAACCCCA ATCCTTATGG TTCACCATAT ACTCAGAATC CTGGACAGCA GATTGGAGCC AGTGGCCTTG GTCTCCAGAT TCAGACAAAA ACTGTACTAT 2040 CAAATAACTT ATCTCCATTT GCTATGGACA AAAAGGCAGT TCCTGGTGGA GGAATGCCCA 2100 ACATGGGTCA ACAGCCAGCC CCGCAGGTCC AGCAGCCAGG TCTGGTGACT CCAGTTGCCC 2160 AAGGGATGGG TTCTGGAGCA CATACAGCTG ATCCAGAGAA GCGCAAGCTC ATCCAGCAGC 2220 AGCTTGTTCT CCTTTTGCAT GCTCACAAGT GCCAGCGCCG GGAACAGGCC AATGGGGAAG 2280 TGAGGCAGTG CAACCTTCCC CACTGTCGCA CAATGAAGAA TGTCCTAAAC CACATGACAC 2340 ACTGCCAGTC AGGCAAGTCT TGCCAAGTGG CACACTGTGC ATCTTCTCGA CAAATCATTT CACACTGGAA GAATTGTACA AGACATGATT GTCCTGTGTG TCTCCCCCTC AAAAATGCTG GTGATAAGAG AAATCAACAG CCAATTTTGA CTGGAGCACC CGTTGGACTT GGAAATCCTA 2520 GCTCTCTAGG GGTGGGTCAA CAGTCTGCCC CCAACCTAAG CACTGTTAGT CAGATTGATC CCAGCTCCAT AGAAAGAGCC TATGCAGCTC TTGGACTACC CTATCAAGTA AATCAGATGC CGACACACC CCAGGTGCAA GCAAAGAACC AGCAGAATCA GCAGCCTGGG CAGTCTCCCC AAGGCATGCG GCCCATGAGC AACATGAGTG CTAGTCCTAT GGGAGTAAAT GGAGGTGTAG GAGTTCAAAC GCCGAGTCTT CTTTCTGACT CAATGTTGCA TTCAGCCATA AATTCTCAAA 2820 ACCCAATGAT GAGTGAAAAT GCCAGTGTGC CCTCCCTGGG TCCTATGCCA ACAGCAGCTC 2880 AACCATCCAC TACTGGAATT CGGAAACAGT GGCACGAAGA TATTACTCAG GATCTTCGAA 2940 ATCATCTTGT TCACAAACTC GTCCAAGCCA TATTTCCTAC GCCGGATCCT GCTGCTTTAA AAGACAGACG GATGGAAAAC CTAGTTGCAT ATGCTCGGAA AGTTGAAGGG GACATGTATG AATCTGCAAA CAATCGAGCG GAATACTACC ACCTTCTAGC TGAGAAAATC TATAAGATCC 3120 AGAAAGAACT AGAAGAAAAA CGAAGGACCA GACTACAGAA GCAGAACATG CTACCAAATG 3180 CTGCAGGCAT GGTTCCAGTT TCCATGAATC CAGGGCCTAA CATGGGACAG CCGCAACCAG GAATGACTTC TAATGGCCCT CTACCTGACC CAAGTATGAT CCGTGGCAGT GTGCCAAACC 3240 AGATGATGCC TCGAATAACT CCACAATCTG GTTTGAATCA ATTTGGCCAG ATGAGCATGG 3360 CCCAGCCCC TATTGTACCC CGGCAAACCC CTCCTCTCA GCACCATGGA CAGTTGGCTC AACCTGGAGC TCTCAACCC CCTATGGGCT ATGGGCCTCG TATGCAACAG CCTTCCAACC 3420 3480 AGGGCCAGTT CCTTCCTCAG ACTCAGTTCC CATCACAGGG AATGAATGTA ACAAATATCC CTTTGGCTCC GTCCAGCGGT CAAGCTCCAG TGTCTCAAGC ACAAATGTCT AGTTCTTCCT GCCCGGTGAA CTCTCCTATA ATGCCTCCAG GGTCTCAGGG GAGCCACATT CACTGTCCCC AGCTTCCTCA ACCAGCTCTT CATCAGAATT CACCCTCGCC TGTACCTAGT CGTACCCCCA CCCCTCACCA TACTCCCCCA AGCATAGGGG CTCAGCAGCC ACCAGCAACA ACAATTCCAG 3780 CCCCTGTTCC TACACCACCA GCCATGCCAC CTGGGCCACA GTCCCAGGCT CTACATCCCC CTCCAAGGCA GACACCTACA CCACCAACAA CACAACTTCC CCAACAAGTG CAGCCTTCAC 3900 TTCCTGCTGC ACCTTCTGCT GACCAGCCC AGCAGCAGCC TCGCTCACAG CAGAGCACAG CAGCGTCTGT TCCTACCCCA AACGCACCGC TGCTTCCTCC GCAGCCTGCA ACTCCACTTT CCCAGCCAGC TGTAAGCATT GAAGGACAGG TATCAAATCC TCCATCTACT AGTAGCACAG 3960 4020 4080 AAGTGAATTC TCAGGCCATT GCTGAGAAGC AGCCTTCCCA GGAAGTGAAG ATGGAGGCCA 4140 AAATGGAAGT GGATCAACCA GAACCAGCAG ATACGCAGCC GGAGGATATT TCAGAGTCTA 4200 AAGTGGAAGA CTGTAAAATG GAATCTACCG AAACAGAAGA GAGAAGCACT GAGTTAAAAA 4260 CTGAAATAAA AGAGGAGGAA GACCAGCCAA GTACTTCAGC TACCCAGTCA TCTCCGGCTC 4320 CAGGACAGTC AAAGAAAAG ATTTTCAAAC CAGAAGAACT ACGACAGGCA CTGATGCCAA 4380 CATTGGAGGC ACTTTACCGT CAGGATCCAG AATCCCTTCC CTTTCGTCAA CCTGTGGACC 4440 CTCAGCTTTT AGGAATCCCT GATTACTTTG ATATTGTGAA GAGCCCCATG GATCTTTCTA 4500 CCATTAAGAG GAAGTTAGAC ACTGGACAGT ATCAGGAGCC CTGGCAGTAT GTCGATGATA
TTTGGCTTAT GTTCAATAAT GCCTGGTTAT ATAACCGGAA AACATCACGG GTATACAAAT 4560 4620 ACTGCTCCAA GCTCTCTGAG GTCTTTGAAC AAGAAATTGA CCCAGTGATG CAAAGCCTTG 4680 GATACTGTTG TGGCAGAAG TTGGAGTTCT CTCCACAGAC ACTGTGTTGC TACGGCAAAC AGTTGTGCAC AATACCTCGT GATGCCACTT ATTACAGTTA CCAGAACAGG TATCATTTCT GTGAGAAGTG TTTCAATGAG ATCCAAGGGG AGAGCGTTTC TTTGGGGGGAT GACCCTTCCC 4860 AGCCTCAAAC TACAATAAAT AAAGAACAAT TTTCCAAGAG AAAAAATGAC ACACTGGATC CTGAACTGTT TGTTGAATGT ACAGAGTGCG GAAGAAAGAT GCATCAGATC TGTGTCCTTC 4980 ACCATGAGAT CATCTGGCCT GCTGGATTCG TCTGTGATGG CTGTTTAAAG AAAAGTGCAC 5040 GAACTAGGAA AGAAAATAAG TTTTCTGCTA AAAGGTTGCC ATCTACCAGA CTTGGCACCT 5100 TTCTAGAGAA TCGTGTGAAT GACTTTCTGA GGCGACAGAA TCACCCTGAG TCAGGAGAGG 5160

91



TCACTGTTAG	AGTAGTTCAT	GCTTCTGACA	AAACCGTGGA	AGTAAAACCA	GGCATGAAAG	5220
CAAGGTTTGT	GGACAGTGGA	GAGATGGCAG	AATCCTTTCC	ATACCGAACC	AAAGCCCTCT	5280
TTGCCTTTGA	AGAAATTGAT	GGTGTTGACC	TGTGCTTCTT	TGGCATGCAT	GTTCAAGAGT	5340
ATGGCTCTGA	CTGCCCTCCA	CCCAACCAGA	GGAGAGTATA	CATATCTTAC	CTCGATAGTG	5400
	CCGTCCTAAA				CTAATTGGAT	5460
	TGTCAAGAAA					5520
GTGAGGGAGA						5580
	GGAATGGTAC		TTGACAAGGC	TGTATCAGAG		5640
	GGATATTTT				GCAAAGGAAT	5700
				AGAAAGCATT		5760
	AGAAGAGAGA					5820
						5880
CCAAGGGAGA				GAAAACCAGC		
	TAGGGGCAAC			CAATGTATCT		5940
	ATATGCCACC			CTTCTTTGTG		6000
		TCCCTGCCTC			CTCATCCCCT	6060
~ · · · · · · · · · · · · · · · · · · ·	GGATGGTCGG			AAGGGACAAG		6120
	CCGAAGAGCC			GCTGGTGGAG		6180
AGAGCCAGGA	CCGCTTTGTC					6240
GGCACTGTAC				CTGCTATAAC		6300
	AATGGAGAAA					6360
CTGCAGCCAC	CCAGAGCCCA	GGCGATTCTC	GCCGCCTGAG	TATCCAGCGC	TGCATCCAGT	6420
CTCTGGTCCA	TGCTTGCCAG	TGTCGGAATG	CCAATTGCTC	ACTGCCATCC	TGCCAGAAGA	6480
TGAAGCGGGT				AACCAATGGC	GGGTGCCCCA	6540
	GCTCATTGCC	CTCTGCTGCT	ACCATGCCAA	GCACTGCCAG	GAGAACAAAT	6600
	GTTCTGCCTA					6660
	GGCCCAAATG					6720
	ACAGGGCCTC					6780
	CACCCGCAG			GCCTCAGCCT		6840
	ACCCTACTTG					6900
	GGTGACCCCT					6960
	AGCAGTGGAA					7020
				CCAGATGCCC		7020
	CGTGCAAATT				•	7080
	CATGGGTATG					7200
	GGGACCGACA			CTGGAGCCAA		
	GCAACTACAG			CATGATGTCA		7260
	TTTGAACATG				ATCAGCCCAC	7320
	CACTGTGTCT		TACAAAACCT	TTTGCGGACT	CTCAGGTCTC	7380
	CCTGCAGCAG		TTAGTATCCT	TCACGCCAAC	CCCCAGCTGT	7440
	CATCAAGCAG					7500
	TGGCATGCCC			GCCACCTACC		7560
	CCACTCCAAT			TCCAATGCAG	GCGGGCGTTC	7620
AGAGGGCTGG	CCTGCCCCAG	CAGCAACCAC	AGCAGCAACT	CCAGCCACCC	ATGGGAGGGA	7680
TGAGCCCCCA	GGCTCAGCAG	ATGAACATGA	ACCACAACAC	CATGCCTTCA	CAATTCCGAG	7740
ACATCTTGAG	ACGACAGCAA	ATGATGCAAC	AGCAGCAGCA	ACAGGGAGCA	GGGCCAGGAA	7800
	AATGGCCAAC					7860
	GCAGCGGATG					7920
AGATAGGCCA	GCTTCCCCAG	GCCTTGGGAG	CAGAGGCAGG	TGCCAGTCTA	CAGGCCTATC	7980
AGCAGCGACT	CCTTCAGCAA	CAGATGGGGT	CCCCTGTTCA	GCCCAACCCC	ATGAGCCCCC	8040
AGCAGCATAT	GCTCCCAAAT	CAGGCCCAGT	CCCCACACCT	ACAAGGCCAG	CAGATCCCTA	8100
	CAATCAAGTG					8160
	CTCCAGTCCT					8220
	AAGTTCCCCA					8280
	TGCCAGCCCG					8340
	CCTCCATGGT					8400
	AAACCTCTCA					8460
					ATTTTTTGA	
	GCCTAAAAGA					8580
	AAGCAAACAT					8,640
	GGGCTGGTTA					
						8700
	GGAGGCTGAG					8760
	ATTATTTTT					8820
	TTATTATTA					8880
TTATGGAAGA	GTTAAAACAT	TTCTAAACCA	GAGGACAAAA	GGGGTTAATG	TTACTTTGAA	8940



ATTACATTCT ATATATAT AAATATAT AAATATAT TAAAATACCA GTTTTTTTC 9000 TCTGGGTGCA AAGATGTTCA TTCTTTTAAA AAATGTTTAA AAAAAA 9046

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7326 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

ATGGCCGAGA	ACTTGCTGGA	CGGACCGCCC	AACCCCAAAC	GAGCCAAACT	CAGCTCGCCC	60
GGCTTCTCCG	CGAATGACAA	CACAGATTTT	GGATCATTGT	TTGACTTGGA	AAATGACCTT	120
		TGGAGAATTA				180
		ACAACTGTCA				240
ATCAACCCAG	GGATAGGCAA	TGTGAGTGCC	AGCAGCCCTG	TGCAACAGGG	CCTTGGTGGC	300
CAGGCTCAGG	GGCAGCCGAA	CAGTACAAAC	ATGGCCAGCT	TAGGTGCCAT	GGGCAAGAGC	360
		ATCAACACCC				420
		CCAAGCACTG				480
GTGACCAGTA	GTCCTGCCAC	ATCACAGACT	GGACCTGGGA	TCTGCATGAA	TGCTAACTTC	540
		TCTCAATAGT				600
		CATGAATGGA				660
		TCCAGCCATG				720
ACCTTGACAC	AGGTTTCCCC	ACAAATGGCT	GGCCATGCTG	GACTAAATAC	AGCACAGGCA	780
GGAGGCATGA	CCAAGATGGG	AATGACTGGT	ACCACAAGTC	CATTTGGACA	ACCCTTTAGT	840
CAAACTGGAG	GGCAGCAGAT	GGGAGCCACT	GGAGTGAACC	CCCAGTTAGC	CAGCAAACAG	900
AGCATGGTCA	ATAGTTTACC	TGCTTTTCCT	ACAGATATCA	AGAATACTTC	AGTCACCACT	960
GTGCCAAATA	TGTCCCAGTT	GCAAACATCA	GTGGGAATTG	TACCCACACA	AGCAATTGCA	1020
ACAGGCCCCA	CAGCAGACCC	TGAAAAACGC	AAACTGATAC	AGCAGCAGCT	GGTTCTACTG	1080
CTTCATGCCC	ACAAATGTCA	GAGACGAGAG	CAAGCAAATG	GAGAGGTTCG	NGCCTGTTCT	1140
		GAAAAACGTT				1200
AAAGCCTGCC	AAGTTGCCCA	TTGTGCATCT	TCACGACAAA	TCATCTCTCA	TTGGAAGAAC	1260
		TGTTTGCCTC				1320
CAACAAACCA	TCCTGGGATC	TCCAGCTAGT	GGAATTCAAA	ACACAATTGG	TTCTGTTGGT	1380
		TTCCTTAAGT				1440
		AGGACTCCCC				1500
		ACCAGCACAG				1560
		CATGAGTGTC				1620
		ATCAGCTCTT			CAATCCACTG	1680
		TGGTAACATT			TACAGCAGCG	1740
CCTCCTTCCA	GCACTGGTGT	TCGAAAAGGC	TGGCATGAAC	ATGTGACTCA	GGACCTACGG	1800
		CGTTCAAGCC				1860
		CCTGGTTGCC				1920
		TGAATACTAT			СТАТААААТА	1980
		GCGGAGGACA			CCTGGGTAAC	2040
		TGGGGCTCAG			CCAGTCTGTA	2100
		GCCTTTGCCA			TCAAGGGATG	2160
AATTCATTTA	ACCCAATGTC	CCTGGGAAAC	GTCCAGTTGC	CACAGGCACC	CATGGGACCT	2220
		CCACTCTGTG				2280
		GATGCCTCAG				2340
		TACTCAGAAC				2400
		GAACAGTGTG				2460
		TGGAGCTGCT			GCTGGCACCC	2520
		CCCACCAGTG				2580
		CATGCCCTCT				2640
		CACTCAGCCA				2700
		GCCCAGCGCT			TACAGTCCAG	2760
		GACTCCACAG				2820
		GCAGCAACCA				2880
		CAGCATTGAT				2940
0000111010						25.10



						2000
AGTGCTGAAA	CCAGTTCCCA	GCAGCCAGGA	CCCGATGTGC	CCATGCTGGA	AATGAAGACA	. 3000
GAGGTGCAGA	CAGATGATGC	TGAGCCTGAA	CCTACTGAAT	CCAAGGGGGA	ACCTCGGTCT	3060
CACATGATGG	AAGAGGATTT	ACAAGGTTCT	TCCCAAGTAA.	AAGAAGAGAC	AGATACGACA	3120
GAGAIGAIGO	CAGAGCCAAT	CCNNCTNCNN	CNNNNGAAAC	CTCDAGTAAA	AGTGGAAGCT	3180
GAGCAGAAGI	CAGAGCCAAI	TEGER RECRE	DALACCERCA C	AATCAACATC	TCCTTCCCAC	3240
AAAGAGGAAG	AAGAGAACAG	TTCGAACGAC	ACAGCCTCAC	AATCAACATC	TCCTTCCCAG	
CCACGCAAAA	AAATCTTTAA	ACCCGAGGAG	CTACGCCAGG	CACTTATGCC	AACTCTAGAA	3300
GCACTCTATC	GACAGGACCC	AGAGTCTTTG	CCTTTTCGTC	AGCCTGTAGA	TCCTCAGCTC	3360
CTACCAATCC	CAGATTATTT	ТСАТАТАСТС	AAGAATCCTA	TGGACCTTTC	TACCATCAAA	3420
CIAGOAAICC	ACACAGGGCA	מייייייייייייייייייייייייייייייייייייי	CCCTCCCAGT	ATGTGGATGA	$TGTC\DeltaGGCTT$	3480
CGAAAGCTGG	ACACAGGGCA	ATATCAAGAA	CCCIGGCAGI	COCCONTON	A TOTOROCCI CT	3540
ATGTTCAACA	ATGCGTGGCT	ATATAATCGT	AAAACGTCCC	GIGIAIAIAA	ATTTTGCAGT	
AAACTTGCAG	AGGTCTTTGA	ACAAGAAATT	GACCCTGTCA	TGCAGTCTCT	TGGATATTGC	3600
TGTGGACGAA	AGTATGAGTT	CTCCCCACAG	ACTTTGTGCT	GTTACGGAAA	GCAGCTGTGT	3660
ACAATTCCTC	GTGATGCAGC	CTACTACAGC	TATCAGAATA	GGTATCATTT	CTGTGGGAAG	3720
MCMMTCTCTC	AGATCCAGGG	CCACAATCTC	ACCCTGGGTG	ACGACCCTTC	CCAACCTCAG	3780
TGTTTCACAG	AGAICCAGGG	DEMORATORO	ACCOLOGIC	AMACCAMACA	TCCTCDACCT	3840
ACGACAATTT	CCAAGGATCA	ATTTGAAAAG	AAGAAAAIG	ATACCTTAGA	D C D C C D C C C C C C C C C C C C C C	
TTTGTTGACT	GCAAAGAGTG	TGGCCGGAAG	ATGCATCAGA	TTTGTGTTCT	ACACTATGAC	3900
ATCATTTGGC	CTTCAGGTTT	TGTGTGTGAC	AACTGTTTGA	AGAAAACTGG	CAGACCTCGG	3960
AAAGAAAACA	AATTCAGTGC	TAAGAGGCTG	CAGACCACAC	GATTGGGAAA	CCACTTAGAA	4020
CACACACTCA	ATAAGTTTTT	GCGGCGCCAG	AATCACCCTG	AAGCTGGGGA	GGTTTTTGTC	4080
GACAGAGIGA	CCAGCTCAGA	CARCACTCTC	CACCECAACC	CCCCAATCAA	CTCAAGGTTT	4140
AGAGTGGTGG	CCAGCTCAGA	CAAGACIGIG	GAGGICAAGC	COUCATIOAA	CHUMCCHUM	4200
GTGGATTCTG	GAGAGATGTC	GGAATCTTTC	CCATATCGTA	CCAAAGCACT	CTTTGCTTTT	
GAGGAGATCG	ATGGAGTCGA	TGTGTGCTTT	TTTGGGATGC	ATGTGCAAGA	TACGGCTCTG	4260
ATTGCCCCCC	ACCAAATACA	AGGCTGTGTA	TACATATCTT	ATCTGGACAG	TATTCATTTC	4320
MECCECCCC	GCTGCCTCCG	GACAGCTGTT	TACCATGAGA	TCCTCATCGG	ATATCTCGAG	4380
7100000000	AATTGGTGTA	TCTCTCTCT	CATATTTCCC	CCTGTCCCCC	AAGTGAAGGA	4440
TATGTGAAGA	AATTGGTGTA	1G1GACAGCA	CAIMITIGG	TCCCCD D D CC	AAAAACCACTA	4500
GATGACTATA	TCTTTCATTG	CCACCCCCT	GACCAGAAAA	TCCCCAAACC	AAAACGACIA	
CAGGAGTGGT	ACAAGAAGAT	GCTGGACAAG	GCGTTTGCAG	AGAGGATCAT	TAACGACTAT	4560
AAGGACATCT	TCAAACAAGC	GAACGAAGAC	AGGCTCACGA	GTGCCAAGGA	GTTGCCCTAT	4620
TTTGAAGGAG	ATTTCTGGCC	TAATGTGTTG	GAAGAAAGCA	TTAAGGAACT	AGAACAAGAA	4680
	GGAAAAAAGA	ACACACTACT	GCAGCGAGTG	AGACTCCTGA.	GGGCAGTCAG	4740
GAAGAAGAAA	AAAATGCGAA	CDDDDDDCDDC	DACABCARCA.	CCAACAAAAA	CANANGCAGC	4800
GGTGACAGCA	AAAATGCGAA	GAAAAAGAAC	AACAAGAAGA		CAAAAGCAGC	
ATTAGCCGCG	CCAACAAGAA	GAAGCCCAGC	ATGCCCAATG	TTTCCAACGA	CCTGTCGCAG	· ~4860
AAGCTGTATG	CCACCATGGA	GAAGCACAAG	GAGGTATTCT	TTGTGATTCA	TCTGCATGCT	4920
GGGCCTGTTA	TCAGCACTCA	GCCCCCCATC	GTGGACCCTG	ATCCTCTGCT	TAGCTGTGAC	4980
CTCATGGATG	GGCGAGATGC	CTTCCTCACC	CTGGCCAGAG	ACAAGCACTG	GGAATTCTCT	5040
CICAIGGAIG	GCTCCAAATG	CTCCACTCTC	TECATECTES	TGCAGCTGCA	CACACAGGG	5100
TCCTTACGCC	GCTCCAAATG	GICCACICIG	TGCAIGCIGG	DECECTOR	D.C.C.C.C.C.C.C.C.C.C.C.C.C.C.C.C.C.C.C	5160
CAGGACCGCT	TTGTTTATAC	CTGCAATGAG	TGCAAACACC	ATGTGGAAAC	ACGCTGGCAC	
TGCACTGTGT	GTGAGGACTA	TGACCTTTGT	ATCAATTGCT	ACAACACAAA	GAGCCACACC	5220
CATAAGATGG	TGAAGTGGGG	GCTAGGCCTA	GATGATGAGG	GCAGCAGTCA	GGGTGAGCCA	5280
CAGTCCAAGA	GCCCCAGGA	ATCCCGGCGT	CTCAGCATCC	AGCGCTGCAT	CCAGTCCCTG	5340
CTCCATCCCT	GCCAGTGTCG	CAATGCCAAC	TGCTCACTGC	CGTCTTGCCA	GAAGATGAAG	5400
GIGCAIGCCI	AGCACACCAA	CCCCTCCAAC	CCCANGACTA	ATGGAGGATG	CCCAGTGTGC	5460
CGAGTCGTGC	AGCACACCAA	GGGCTGCAAG	COCAAGACIA	CCCDDCDDDD	mn n nmccccm	5520
AAGCAGCTCA	TTGCTCTTTG	CTGCTACCAC	GCCAAACACT	GCCAAGAAAA	TAAATGCCCT	
GTGCCCTTCT	GCCTCAACAT	CAAACATAAC	GTCCGCCAGC	AGCAGATCCA	GCACTGCCTG	5580
CAGCAGGCTC	AGCTCATGCG	CCGGCGAATG	GCAACCATGA	ACACCCGCAA	TGTGCCTCAG	5640
CAGAGTTTGC	CTTCTCCTAC	CTCAGCACCA	CCCGGGACTC	CTACACAGCA	GCCCAGCACA	5700
CCCCDDDCDC	CACAGCCCCC	AGCCCAGCCT	CAGCCTTCAC	CTGTTAACAT	GTCACCAGCA	5760
CCCCAAACAC	ATGTAGCCCG	CACTCACCCC	CCAACAATAG	тстстсстсс	GAAGCCTACC	5820
GGCTTCCCTA	AIGIAGCCCG	GACTCAGCCC	CAACAATAO	CECCACCACE	A CA A CCA CCC	5880
AACCAGGTGC	CAGCTCCCCC	ACCCCCTGCC	CAGCCCCCAC	CTGCAGCAGT	AGAAGCAGCC	
CGGCAAATTG	AACGTGAGGC	CCAGCAGCAG	CAGCACCTAT	ACCGAGCAAA	CATCAACAAT	5940
GGCATGCCCC	CAGGACGTGA	CGGTATGGGG	ACCCCAGGAA	ĞCCAAATGAC	TCCTGTGGGC	6000
CTGAATGTGC	CCCGTCCCAA	CCAAGTCAGT	GGGCCTGTCA	TGTCTAGTAT	GCCACCTGGG	6060
CACTCCCACC	AGGCACCCAT	CCCTCAGCAG	CAGCCGATGC	CAGGCATGCC	CAGGCCTGTA	6120
CAGIGGCAGC	AGGCACCCA1	DASORS CREECE	CCCCCACCCA	méccen n mem	CCDCCCDDDC	6180
ATGTCCATGC	AGGCCCAGGC	AGCAGTGGCT	amages agas	CCCCAAIGI	N C C C N C C C C C	6240
AGGAGCATCT	CGCCAAGTGC	CCTGCAAGAC	· CTGCTACGGA	CCCTAAAGTC	ACCCAGCTCT	
CCTCAGCAGC	AGCAGCAGGT	GCTGAACATC	CTTAAATCAA	ACCCACAGCT	AATGGCAGCT	6300
TTCATCAAAC	AGCGCACAGC	CAAGTATGTG	GCCAATCAGC	CTGGCATGCA	GCCCCAGCCC	6360
GCDCTTCDNT	CCCAGCCTGG	TATGCAGCCC	CAGCCTGGCA	TGCACCAGCA	GCCTAGTTTG	6420
CANANCCMCA	. ACGCAATGCA	ACCTGGTGTG	. CCACGGCCTG	GTGTGCCTCC	ACCACAACCA	6480
CAAAACCTGA	ACGUAAIGCA	CCRCCCCCC	CCMCMCAACT	mchmchhcccc	ACCACACTOR	6540
GCAATGGGAG	GCCTGAATCC	CCAGGGACAA	GCTCTGAACA	TCATGAACCC	AGGACACAAC	
CCCAACATGA	CAAACATGAA	. TCCACAGTAC	CGAGAAATGG	TGAGGAGACA	GCTGCTACAG	6600
CACCAGCAGC	AGCAGCAGCA	. ACAGCAGCAG	CAGCAGCAGC	AACAACAAAA	TAGTGCCAGC	6660
TTGGCCGGGG	GCATGGCGGG	ACACAGCCAG	TTCCAGCAGC	CACAAGGACC	TGGAGGTTAT	6720



GCCCCAGCCA'	TGCAGCAGCA	ACGCATGCAA	CAGCACCTCC	CCATCCAGGG	CAGCTCCATG	6780
GGCCAGATGG	CTGCTCCAAT	GGGACAACTT	GGCCAGATGG	GGCAGCCTGG	GCTAGGGGCA	6840
GACAGCACCC	CTAATATCCA	GCAGGCCCTG	CAGCAACGGA	TTCTGCAGCA	GCAGCAGATG	6900
AAGCAACAAA	TTGGGTCACC	AGGCCAGCCG	AACCCCATGA	GCCCCCAGCA	GCACATGCTC	6960
TCAGGACAGC	CACAGGCCTC	ACATCTCCCT	GGCCAGCAGA	TCGCCACATC	CCTTAGTAAC	7020
CAGGTGCGAT	CTCCAGCCCC	TGTGCAGTCT	CCACGGCCCC	AATCCCAACC	TCCACATTCC	7080
AGCCCGTCAC	CACGGATACA	ACCCCAGCCT-	TCACCACACC	ATGTTTCACC	CCAGACTGGA	7140
ACCCCTCACC	CTGGACTCGC	AGTCACCATG	GCCAGCTCCA	TGGATCAGGG	ACACCTGGGG	7200
AACCCTGAAC	AGAGTGCAAT	GCTCCCCCAG	CTGAATACCC	CCAACAGGAG	CGCACTGTCC	7260
AGTGAACTGT	CCCTGGTTGG	TGATACCACG	GGAGACACAC	TAGAAAAGTT	TGTGGAGGGT	7320
TTGTAG						7326

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2499 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single .
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TCACTTGTCA ATTAATCCAG CTTCCTTAAT TTTACTGAAG AAGAATTTCT CCAGGATATT GGCACATTTG TAGTATTCAC TCTCAGGGGC GTTGTACTCT TTGCAATTGG TAAAGACTCG 120 CTGTAAGTCT GCCATGAATA ATTTCTTAGA CACGTAGTAC CTATTCTTGA GGCGTTCACT 180 CATGGTTTTG AGATCCATGG GGGACCTTAT AACTTCATAA TATCCTGGAG CTTCTGTTCT CTTCACAGGT TCCATGAAGG GCCAAGCGCT TTGATGGCTC TTCACCTGCT GGAGGATGCT 300 CTTGAGCGTG CTGTAAAGCT GGTCAGGGTC TCTGGGCTCT TTACTTTTCT CTTTTCCACT 360 CGGTTTCCAG CCTGTCTCTC TAATTCCAGG AATGCTTTCT ATAGGAATCT GTCGAACTCC 420 ATCTTTAAAA CATGAAAGTC CAGGGTAAAC TTTTCGAATT TGTGCCTGTT TTCTTTCAAT CAGTTTTTTA ATTATCTCCT TCTGCTTTTT AATGATGACA GAAAATTCTG TGTACGGGAT 540 CCGTGGATTT AGCTCACATC CCATTAAAGT GGCTCCTTCA TAATCCTTGA TATAGCCAAC 600 ATATTTGGTT TTAGGTATTT TAATTTCTTT GGAGAAACCC TGTTTCTTAA AGTATCCAAT TGCATATCA TCTGCATATG TGAGGAAGTT CAGGATGTCA TGCTTTATGT GATATTCTTT 660 720 CAAATGATTC ATCAGGTGTG TTCCATAGCC CTTGACTTGC TCATTTGAGG TTACAGCACA GAAGACAATC TCTGTGAATC CTTGAGATGG GAACATACGG AAACAGATAC CACCAATAAC ACGGCCATCT TTAATTAAAG CAAGGGTTTT GTGTTTCGGG TCAAAGACGA GCCGTGTGAT 900 GTATTCTTTT GGCATTCGGG GCAGCTGGTG GGAGAAAACG TTCTGTAGGC CAACCAGCCA 960 CATCAGGATC TTCTTGTTTG GTTTCTGGTT GAGGGAATTG CCAACCACGT GAAATTCAAT 1020 TACACCCCTG CGCTCTTCCA ACCTTGCCGC CTCATCCCTG GCCGAGTGTG CTGACAGAAA 1080 ATTGGTCTCT GGTCCAAGCA TTGCTGCAGG GTCCGTGATG GTAGACATAA CCTCGTTGAT 1140 TAATTCCATC GGAATATCCC CCATAACTCG GGGTTTCTTG GCCTCCTCCA GAACATGAGA 1200 ATCAGTCATT TTCCTCTTTT CTCCTGGGTT TGCCTCAAGT CCAGAAGAGG CTTTGCAGGC 1260 AGGACTGCTG CTCCCTGCGT TTGGCTGCTC AAGGGAAGAT GAGGTTGAAT TGTATGAAAT 1320 TGTCCCAGCC ACAGGAGGTG GATTGATAAC TGTTTGGATG CCTAGCTGGC TGGTTCTGGA 1380 AGAGGCTGAG AGAAAATCCT GATCCCAGAT GGGAGAGTTT TGACTATATA CTTCTTCTC 1440 TAGCATGGAC AGAAATTTTG GGAAATGAGT GAGGATTAGA GTTCGTTTTT CAAGAGGCAG 1500 TTTATCTTTT TCCTGTCTTG CTTGTTCCAG GAGTTGTCGC CTCATAACAG TGAAGACCGA 1560 GCGAAGCAAT GTTCTCCCAA ACACCTGTGT GGTTTCGTAC CGAGGTAGAC TGTCGCAGAA CTGTGGCACG TTGCAGTAAC ACAGCCACCT TGTGTAGTTC TCTTTGTATC CAGAAATATC 1680 ATCATTGGGA GATCGCAGTC TTCGTTGAGA TGGTGCCTCC AGATGCCAAT AGTTGATGCG 1740 GTTTAGGAAC ATTTTTGCCA ACTCAACTAT TGTTTGCCTT TCTTTTGCTG GCAGGTGACT 1800 AAATTTGTAC TGCACAAAGT TATTCACACC CTGTTCAATG CTAGGTTTTT CAAATGGGGG 1860 TTTCTTTCC AAAGAGCCTT CAACCACAGG TTTTCCTCTT TGTAAAATAG ACTTTCTCAA 1920 GAGCTTAAAT AGATAGAAAT AAACTTGTTT GGTATCTGCA TCTTCTTCCT TGTGGACACA 1980 GGTAAAGAGA TATTCCACAT CCAATACTAT TCCCAGGAGT CTGTTCATTT CTTCCTCTGA CACATTCTCC AGGTGGGAAA CATGAGCAGC TAGGGCATGG CTACAACTCC GACAGGATTC 2100 TGTTAGACTG ACAATTATTT GCTGCAGGTC GGCTCTGGGG GGAGTGGGTG AGGGGTTAGG 2160 GTTTTTCCAG CCATTACATT TACAAGACTC CTCGGCCTTG CAGGCGGAGT ACACTCCGAG 2220 TTTCTCCAGT TTCTTGGCCC GCGGAGCGGA GCGTAGTTGC GCTTTCTTCA CGGCGATTCG 2280 GGCCGAGCCA CCGCCTCCCG GTCCTTCGGC CGTGCCCGCT GCAGCCACTG CCGTCGCCGG 2340 ACCGCAGGCG CCCGAGCCCC CGGCGGCAGC GGCGCAGGGG GAGCCCTGCG GGGGCGCGGG 2400

CGGAAGCGCC GCAGGCTGCG GGGGCAGCGC CCCGGGCCCG GCCCCTGCCC CGGCTCCTGC 2460 CCCGCAGCCG CCCGCCAGC CTCGGACAT 2499

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2442 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

WO 98/03652

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TCACTTGTCA	ATCAACCCTG	CTTCCTTAAT	TTTACTGAAG	AAGAACTTCT	CCAGGATGCT	60
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CATGGTTTTC	AGATCCATGG	GGAACCTTAT	AACTTCATAA	TATCCCGGAG	CTTCTGTTCT	240
CTTCACTGGT	TCCATGAAAG	GCCAAGCATT	TGGATGGTTC	TTCACCTGCT	GCAGGATGTT	300
CTTGAGGGTG	CTGTAAACGT	GCTCAGGGTC	TTTGGGCTCT	TTACTTTTCT	CTTTTCCACT	360
TGGTTTCCAG	CCTGTCTCTC	TGATTCCAGG	AATGCTTTCT	ATAGGAATCT	GCCGAACTCC	420
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ATATTTGGTT	TTAGGTATTT	TGATTTCTTT	GGAGAAACCC	TGCTTCTTGA	AATAGCCGAT	660
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ACGGCCATCT	TTAATTAAAG	CAAGGGTTTT	GTGTTTCGGG	TCAAAGACGA	GCCGTGTGAT	900
GTACTCTTTG	GGCATTCTGG	GCAGCTGGTG	GGAAAACACA		CCACGAGCCA	960
CATCAGGATC	TTCTTGTTTG	GTTTCTGGTT	CAGGGAGTTG	CCCACCACGT	GGAATTCAAT	1020
GACACCCCTG	CGTTCTTCCA	GCCGTGCCGC	CTCATCTCTG	GCCGAATGGG	CTGACAGAAA	1080
ATTGGTCTCT	GGTCCAAGCA	TCCCTGCAGG	GTCTGTGATG	GTAGACATGA	CCTCATTGAT	1140
CAATTCCACG	GGAATATCCC	CCATCACTCG	AGATCTCTTG	GCCTCCTCGG	GAGCATGAGA	1200
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AGGACTTGTT	CTCCCTCCAT	TGATCTGCTC	ATGGGAAGTT	GAATTTGAAC	TGAACAATGC	1320
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(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

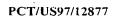


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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16: CCCGCCAGCC TCGGACATGC 20 (2) INFORMATION FOR SEQ ID NO:17: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17: CCCGCCAGCC TCGGCCATGC 20 (2) INFORMATION FOR SEQ ID NO:18: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2442 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

ATGGCCGAGG CTGGCGGGGC CGGGTCCCCA GCACTGCCTC CCGCGCCCCC, GCACGGTTCC. CCCCGGACCC TGCCCACCGC TGCCGGGAGC TCTGCTTCCT GCGGGCCAGC GACGCCGGTG 120 GCCGCGGCGG GCACCGCCGA GGGACCGGGA GGAGGCGGCT CGCCCGGAT CGCCGTGAAG 180 AAGGCGCAGT TGCGCTCTGC TCCGCGGGCG AAGAAGCTGG AGAAACTCGG CGTGTACTCC 240 GCCTGCAAGG CAGAGGAGTC CTGTAAATGC AATGGCTGGA AGAACCCTAA CCCCTCTCCT 300 ACTCCACCAA GAGGAGACCT CCAGCAGATA ATTGTCAGTT TGACAGAATC CTGTCGAAGC TGTAGCCATG CCCTTGCTGC TCACGTTTCT CACTTGGAGA ATGTGTCAGA GGAAGAGATG 360 GACAGACTCC TGGGAATTGT GTTGGATGTG GAGTACCTCT TCACCTGCGT CCACAAAGAA 480 GAAGATGCAG ATACCAAACA AGTGTACTTC TACCTATTCA AGCTCTTGAG AAAGTCAATT 540 TTACAAAGAG GAAAACCTGT GGTTGAAGGC TCCTTGGAGA AGAAGCCGCC ATTTGAGAAG 600 CCCAGTATTG AACAGGGTGT GAACAACTTC GTGCAGTACA AGTTTAGTCA CTTGCCATCG 660 AAAGAGAGGC AGACAACGAT CGAGCTGGCC AAGATGTTTC TGAACCGCAT CAACTACTGG CATCTGGAGG CTCCATCTCA GCGGAGACTA CGGTCTCCCA ATGATGACAT CTCTGGATAC 780 AAGGAAACT ACACAAGGTG GTTGTGCTAC TGCAATGTAC CGCAGTTCTG TGACAGCTTA 840 CCTCGGTACG AAACCACAAA GGTGTTTGGG AGAACATTGC TTCGCTCGGT CTTCACCATC 900 ATGAGACGAC AGCTCTTGGA ACAAGCCAGA CAGAAAAAAG ACAAACTGCC TCTTGAGAAA CGCACGCTTA TCCTCACACA TTTCCCAAAG TTTCTGTCCA TGTTGGAAGA AGAAGTGTAT AGTCAAAATT CTCCTATCTG GGATCAGGAT TTTCTCTCAG CCTCTTCCAG AACCAGCCCG 1080 CTAGGAATCC AAACAGTAAT CAGTCCTCCT GTTACTGGGA CAGCATTGTT CAGTTCAAAT 1140 TCAACTTCCC ATGAGCAGAT CAATGGAGGG AGAACAAGTC CTGGATGCAG AGGCTCTTCT 1200 GGGCTTGAAG CAAACCCGGG AGAAAAGAGG AAAATGAACA ACTCTCATGC TCCCGAGGAG 1260 GCCAAGAGAT CTCGAGTGAT GGGGGATATT CCCGTGGAAT TGATCAATGA GGTCATGTCT 1320 ACCATCACAG ACCCTGCAGG GATGCTTGGA CCAGAGACCA ATTTTCTGTC AGCCCATTCG 1380 GCCAGAGATG AGGCGGCACG GCTGGAAGAA CGCAGGGGTG TCATTGAATT CCACGTGGTG 1440 GGCAACTCCC TGAACCAGAA ACCAAACAAG AAGATCCTGA TGTGGCTCGT GGGCCTCCAG 1500 AATGTGTTTT CCCACCAGCT GCCCAGAATG CCCAAAGAGT ACATCACACG GCTCGTCTTT 1560
GACCCGAAAC ACAAAACCCT TGCTTTAATT AAAGATGGCC GTGTCATTGG TGGTATCTGT 1620 TTCCGGATGT TTCCATCCCA GGGATTCACA GAGATTGTTT TCTGTGCAGT AACCTCAAAT GAACAAGTCA AGGGCTATGG AACCCACCTG ATGAACCATC TCAAAGAATA CCACATAAAG 1740 CACGAGATCC TCAACTTCCT CACATATGCA GATGAGTATG CCATCGGCTA TTTCAAGAAG 1800 CAGGGTTTCT CCAAAGAAT CAAAATACCT AAAACCAAAT ATGTTGGCTA CATCAAGGAT 1860 TATGAAGGGG CCACTTTGAT GGGATGTGAG CTGAACCCTC AGATCCCATA CACAGAGTTC 1920 TCTGTCATCA TTAAAAAGCA GAAGGAGATC ATTAAAAAGC TGATAGAAAG AAAACAAGCC 1980 CAGATTCGAA AAGTCTACCC TGGACTTTCG TGTTTCAAAG ATGGAGTTCG GCAGATTCCT 2040







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AAGAACCATC	CAAATGCTTG	GCCTTTCATG	GAACCAGTGA	AGAGAACAGA	AGCTCCGGGA	2220
TATTATGAAG	TTATAAGGTT	CCCCATGGAT	CTGAAAACCA	TGAGTGAACG	CCTCAGGAAC	2280
AGGTACTATG	TGTCTAAGAA	GTTATTCATG	GCGGACTTGC	AACGAGTGTT	CACCAACTGC	2340
AAGGAGTACA	ACCCTCCCGA	GAGCGAGTAC	TACAAATGCG	CCAGCATCCT	GGAGAAGTTC	2400
TTCTTCAGTA	AAATTAAGGA	AGCAGGGTTG	ATTGACAAGT	GA		2442



What is claimed is:

- 1. A purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones.
- 2. The protein of claim 1 consisting of the amino acid sequence of SEQ ID NO:1.
- 3. The protein of claim 1 comprising the amino acid sequence of SEQ ID NO:2.
- 4. The protein of claim 1, which also binds to the amino acid sequence of SEQ ID NO:3 on a p300 cellular protein and to amino acid residues 1805-1854 of a CBP cellular protein (SEQ ID NO:9).
- 5. A fragment of the protein of claim 1 having histone acetyltransferase activity.
- 6. A polypeptide consisting of the amino acid sequence of SEQ ID NO: 2.
- 7. A fragment of the protein of claim 1 which binds to the amino acid sequence of SEQ ID NO: 3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- 8. A polypeptide consisting of the amino acid sequence of SEQ ID NO:4.
- 9. A nucleic acid consisting of the nucleotide sequence of SEQ ID NO:10.
- 10. A nucleic acid having a nucleotide sequence which encodes the protein of claim
- 1.



- 11 A nucleic acid having a nucleotide sequence which encodes the protein of claim 2.
- 12 A nucleic acid having a nucleotide sequence which encodes the protein of claim
 3.
- 13. A nucleic acid consisting of the nucleotide sequence which encodes the protein of claim 4.
- 14 A nucleic acid complementary to and which selectively hybridizes with the nucleic acid of claim 11 under stringent hybridization conditions.
- 15. A fragment of the nucleic acid of claim 9, which encodes a polypeptide that acetylates histones.
- A fragment of the nucleic acid of claim 9, which encodes a polypeptide which binds to the amino acid sequence of SEQ ID NO:3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- 17. A purified antibody which specifically binds the protein of claim 1.
- 18. A purified antibody which specifically binds the protein of claim 2.
- 19. A purified antibody which specifically binds the protein of claim 3.
- 20. A purified antibody which specifically binds the protein of claim 4.
- An assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of P/CAF comprising:
- a) contacting the substance with a system in which histone acetylation by P/CAF can be determined;



- b) determining the amount of histone acetylation by P/CAF in the presence of the substance, and
- c) comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.
- 22. An assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising:
- a) contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined;
- b) determining the amount of P/CAF binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP.
 - The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the p300 protein comprising amino acid residues 1767-1816 (SEQ ID NO:3) and the protein of claim 4.
 - The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising amino acid residues 1805-1854 (SEQ ID NO.9) and the protein of claim 4.
 - The method of claim 22, wherein the system consists of a cell extract produced from cells producing both p300 and P/CAF

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- An assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP comprising:
- a) contacting the substance with a system in which histone acetylation by p300/CBP can be determined;
- b) determining the amount of histone acetylation by p300/CBP in the presence of the substance; and
- c) comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP.
 - 27. An assay for screening substances for the ability to inhibit binding of a DNA-binding transcription factor to p300/CBP comprising:
 - a) contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined,
 - b) determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.
 - The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a DNA-binding transcription factor and p300/CBP.
 - 29. The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising a DNA-binding transcription factor and p300/CBP.



- The method of claim 27, wherein the system consists of a cell extract produced from cells producing both a DNA-binding transcription factor and p300/CBP.
- The method of claim 27, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1
- 32. A method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.
- 33. The method of claim 32, wherein the substance can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP.
- A method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier.
- 35. The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by promoting the binding of P/CAF to p300/CBP.
- 36. The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by stimulating the histone acetlytransferase activity of P/CAF.
- 37. A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

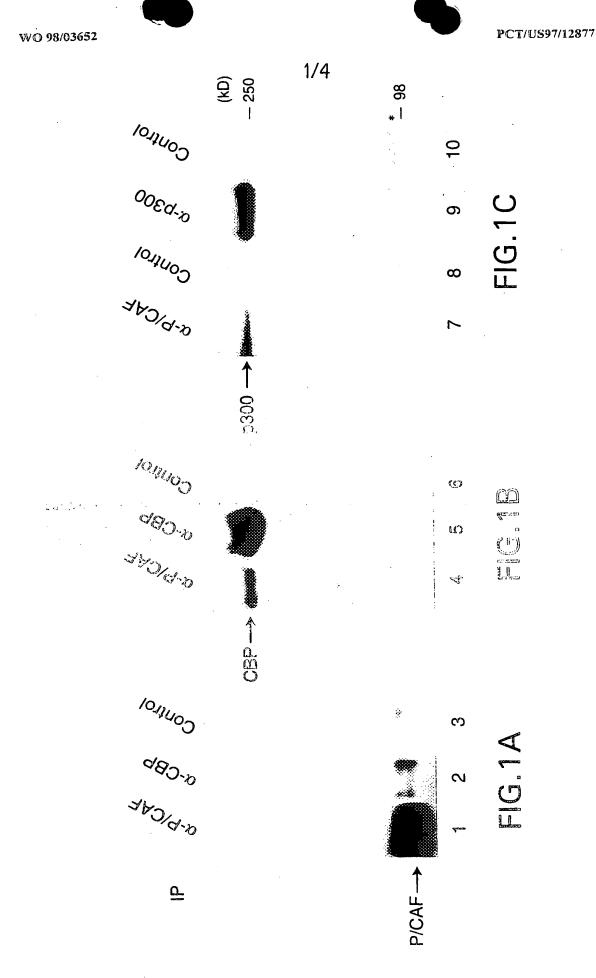




The method of claim 37, wherein the substance can inhibit the transcription modulating activity of p300/CBP by preventing the binding of a DNA-binding transcription factor to p300/CBP.

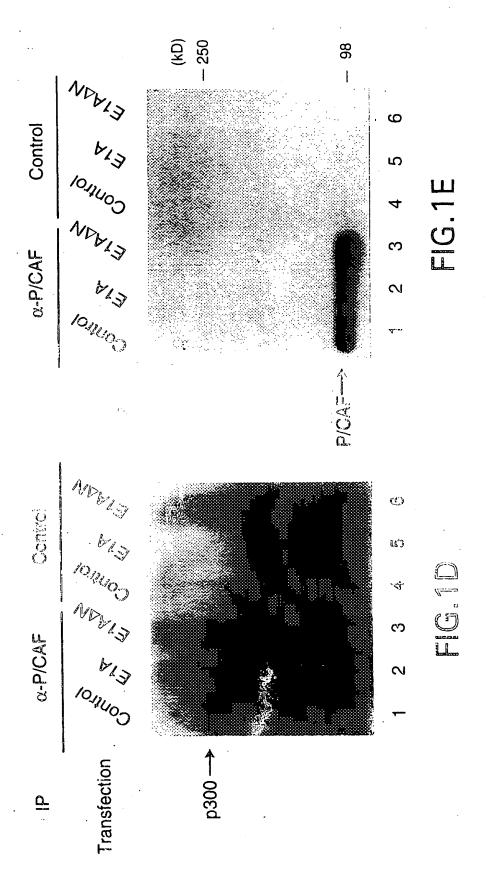
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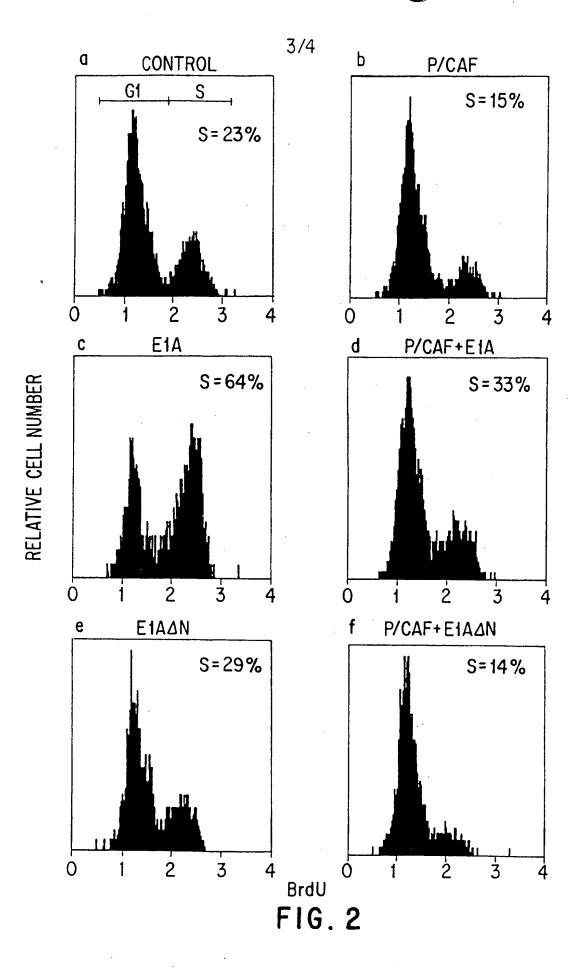
- The method of claim 38, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1
- The method of claim 37, wherein the substance is an antibody which binds p300/CBP.
- A method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.
- The method of claim 41, wherein the substance can stimulate the histone acetyltransferase activity of p300/CBP by promoting the binding of a DNA-binding transcription factor to p300/CBP.



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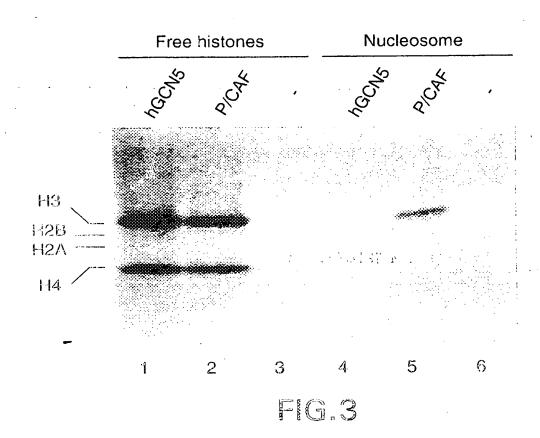
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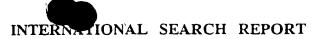






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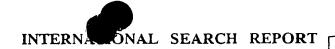


Inter .nal Application No PCT/US 97/12877

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A. CLASSIF	FICATION OF SUBJECT MATTER C12N15/12 C07K14/47 G01N33/	50 A61K38/17			
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According to	International Patent Classification (IPC) or to both national classific	ation and IPC			
B. FIELDS					
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Documentat	ion searched other than minimum documentation to the extent that s	uch documents are included in the helds search			
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	ENTS CONSIDERED TO BE RELEVANT		Dalaman and Att		
Category °	Citation of document, with indication, where appropriate, of the rel	evant passages	Relevant to claim No.		
A	EMPL EST Aggression numberN20522	•	1		
Α	EMBL EST, Accession numberN39522 Sequence no yv27b08.sl Homo sapi		-		
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	25 January 1996				
	XP002050402 see the whole document				
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	EMBO JOURNAL.,	60			
	vol. 11, 1992, EYNSHAM, 0XFORD pages 4145-4152, XP002050399	GB,			
	see the whole document				
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X Furt	her documents are listed in the continuation of box C.	Patent family members are listed in	annex.		
° Special ca	ategories of cited documents :	"T" later document published after the interna	ational filing date		
	ent defining the general state of the art which is not	or priority date and not in conflict with the cited to understand the principle or theorem.	e application but		
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1	17 December 1997	1 4. 01. 30			
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	European Patent Office, P.B. 5818 Patentiaan 2				
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Form PCT/ISA/210 (second sheet) (July 1992)

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Inters nal Application No PCT/US 97/12877

•) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category Cit.	ation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	YANG, X.Y. ET AL.: "A p300-CBP-associated factor that competes with the adenoviral oncoprotein E1A" NATURE., vol. 382, no. 8589, 25 July 1996, LONDON GB, pages 319-324, XP002050400 see the whole document	1
P,X	OGRYZKO, V.V. ET AL.: "The transcriptional coactivators p300 and CBP are histone acetyltransferases" CELL, vol. 87, no. 5, November 1996, NA US, pages 953-959, XP002050401 see the whole document	1
P,X	EMBL EST, Accession number U57316, Sequence reference human GCN5 (hGCN5) complete cds. 26 august 1996 XP002050403 see the whole document	



INTERNATIONAL SEARCH REPORT

International application No. PCT/US 97/12877

Box	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Int	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: see FURTHER INFORMATION sheet PCT/ISA/210
2.	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This In	ternational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Rema	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.



FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Remark: Although claims 32 to 42 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the composition.